

July 1984

Research and Development

ENVIRONMENTAL ASSESSMENT OF
A CRUDE-OIL HEATER USING STAGED AIR
LANCES FOR NO_X REDUCTION
Volume II. Data Supplement

Prepared for

Office of Air Quality Planning and Standards

Prepared by

Industrial Environmental Research
Laboratory
Research Triangle Park NC 27711

RESEARCH REPORTING SERIES

Research reports of the Office of Research and Development, U.S. Environmental Protection Agency, have been grouped into nine series. These nine broad categories were established to facilitate further development and application of environmental technology. Elimination of traditional grouping was consciously planned to foster technology transfer and a maximum interface in related fields. The nine series are:

- 1. Environmental Health Effects Research**
- 2. Environmental Protection Technology**
- 3. Ecological Research**
- 4. Environmental Monitoring**
- 5. Socioeconomic Environmental Studies**
- 6. Scientific