

DICHLOROBENZIDINE

Ambient Water Quality Criteria

Criteria and Standards Division  
Office of Water Planning and Standards  
U.S. Environmental Protection Agency  
Washington, D.C.

### 3,3'-DICHLOROBENZIDINE (DCB)

#### CRITERIA SUMMARY

##### Freshwater Aquatic Life

For freshwater aquatic life, no criterion for any dichlorobenzidine can be derived using the Guidelines, and there are insufficient data to estimate a criterion using other procedures.

##### Saltwater Aquatic Life

For saltwater aquatic life, no criterion for any dichlorobenzidine can be derived using the Guidelines, and there are insufficient data to estimate a criterion using other procedures.

##### Human Health

For the maximum protection of human health from the potential carcinogenic effects of exposure to dichlorobenzidine through ingestion of water and contaminated aquatic organisms, the ambient water concentration is zero. Concentrations of  $10^{-5}$  to  $10^{-7}$   $\mu\text{g/l}$  are estimated to result in additional lifetime cancer risks ranging from no additional risk to an additional risk of 1 in 100,000 are presented in the Criterion Formulation section of this document. The Agency is considering setting criteria at an interim target risk level in the range of  $10^{-5}$ ,  $10^{-6}$ , or  $10^{-7}$  with corresponding criteria of  $1.69 \times 10^{-2}$   $\mu\text{g/l}$ ,  $1.69 \times 10^{-3}$   $\mu\text{g/l}$  and  $1.69 \times 10^{-4}$   $\mu\text{g/l}$  respectively.

## DICHLOROBENZIDINE

### Introduction

Dichlorobenzidine (4,4'-diamino-3,3'-dichlorobiphenyl) (DCB) is used in the production of dyes and pigments and as a curing agent for polyurethanes. This compound is soluble in organic solvents, but it is nearly insoluble in water (Stecher, 1968).

The affinity of DCB for suspended particulates in water is not clear; its basic nature suggests that it may be fairly tightly bound to humic materials in soils (Radding, et al. 1975). Soils may be moderate to long term reservoirs.

DCB has been shown to be a carcinogen in non-human mammals under controlled laboratory conditions. Exposure to DCB results in various types of sarcomas and adenocarcinomas. Tumors have been induced both locally (at the site of injection) and remotely (multi-system involvement after feeding) (Rye, et al. 1970). Experiments show DCB to be a much less potent carcinogen in animals than the unsubstituted base (benzidine) (Rye, et al. 1970).

No definitive evidence exists to demonstrate DCB as a carcinogen in man. Unlike benzidine, DCB exposure to humans produced no increases in urinary bladder carcinomas (Gerarde and Gerarde, 1974; MacIntyre, 1975). Because of the dichloro-substitution, this compound may be metabolized differently from benzidine, which is carcinogenic by virtue of metabolic activation.

The molecular formula of 3,3'-dichlorobenzidine (4,4'-diamino-3,3'-dichlorobiphenyl) is C-12 H-10 Cl-2 N-2 and the molecular weight is 253.13 (Stecher, 1968).

DCB forms brownish needles with a melting point of 132°C to 133°C (Pollock and Stevens, 1965). It is readily soluble in alcohol, benzene, and glacial acetic acid (Stecher, 1968), slightly soluble in HCl (Radding, et al. 1975), and sparingly soluble in water (0.7 g/l at 15° (Stecher, 1968). When combined with ferric chloride or bleaching powder, a green color is produced (Pollock and Stevens, 1965).

DCB's affinity for water-suspended particulate matter and soils is uncertain; the fact that it is an organic base suggests that it may be strongly bound to soil materials (Radding, et al. 1975).

Pyrolysis of DCB will most likely lead to the release of HCl. Because of the halogen substitution, DCB compounds probably biodegrade at a slower rate than benzidine alone. The photochemistry of DCB is unknown.

Assuming the clean air concentrations of ozone ( $2 \times 10^{-9}\text{M}$ ) and an average atmospheric concentration of hydroxyl radicals ( $3 \times 10^{-15}\text{M}$ ), the half-life for oxidation of DCB's by either of these chemical species is on the order of one and one to ten days, respectively. Furthermore, assuming a representative concentration of  $10^{-10}\text{M}$  for peroxy radicals in sunlit oxygenated water, the half-life for oxidation by these species is approximately 100 days, given the variability of environmental, atmospheric, and aqueous conditions (Radding, et al. 1975).

There are few data available on the bioconcentration, bioaccumulation, and biomagnification of DCB in the aquatic environment. DCB has been shown to be experimentally bioconcentrated by fish to a significant degree - approximately 1150 fold (Appleton and Sikka, Manuscript). However, no DCB was detected in fish sampled from the vicinity of a DCB-contaminated waste lagoon using analytical methods with sensitivities of 10 to 100  $\mu\text{g/kg}$  (Diachenko, 1978).

## REFERENCES

Appleton, H.T., and H.C. Sikka. Accumulation, elimination, and metabolism of dichlorobenzidine in the Bluegill Sunfish. Manuscript.

Diachenko, G. 1978. Personal communication. U.S. Food and Drug Administration.

Gerarde, H.W., and D.F. Gerarde. 1974. Industrial experience with 3,3'-dichlorobenzidine. Epidemiological study of a chemical manufacturing plant. Jour. Occup. Med. 16: 322.

MacIntyre, D.I. 1975. Experience of tumors in a British plant handling 3,3'-dichlorobenzidine. Jour. Occup. Med. 17: 23.

Pollock, J.R.A., and R. Stevens, eds. 1965. Dictionary of organic compounds. Eyre and Spottiswoode, London.

Radding, S.B., et al. 1975. Review of the environmental fate of selected chemicals. U.S. Environ. Prot. Agency, Washington, D.C.

Rye, W.A., et al. 1970. Facts and myths concerning aromatic diamine curing agents. Jour. Occup. Med. 12: 211.

Stecker, P.G., ed. 1968. The Merck Index. 8th ed. Merck and Co., Rahway, N.J.

#### AQUATIC LIFE TOXICOLOGY

There are few data available on the toxicity of DCB to freshwater or saltwater organisms. DCB was found to be acutely toxic to bluegill sunfish at levels of 0.5 mg/l or greater in the water (Sikka, et al. 1978).

#### CRITERION FORMULATION

No freshwater or saltwater criterion can be derived for any dichlorobenzidine using the Guidelines because no Final Chronic Value for either fish or invertebrate species or a good substitute for either value is available. There are insufficient data to estimate a criterion using other procedures.



## REFERENCES

Sikka, H.C., et al. 1978. Fate of 3,3'-dichlorobenzidine in aquatic environment. U.S. Environ. Prot. Agency, 600/3-8-068.

### 3,3' DICHLOROBENZIDINE (DCB)

#### Mammalian Toxicology and Human Health Effects

##### EXPOSURE

##### Ingestion from Water

To date, few systematic measurements of DCB in water supplies have been undertaken. In one instance, analysis of purge wells and seepage water near a waste disposal lagoon receiving DCB-manufacture wastes showed levels of DCB ranging from 0.13 to 0.27 mg/l. High levels of benzidine (up to 2.5 mg/l) were also seen, which may have arisen from photo-degradation of DCB (Sikka, et al. 1978), since benzidine is no longer manufactured in the U.S. Several other dichlorobenzidine isomers were also detected at levels of 1 to 8 mg/l. The use of lagoons to handle DCB-containing wastes might lead to contamination of ground water and pose a threat to persons relying on nearby wells for drinking water.

Takemura, et al. (1965) analyzed the water of the Sumida River in Tokyo during 1964. This river receives the waste effluents of several dye and pigment factories. The presence of DCB was demonstrated by thin layer chromatography. Although levels of DCB itself were not quantified, colorimetric analysis revealed that total aromatic amine content of the water (including benzidine, dichlorobenzidine,  $\alpha$ -naphthylamine, and  $\beta$ -naphthylamine) reached levels up to 0.562 mg/l. The authors suggested that the presence of the free amines might be due to chemical reduction of the azo-dyes by the high levels of  $H_2S$  and  $SO_2$  in the river.

### Ingestion from Foods

Few studies have attempted to identify DCB as a contaminant of human food. Since DCB has never had an application as an agricultural or food chemical, the most likely source of dietary DCB would be through consumption of contaminated fish.

A bioconcentration factor (BCF) relates the concentration of a chemical in water to the concentration in aquatic organisms, but BCF's are not available for the edible portions of all four major groups of aquatic organisms consumed in the United States. Since data indicate that the BCF for lipid-soluble compounds is proportional to percent lipids and the amounts of various species consumed by Americans. A recent survey on fish and shellfish consumption in the United States (Cordle, et al. 1978) found that the per capita consumption is 18.7 g/day. From the data on the nineteen major species identified in the survey and data on the fat content of the edible portion of these species (Sidwell, et al. 1974), the relative consumption of the four major groups and the weighted average percent lipids for each group can be calculated:

<u>Group</u>	<u>Consumption (Percent)</u>	<u>Weighted Average Percent Lipids</u>
Freshwater fishes	12	4.8
Saltwater fishes	61	2.3
Saltwater molluscs	9	1.2
Saltwater decapods	18	1.2

Using the percentages for consumption and lipids for each of these groups, the weighted average percent lipids is 2.3 for consumed fish and shellfish.

An average measured steady-state bioconcentration factor of 500 was obtained for 3,3'-dichlorobenzidine in the whole body of bluegills (Appleton and Sikka, manuscript). These fish probably contained about one percent lipids. An adjustment factor of  $2.3/1.0 = 2.3$  can be used to adjust the measured BCF from the 1.0 percent lipids of the bluegill to the 2.3 percent lipids that is the weighted average for consumed fish and shellfish. Thus, the weighted average bioconcentration factor for 3,3'-dichlorobenzidine and the edible portion of all aquatic organisms consumed by Americans is calculated to be  $500 \times 2.3 = 1,150$ .

No DCB was detected in fish sampled from the vicinity of a DCB-contaminated waste lagoon using analytical methods with sensitivity of 10 - 100 ug/kg (Diachenko, 1978).

#### Inhalation

The physical properties of DCB (low volatility, large crystal structure) probably minimize the risk of exposure of general populations to DCB through inhalation of air contaminated through industrial processes. However, inhalation might represent a major source of occupational exposure under sub-optimum working conditions. Akiyama (1970) examined the exposure of workers to DCB in a pigment plant in Japan and determined that during the addition of DCB to reaction vessels for synthesis of DCB pigments, the concentration of DCB in air reached  $2.5 \text{ mg}/100 \text{ m}^3$  in 10 minutes of charging of reaction vessels, and decreased to  $0.2 \text{ mg}/100 \text{ m}^3$  within 20 minutes. The distance of the sampling device from the operation was not specified. Also, the amount of total aromatic amines was elevated in exposed personnel (presumably

due to the presence of DCB). The mean urinary concentration of aromatic amines in process workers charging the reaction vessels with DCB and plant laboratory workers was 20.1 ppm and 21.1 ppm, respectively. Levels were only 14.5 ppm in workers who dried and cracked the pigments, 12.7 ppm in office clerks, and 13.6 ppm in controls (medical students). Although the concentrations detected were highly variable (i.e., the mean 20.1 ppm from the charging personnel was derived from data ranging from 48.5 to 10 ppm), it is possible that the elevated levels result from DCB exposure, since Akiyama claims that few or no precautions were taken to prevent exposure, particularly on hot days. It is uncertain whether the amines entered the body through respiration or through dermal absorption.

Gerarde and Gerarde (1974) reported on an industrial process in which both DCB and the DCB diarylide pigments were manufactured. Most steps in the process were performed in closed systems, and the DCB was handled in a salt form in a slurry (ca. 80 percent water content). DCB dust was said not to be a problem. The possibility that DCB contamination exposure could, however, occur is indicated by the statement that "...the floor and accessible surfaces contaminated with the slurry were usually hosed down to prevent accumulation of dried material...." Also, an outbreak of dermatitis in the plant was attributed to a process change in DCB production. In utilizing DCB in pigment production, the major sources of potential exposure are listed as the weighing process and charging of the tanks. Prior to May 1973, operators wore gloves and goggles but not dust face

masks. DCB was manufactured in this plant from 1938 to 1957. Thereafter, DCB was purchased from an outside supplier. On-site inspection of three DCB utilizing plants showed that two of the plants posed relatively low exposure potential which was due to use of metal reactors and protective arrangements at the point of tank charging. However, in the third plant, chemicals were dumped into open reaction vessels from an elevated platform, posing an enhanced potential for exposure. Therefore, a great deal of variability concerning the exposure of individuals to DCB may exist among various operations.

#### Dermal

Because of large particle size and increased usage of closed systems and protective clothing, dermal absorption of DCB probably represents a relatively minor route of DCB exposure in humans at present. However, Meigs, et al. (1954) presented some experimental evidence that under certain environmental conditions favoring moist skin conditions, such as high relative humidity and high air temperature, the dermal absorption by humans of benzidine and possibly other congenors such as DCB may be enhanced. An experiment using one rabbit suggested that exposure to DCB may occur indirectly by reductive metabolism of DCB-based azo pigments, liberating the free amine in vivo (Akiyama, 1970).

### PHARMACOKINETICS

#### Absorption

Virtually no information exists that quantifies the degree and rate of absorption of DCB in experimental animals

or in humans, although Meigs, et al. (1954) detected DCB in the urine of DCB process and manufacturing workers.

#### Distribution

A detailed distributional study of DCB in rats, monkeys, and dogs given 0.2 mg/kg of  $^{14}\text{C}$ -DCB by intravenous injection was reported by Kellner, et al. (1973). The results indicate a rather general distribution within the body after a 14-day observation period with highest levels found in the liver of all three species. The bile of monkeys and the lungs of dogs showed significant levels.

#### Metabolism

Little information exists on the metabolism of DCB in vertebrates.

DCB metabolites have not been detected in the excreta of dogs administered DCB by intraperitoneal injection (Sciarini and Meigs, 1961), or orally (Gerarde and Gerarde, 1974).

Kellner, et al. (1973) examined the urine of a rhesus monkey given 0.2 mg/kg  $^{14}\text{C}$ -DCB intravenously and found that in the first four hours following injection, about one-third of the urinary  $^{14}\text{C}$  was unchanged DCB, with another third identified as mono-N-acetyl DCB, based on chromatographic properties. The remainder of the urinary  $^{14}\text{C}$  was not recoverable into ether at pH = 11. At later intervals, mostly metabolites were excreted, with non-extractable  $^{14}\text{C}$  comprising the majority of this material.

No ortho hydroxy metabolites of DCB were detected in the urine of human subjects after oral dosing (Gerarde and Gerarde, 1974).

Aksamitnaia (1959) reported that prolonged ingestion (7.5-8.5 months) of small doses or a single large dose of DCB in rats led to the appearance of four transformation products, including benzidine and possibly glucuronide conjugates. At best, this conclusion is tenuous because analysis was done by paper chromatography (one solvent system) without benefit of radiotracer techniques, and the products were not quantified or further characterized. DCB was never detected in the urine in any of the experiments, and products were not seen until over 7 months of chronic DCB ingestion.

In a study of the bioconcentration of DCB in bluegill sunfish, over one-half of the DCB residues in the fish were in the form of a conjugate which, under very mildly acidic conditions, hydrolyzed to reform free DCB (Sikka, et al. 1978).

Hirai and Yasuhira (1972) noted that DCB was not oxidized by cytochrome c, whereas benzidine and other derivatives were oxidized.

The majority of information available at present suggests that DCB is resistant to metabolism, with the exception of certain conjugative mechanisms and possibly certain bioactivation steps. Ring chlorination of benzidine probably blocks ring hydroxylation reactions of DCB for both electronic and steric reasons (Shriner, et al. 1978).

#### Excretion

The excretion of DCB and metabolites following a 0.2 mg/kg intravenous dose of  $^{14}\text{C}$ -DCB was studied by Kellner, et al. (1973) in rats, dogs, and monkeys. With all species,



measurable elimination had ceased within 7 days of administration (Table 1). Fecal excretion was the predominant route of elimination in rats and dogs, and possibly in monkeys.

Sciarini and Meigs (1961) also noted a preponderance of fecal elimination of DCB in dogs. Finally, Gerarde and Gerarde (1974) cite an unpublished study utilizing human volunteers which concluded that DCB is excreted largely by the fecal route in man as well as in dogs.

Insufficient data is available to assess the ability of the body to accumulate significant burdens of DCB through repeated exposures.

#### EFFECTS

##### Acute, Sub-acute, and Chronic Toxicity

Gaines and Nelson (1977) reported the acute oral toxicity of DCB to male and female mice. The  $LD_{50}$  (mg/kg/day) for DCB given daily for seven days was 352 for female mice (slope = 27.39) and 386 for male mice (slope = 23.15). The single dose  $LD_{50}$  (mg/kg) was 488 for females and 676 for males.

Gerarde and Gerarde (1974) listed results of several toxicological studies with DCB. DCB-dihydrochloride failed to produce skin irritation in rabbits at an unspecified dose. An intradermal dose of 700 mg/kg also gave a negative reaction. One hundred mg of DCB-free amine placed in the conjunctival sac of the eye of a rabbit gave a negative reaction, while 20 mg of DCB dihydrochloride produced erythema, pus, and opacity of the eye, giving a score of 84 of a possible 110 in one hour according to the method of

TABLE 1  
Excretion of  $^{14}\text{C}$ -DCB in Rats, Dogs and Monkeys Administered  $^{14}\text{C}$ -DCB

Species; compound	Interval	Elimination of Total Dose Administered					
		Urine	Feces	$t_{50}$ Phase IV (hr)	Cage, feed	Balance	Residues
	(d)	%	%		%	0-7 day %	%
Rat	0-6	18 $\pm$ 4	79 $\pm$ 12	45	1	98 $\pm$ 12	2
Dog	0-7	8 $\pm$ 6	84 $\pm$ 11	--	5	97 $\pm$ 8	3
Monkey 1	0-7	27	46	--	21	94	
Monkey 2	0-7	37	26	--	20	83	

Adapted from Kellner, et al. Arch. Toxicol. 1973.

Draize. The oral LD<sub>50</sub> was given as 7.07 g/kg in albino rats for DCB free amine, and 3.82 g/kg in male and female Sprague-Dawley rats for DCB dihydrochloride. For topical application to skin, an LD<sub>50</sub> of 8 g/kg in male and female rats was seen. Pliss (1959) noted that rats given 120 mg of DCB subcutaneously exhibited a state of excitation with short-lived convulsions.

DCB was acutely toxic to bluegill sunfish at levels of 0.5 mg/l or greater in the water (Sikka, et al. 1978).

No incidence of human fatality resulting from exposure to DCB has been noted.

Ten rats exposed to a concentrated atmospheric dust of DCB dihydrochloride for 14 days showed, upon autopsy, slight to moderate pulmonary congestion and one pulmonary abcess (Gerarde and Gerarde, 1974). An irritant effect from HCl cannot be discounted in the study.

Freeman, et al. (1973) noted that DCB was cytotoxic to embryonic rat cells in culture at concentrations of 5 ppm or greater.

No mortalities were obtained in inhalation studies where rats were exposed to a concentrated atmosphere of concentrated DCB dihydrochloride dust for 14 days, or to 355 mg DCB free amine for 2 hours daily for 7 days (Gerarde and Gerarde, 1974).

Gerarde and Gerarde (1974) listed the principal reasons for visits to a company medical clinic by employees working with DCB. These were as follows: 1) gastro-intestinal upset; 2) upper respiratory infection; 3) sore throat; 4)

caustic burns; 5) headache; 6) dizziness; 7) dermatitis. The only illness apparently directly related to DCB was dermatitis. An outbreak of dermatitis was attributed to a manufacturing process change which led to small amounts of DCB-free base in the isolated DCB sulfate salt. Two cases of acute cystitis were found in the medical record review of the workers. One was of infectious origin and the other related to the presence of renal calculi. Cystoscopic examination of three other workers with urinary system symptoms revealed two had renal calculi, and another had cystitis cystica.

#### Synergistic or Antagonistic Compounds

No data are available concerning compounds which synergize or antagonize the toxicity of DCB.

#### Teratogenicity

No information is available defining the teratogenic potential of DCB. While perhaps not directly relevant to the question of DCB-induced teratogenesis, several studies show that DCB can cross the placental barrier and can also affect developmental systems.

DCB has been demonstrated to significantly increase the incidence of leukemia in the offspring of pregnant female mice given comparatively low doses (ca. 8-10 mg) of DCB by subcutaneous injection in the last week of gestation (Golub, et al. 1974). This could have been due to post-natal transfer of DCB to the young through lactation. However, transplacental effects of DCB have also been observed. Shabad, et al. (1972) and Golub (1969) noted that kidney

tissue taken from embryos of pregnant female mice treated with DCB exhibited altered behavior in organ culture, including increased survival and hyperplastic changes in epithelium not seen in controls.

The degree of exposure of pregnant women to DCB is probably low. The work force involved in the manufacture and utilization of DCB is predominantly or totally male. MacIntyre (1975) lists 5 women, all between the ages of 20 and 34 years, as having been DCB service or production workers in a plant in Great Britain as opposed to 217 men. Therefore, such exposure may be through consumption of contaminated fish or water, from DCB impurity in various DCB-based pigment products, or in vivo release of DCB from absorbed or ingested pigments. There is little evidence for any of these occurrences at present.

#### Mutagenicity

Garner, et al. (1975) compared the relative mutagenicity of benzidine, DCB, and other analogs in the bacterial mutagenesis system developed by Ames, et al. (1973), utilizing the Salmonella typhimurium tester strain TA 1598, an indicator of frame shift mutagenesis. The relevant data are summarized in Table 2. These results show that DCB is considerably more potent as a frameshift mutagen in this system than is benzidine. Also, a low degree of mutation is elicited by DCB but not by benzidine in the absence of the S-9 activation enzyme system.

TABLE 2  
Mutagenicity of DCB in the Ames Assay

	<u>µg chemical/ plate</u>	<u>S-9<sup>2</sup></u>	<u>Revertants/ plate</u>
3,3'-dichlorobenzidine	50	+	3360
	100	+	7520
	50	-	114
	100	-	131
3,3'-dichlorobenzidine sulfate salt, technical grade	50	+	5490
	100	+	8350
	50	-	127
	100	-	129
benzidine	50	+	430
	100	+	640
	50	-	5
	100	-	15
Dimethyl sulfoxide (control)		+	16
		-	8

<sup>1</sup>Adapted from Garner, et al. Cancer Lett. 1:39, 1975.

<sup>2</sup>S9 is the NADPH-fortified rat liver activation enzyme preparation.  
+ signifies preparation present; -, preparation absent.

Similar observations were made by Lazear and Louis (1977), utilizing an enzyme activation system obtained from the livers of male mice and Ames tester strain TA98 (an indicator of frameshift mutation). As before, DCB was much more mutagenic than benzidine and, unlike benzidine, retained an appreciable mutagenic activity without the liver enzymes. DCB was also slightly mutagenic towards tester strain TA100, indicating base-pair substitution mutation.

#### Carcinogenicity

Stula, et al. (1975) maintained 50 male and 50 female rats on a dietary level of DCB of 1000 mg/kg. The average 50 percent survival was 356 days with average days on the test of 349 days for females and 353 days for males. The range of days on the test was 118-486 days for males and 143-488 days for females. The rats were 38 days old at the start of the assay and were apparently autopsied at time of death or after 486-488 days (not specified). The results of this study are listed in Table 3.

TABLE 3  
Induction of Cancer in Male and Female Rats  
by 1000 ppm Dietary DCB

Type of Cancer	No. of Cancers			
	Male <sup>3</sup> DCB	vehicle control	Female <sup>3</sup> DCB	vehicle control
Mammary adenocarcinoma	7 <sup>2</sup>	0	26 <sup>2</sup>	3
Granulocytic leukemia	9 <sup>2</sup>	2	0	0
Zymbal gland carcinoma	8 <sup>2</sup>	0	1	0

<sup>1</sup>Adapted from Stula, et al. Toxicol. Appl. Pharmacol. 1975.

<sup>2</sup>Significantly greater than controls at p 0.05

<sup>3</sup>The number of animals examined histologically was 44 each for male and female.

In addition to the cancers listed in Table 3, the occurrence of malignant lymphoma was elevated over controls but not at statistically significant ( $p < 0.05$ ) levels. No bladder cancer was noted.

In a recent study, Stula, et al. (1978) reported on the induction of both papillary transitional cell carcinomas of the urinary bladder and hepatic carcinomas in female beagle dogs. An oral dose of 100 mg DCB was administered to the experimental animals, 3 times per week for 6 weeks, then 5 times per week continuously for periods up to 7.1 years. DCB was found to be carcinogenic at statistically significant levels ( $p < .025$ ). The incidences of hepatic carcinomas were 4/5 and 0/6 in DCB-treated and control groups, respectively. The incidences of urinary bladder carcinomas were 5/5 and 0/6, respectively.

For 12 months, 6 times weekly, Pliss (1959) added 0.5 to 1.0 ml of a 4.4% suspension of DCB to the feed of rats (mixed sex) of a strain assumed by Pliss to have a low spontaneous tumor rate. Each rat received a total dose of 4.53 g. Neoplasms were detected in 22 of 29 (75.8 percent) surviving animals. Tumors, primarily carcinomas, were observed in a broad spectrum of organs including mammary gland, Zymbal glands (sebaceous gland of the external auditory meatus), bladder, skin, small intestine, liver, thyroid gland, kidney, hematopoietic (lymphatic) system, and salivary glands.

An assay of DCB carcinogenicity was also done with mice (Pliss, 1959). The mice received 0.1 ml of a 1.1 percent DCB suspension in their food for 10 months, receiving a



total dose of 127.5 to 135 mg DCB. Hepatic tumors were found in 4 of 18 mice surviving after 18.5 months (22.2 percent). A sebaceous gland carcinoma and a lung adenoma were also seen.

The Pliss studies show that DCB may possess carcinogenic activity in both rats and mice. However, the massive and apparently acutely-toxic dose levels employed, the uncertain purity of the commercial product used, the virtual lack of dose-response data, and the lack of adequate controls limit the studies' utility for assessing human health hazards.

Carcinogenicity assays were performed using rats and mice which received DCB by subcutaneous injection (Pliss, 1959; Pliss, 1963), but are not considered here because of the irrelevancy of the subcutaneous route of administration to human exposure.

Griswold, et al. (1968) examined the potency of cancer induction by DCB, benzidine, and other compounds, using induction of mammary cancer in young female Sprague-Dawley rats as the major index. Forty day-old female Sprague-Dawley rats were given 30 mg of DCB every 3 days for 30 days by gavage and were then observed for 9 months. Under the conditions of this assay, DCB was ineffective as a mammary carcinogen but benzidine was highly effective at lower doses.

Sellakumar, et al. (1969) maintained male and female hamsters on a diet containing 0.1 percent (1000 ppm) of DCB. The duration of the study was not specified. With 30 animals of each sex, no cancer was observed. However,

at 0.3 percent dietary DCB, 4 transitional cell bladder carcinomas, some liver tumors and diffuse chronic intrahepatic obstructing cholangitis were seen. At 0.1 percent in the diet of benzidine, many liver tumors were obtained but no bladder cancer was found.

DCB was also found to produce transformation in cultured rat embryo cells infected with Rauscher leukemia virus (Freeman, et al. 1973). The index of transformation was the development of macroscopic foci of spindle cells, lacking polar orientation and contact inhibition. Cells from typical foci were tumorigenic when transplanted into newborn Fisher rats. Transplantability of DCB-transformed cells was not specified. DCB-induced transformation was seen at concentration of 5 ppm in the medium, but not at 1 ppm. Levels of 10 ppm or higher were cytotoxic. The in vitro test system detected transformation-activity in 6 of 7 aromatic amines characterized as active in vivo carcinogens, 1 of 2 aromatic amines classed as weak in vivo carcinogens, and 0 of 3 aromatic amines classed as non-carcinogenic in vivo. DCB was classed by the authors as a weak carcinogen.

The history of human industrial experience with DCB has been summarized and analyzed by Gerarde and Gerarde (1974) and Rye, et al. (1970) in the United States; by MacIntyre (1975) and Gadian (1975) in Great Britain; and by Akiyama (1970) in Japan. The concensus of these authors, achieved through epidemiological studies, is that there is no evidence that DCB itself has induced bladder cancer, the characteristic lesion induced by benzidine, naphthylamine, and other carcinogenic aromatic amines used in the dye and pigment industry.

The case for DCB carcinogenicity has been made largely on the basis of its structural similarity to benzidine and its tumorigenicity in several species of animals (MacIntyre, 1975). One problem associated with epidemiological studies of DCB effects in humans is that the population which has been exposed only to DCB is small. Many workers have also handled benzidine or other carcinogens. Also, the characteristic latency period for induction of bladder cancer by chemicals is quite long, exceeding 16 years for benzidine (Haley, 1975), and may not have elapsed for many workers. Finally, most of these studies have focused solely upon bladder cancer as the disease of interest. As discussed below, this approach may be misleading and fallacious in view of the pattern of DCB carcinogenesis in animals and the nature of cancer observed in DCB process workers.

Gadian (1975) examined the health records of 59 workers at a dyestuff plant in Great Britain who were exposed from 1953 through 1973 to DCB only and compared them to those working with both benzidine and DCB, and to unexposed populations. This time was justified as the average latency period for chemically-induced bladder cancer in humans (ca. 18 years). It was calculated that the DCB process worker was actually exposed to DCB for a maximum of 10 hours per work week. Men whose total DCB exposure was less than 245 hours (6 months full time work) were excluded from the study, leaving 35 segregated DCB workers. These 35 workers, representing a total of 68,505 hours of DCB exposure, had no urinary tract tumors, no other tumors, and two deaths from other causes (coronary thrombosis, cerebral hemorrhage).

In contrast, among 14 mixed benzidine and DCB workers with 16,200 hours exposure (approximately 60 percent worked with benzidine, 40 percent worked with DCB), three men developed tumors of the bladder, and one man developed carcinoma of the bronchus. One death from coronary thrombosis occurred. Since the use of benzidine ceased in 1964, the mixed group had a longer time to develop tumors than the DCB-segregated group. Therefore, the DCB-alone hours worked during the same period (1953-1964) as the mixed group was 31,945 hours. These results, while admitting that the population studied was small, were taken as evidence that DCB can be safely used if the provisions of the Carcinogenic Substances Regulations are observed.

MacIntyre (1975) also surveyed the health history of a DCB-utilizing plant in Great Britain. It was noted that the vast majority (209 out of 217) of production and service workers had received first exposure to DCB less than 20 years before the time of the report, indicating that the latent period for tumor formation might not have elapsed. Only 3 of the 217 exposed workers were deceased. The causes of death were amyotrophic lateral sclerosis (age 55 years, 15 years of DCB exposure, 39 years since first exposed), carcinoma of the lung (age-61 years, 1 year of DCB exposure, 12 years since first exposed), and pneumonia (age - 70 years, 10 years of DCB exposure, 43 years since first exposed). Three other employees who had not been exposed to DCB died of bronchial carcinoma. All employees exposed to DCB since 1965 have received cytological testing twice yearly, with

all tests proving negative. A 1974 meeting of occupational physicians is also cited, stating that in Europe approximately 1000 persons have been exposed to DCB with a zero incidence of bladder cancer.

Gerarde and Gerarde (1974) reported the results of an epidemiological study of workers exposed to DCB in manufacture and utilization in a plant in the United States. A survey of the number of DCB-exposed workers who developed neoplasms and the type of neoplasm was presented. These included lung cancer (2 workers), leukemia-bone marrow (1), lipoma (6), rectum-papilloma (3), sigmoid colon carcinoma (2), prostate carcinoma (1), breast muscle myoblastoma (1), skin basal cell epithelioma (1). A total of 17 workers of the total of 207 workers surveyed had developed neoplasma.

The etiology of bladder cancer was discussed and the data treated, using several epidemiological and statistical approaches. According to this approach, if DCB were as potent as benzidine as a bladder carcinogen, a total of 22 cases of bladder cancer out of 163 DCB production workers would have been observed, whereas none were seen. The possible induction by DCB of tumors at sites other than the bladder was not considered.

### Summary

Based upon existing data, there is little doubt that DCB is carcinogenic in several animal species including rats, mice, hamsters, and dogs. According to current methodology, the experimental evidence serves as an indication that a potential carcinogenic risk is posed to man. DCB induces tumors in a variety of tissues in animals, with mammary, hematopoietic, and skin (Zymbal gland) tissue being the most affected. Many of the tumors have been characterized as malignant.

## CRITERION FORMULATION

### Existing Guidelines and Standards

The American Conference of Governmental Industrial Hygienists (1977) has recommended that no exposure to DCB by any route should be permitted, because of a demonstrated high carcinogenic response in animals. Strict regulations have recently been promulgated by the Occupational Safety and Health Administration to minimize or eliminate occupational exposure to DCB (CFR, 1977). To date, no standards have been placed on permissible levels of DCB in the environment or in food.

### Current Levels of Exposure and Special Groups at Risk

It is estimated that between 250 and 2500 workers receive exposure to DCB in the U.S., compared to 62 for benzidine (Fishbein, 1977). Given the stringent precautions which must be taken in the manufacture and use of DCB, the level of exposure may be minimal at present, although no data is available. However, past exposure of individuals working without benefit of protective measures must present a cause for concern. In addition, the general population may receive exposure to DCB through contaminated drinking water or food (fish), although there is no significant evidence for this at the present.

Additional groups that may be at risk include worker in the printing or graphic arts professions handling the DCB-based azo pigments. DCB may be present as an impurity in the pigments, and there is some evidence that DCB may be metabolically liberated from the azo pigment. More information on the level of exposure to the pigments is needed.

### Basis and Derivation of Criterion

The safe dose of DCB in water was calculated from the carcinogenicity assays, using a linear, non-threshold mathematical model described in the appendix. The calculation assumes a risk of 1 in 100,000 of developing cancer as a result of daily consumption of 2 liters of water and DCB-contaminated fish having a bioconcentration factor of 1150. Although several carcinogenicity studies are available for use in calculating a criterion for DCB in drinking water, only the work of Stula and coworkers (1975, 1978) was considered, since the studies by Pliss (1959, 1963) lack appropriate control data. More specifically, the data on induction of papillary transitional cell carcinomas of the urinary bladder and hepatic carcinomas in female beagle dogs (Stula, et al. 1978) were chosen as a base for the calculations. Based on these data, a DCB criterion of  $1.69 \times 10^{-2}$   $\mu\text{g/l}$  is judged to be adequate to protect the population consuming the water. This dose is low from an occupational viewpoint and should justify efforts to eliminate exposure of workers to DCB.

Under the Consent Decree in NRDC vs. Train, criteria are to state "recommended maximum permissible concentrations (including where appropriate, zero) consistent with the protection of aquatic organisms, human health, and recreational activities." DCB is suspected of being a human carcinogen. Because there is no recognized safe concentration for a human carcinogen, the recommended concentration of DCB in water for maximum protection of human health is zero.



Because attaining a zero concentration level may be infeasible in some cases and in order to assist the Agency and States in the possible future development of water quality regulations, the concentrations of DCB corresponding to several incremental lifetime cancer risk levels have been estimated. A cancer risk level provides an estimate of the additional incidence of cancer that may be expected in an exposed population. A risk of  $10^{-5}$  for example, indicates a probability of one additional case of cancer for every 100,000 people exposed, a risk of  $10^{-6}$  indicates one additional case of cancer for every million people exposed, and so forth.

In the Federal Register notice of availability of draft ambient water quality criteria, EPA stated that it is considering setting criteria at an interim target risk level of  $10^{-5}$ ,  $10^{-6}$  or  $10^{-7}$  as shown in the table below.

<u>Exposure Assumptions</u>	<u>Risk Levels and Corresponding Criteria</u> <sup>(1)</sup>			
	<u>0</u>	<u><math>10^{-7}</math></u>	<u><math>10^{-6}</math></u>	<u><math>10^{-5}</math></u>
2 liters of drinking water and consumption of 18.7 grams of fish and shellfish (2)	0	$1.69 \times 10^{-4}$ $\mu\text{g/l}$	$1.69 \times 10^{-3}$ $\mu\text{g/l}$	$1.69 \times 10^{-2}$ $\mu\text{g/l}$
Consumption of fish shellfish only.	0	$1.85 \times 10^{-4}$ $\mu\text{g/l}$	$1.85 \times 10^{-3}$ $\mu\text{g/l}$	$1.85 \times 10^{-2}$ $\mu\text{g/l}$

(1) Calculated by applying a modified "one hit" extrapolation model described in the FR 15926, 1979. Appropriate bioassay data used in the calculation of the model are presented in Appendix 1. Since the extrapolation model is linear to low doses, the additional lifetime risk is directly proportional to the water concentration. Therefore, water concentrations corresponding to other risk levels can be derived by multiplying or dividing one of the risk levels and corresponding water concentrations shown in the table by factors such as 10, 100, 1,000, and so forth.

(2) Ninety-one percent of DCB exposure results from the consumption of aquatic organisms which exhibit an average bioconcentration potential of 1150 fold. The remaining 9 percent of DCB exposure results from drinking water.

Concentration levels were derived assuming a lifetime exposure to various amounts of DCB, (1) occurring from the consumption of both drinking water and aquatic life grown in water containing the corresponding DCB concentrations and, (2) occurring solely from consumption of aquatic life grown in the waters containing the corresponding DCB concentrations.

Although total exposure information for DCB is discussed and an estimate of the contributions from other sources of exposure can be made, this data will not be factored into the ambient water quality criteria formulation because of the tenuous estimates. The criteria presented, therefore, assume an incremental risk from ambient water exposure only.

## APPENDIX I

### Summary and Conclusions Regarding the Carcinogenicity of 3,3'-Dichlorobenzidine (DCB)\*

3,3'-Dichlorobenzidine (DCB) is used as an intermediate in the synthesis of dyes and pigments. It is structurally related to carcinogenic aromatic amines, which have been used in the dye and pigment industries.

Five epidemiological studies of employees handling DCB in chemical plants in the United States, Great Britain, and Japan have provided no evidence of DCB-induced cancers. However, investigative problems associated with these studies, such as too short a follow-up time and small sample size, make them unreliable as the sole basis for making conclusions about human cancer risks from DCB.

DCB has induced carcinomas in three species of experimental animals receiving oral doses of the chemical. Dogs (female) developed papillary transitional cell carcinomas of the urinary bladder and hepatocellular carcinomas. Hamsters developed transitional cell bladder carcinomas, liver cell and cholangiomatous tumors. Rats developed mammary adenocarcinomas (male and female), granulocytic leukemia (males), and Zymbal gland carcinomas (males).

Two studies of the mutagenicity of DCB showed that it was mutagenic in two Salmonella typhimurium tester strains (TA 1538, TA 98) in the presence and absence of an S-9 liver enzyme system. DCB also transformed cultured rat embryo cells and the transformed cells were tumorigenic when transplanted into newborn rats.

The carcinogenic, mutagenic, and transforming activities of DCB in laboratory organisms and its chemical similarity to benzidine, a human bladder carcinogen, are strong evidence that it is likely to be a human carcinogen.

The water quality criterion for DCB is based on the induction of papillary transitional cell carcinomas of the urinary bladder and hepatic carcinomas in female beagle dogs, given an oral dose of 100 mg 3,3'-dichlorobenzidine, 3 times per week for 6 weeks, then 5 times per week continuously for up to 7.1 years (Stula, et al. 1978). Dose-response data for dogs were selected because dogs developed urinary bladder tumors, as do humans, when exposed to certain aromatic amines. The concentration of DCB in water, calculated to keep the lifetime cancer risk below  $10^{-5}$ , is  $1.69 \times 10^{-2}$  micrograms per liter.

\*This summary has been prepared and approved by the Carcinogens Assessment Group, EPA, on June 15, 1979.

### Summary of Pertinent Data

The water quality criterion for DCB is based on the induction of papillary transitional cell carcinomas of the urinary bladder and hepatic carcinomas in female beagle dogs, given an oral dose of 100 mg DCB, three times per week for six weeks, then five times per week continuously for periods up to 7.1 years (Stula, et al. 1978). The incidence of urinary bladder carcinomas observed in DCB-treated dogs was 5/5 as compared to 0/6 in the control group. The incidences of hepatic carcinomas were 4/5 and 0/6 in DCB-treated and control groups, respectively. The criterion was calculated from the following parameters:

$n_{tu}$	= 4.5*	(urinary bladder carcinomas)
$N_{tu}$	= 5	
$n_{th}$	= 4	(hepatic carcinoma)
$N_{th}$	= 5	
$n_c$	= 0	d (timeweight average)
$N_c$	= 6	concentration) = 7.36 mg/kg/day
$L_c$	= 7.1 yrs.	F = 0.0187 kg
$le$	= 7.1 yrs.	R = 1150
$L$	= 8.65 yrs.	W = 11.391 kg

Based on these parameters, the one-hit slope ( $B_H$ ) is 1.036 (mg/kg/day)<sup>-1</sup> for urinary bladder carcinomas and 0.724 (mg/kg/day)<sup>-1</sup> for hepatic carcinomas. The resulting water concentration for DCB, calculated to keep the individual lifetime cancer risk below 10<sup>-5</sup>, is 1.69 x 10<sup>-2</sup> micrograms per liter.

\*This underestimate of the true number, based on Burkson's correction factor, was chosen in order to obtain a finite mathematical estimate.

## REFERENCES

- Akiyama, T. 1970. The investigation on the manufacturing plant of organic pigment. Jikei. Med. Jour. 17: 1.
- Aksamitnaia, I.A. 1959. Some data on transformation products of 3,3-dichlorobenzidine excreted with rat urine. Vopr. Onkol. 5: 18.
- American Conference of Governmental Industrial Hygienists. 1971. Documentation of the threshold limit values for substances in workroom air. 3rd Ed., 1977. : 77.
- Ames, B.N., et al. 1973. Carcinogens are mutagens: a simple test system combining liver homogenates for activation and bacteria for detection. Proc. Natl. Acad. Sci. U.S.A. 70: 2281.
- Appleton, H.T., and H.C. Sikka. Accumulation, elimination, and metabolism of dichlorobenzidine in the Bluegill Sunfish. Manuscript.
- Code of Federal Regulations. 1977. Title 29, Part 1910.
- Cordle, F., et al. 1978. Human exposure to polychlorinated biphenyls and polybrominated biphenyls. Environ. Health Perspect. 24: 157.

Diachenko. G. 1978. Personal communication, U.S. Food and Drug Administration.

Fishbein. 1977. Potential industrial carcinogens and mutagens. Office of Toxic Substances, Environ. Prot. Agency, Washington, D.C. p. 27.

Freeman, A.E., et al. 1973. Transformation of cell cultures as an indication of the carcinogenic potential of chemicals. Jour. Natl. Cancer Inst. 51: 799.

Gadian, T. 1975. Carcinogens in industry, with special reference to dichlorobenzidine. Chem. Ind. (Great Britain) 19: 821.

Gaines, T.B., and C.J. Nelson. 1977. In: 5th Anniversary Report, National Center for Toxicological Research (FOA). 77: 1029.

Garner, R.C., et al. 1975. Testing of some benzidine analogues for microsomal activation to bacterial mutagens. Cancer Lett. 1: 39.

Gerarde, H.W., and D.F. Gerarde. 1974. Industrial experience with 3,3'-dichlorobenzidine: an epidemiological study of a chemical manufacturing plant. Jour. Occup. Med. 16: 322.

Golub, N.I. 1969. Transplacental action of 3,3'-dichlorobenzidine and orthotolidine on organ cultures of embryonic mouse kidney tissue. Bull. Exp. Biol. Med. (U.S.S.R.) 68: 1280.

Golub, N.I., et al. 1974. Oncogenic action of some nitrogen compounds on the progeny of experimental mice. Bull. Exp. Biol. Med. (U.S.S.R.) 78: 62.

Griswold, D.P., et al. 1968. The carcinogenicity of multiple intragastric doses of aromatic and heterocyclic nitro or amino derivatives in young female Sprague-Dawley rats. Cancer Res. 28: 924.

Haley, T.J. 1975. Benzidine revisited: a review of the literature and problems associated with the use of benzidine and its congeners. Clin. Toxicol. 8: 13.

Hirai, K., and Y. Yasuhira. 1972. Mitochondrial oxidation of 3,3'-diaminobenzidine and related compounds, and their possible relation to carcinogenesis. GANN. 63: 665.

Kellner, H.M., et al. 1973. Animal studies on the kinetics of benzidine and 3,3'-dichlorobenzidine. Arch. Toxicol. 31: 61.

Lazear, E.J., and S.C. Louis. 1977. Mutagenicity of some congeners of benzidine in the Salmonella typhimurium assay system. Cancer Lett. 4: 21.



MacIntyre, I. 1975. Experience of tumors in a British plant handling 3,3'-dichlorobenzidine. Jour. Occup. Med. 17: 23.

Meigs, J.W., et al. 1951. A study of exposure of benzidine and substituted benzidine in a chemical plant. Arch. Ind. Hyg. Occup. Med. 4: 533.

Meigs, J.W., et al. 1954. Skin penetration by diamines of the benzidine group. Arch. Ind. Hyg. Occup. Med. 9: 122.

Pliss, G.B. 1959. Dichlorobenzidine as a blastomogenic agent. Vopr. Onkol. 5: 524.

Pliss, G.B. 1963. On some regular relationships between carcinogenicity of aminodiphenyl derivatives and the structure of substance. Acta Unio. Int. Contra. Cancrum. 19: 499.

Sciarini, L.J., and J.W. Meigs. 1961. Biotransformation of the benzidines. III. Studies on diorthotolidine, dianisidine, and dichlorobenzidine:3,3'-disubstituted congeners of benzidine (4,4'-diamino-biphenyl). Arch. Environ. Health 2: 108.

Sellakumar, A.R., et al. 1969. Aromatic amines carcinogenicity in hamsters. Proc. Am. Assoc. Cancer Res. 10: 78.

Shabad, L.M., et al. 1972. Transplacental effects of some chemical compounds on organ cultures of embryonic kidney tissue. Cancer Res. 32: 617.

Shriner, C.R., et al. 1978. Reviews of the Environmental Effects of Pollutants: II. Benzidine. ORNL/ELS-86 EPA 600/1-78-024.

Sidwell, V.D., et al. 1974. Composition of the edible portion of raw (fresh or frozen) crustaceans, finfish, and mollusks. I. Protein, fat, moisture, ash, carbohydrate, energy value, and cholesterol. Mar. Fish. Rev. 36: 21.

Sikka, H.C. 1978. Personal communication.

Sikka, H.C., et al. 1978. Fate of 3,3'-dichlorobenzidine in aquatic environments. U.S. Environ. Prot. Agency 600/3-8-068.

Stula, E.F., et al. 1975. Experimental neoplasia in rats from oral administration of 3,3'-dichlorobenzidine, 4,4'-methylene-bis(2-chloroaniline), and 4,4'-methylene-bis(2-methylaniline). Toxicol. Appl. Pharmacol. 31: 159.

Takemura, N., et al. 1965. A survey of the pollution of the Sumida River, especially on the aromatic amines in the water. Internat. Jour. Air Water Pollut. 9: 665.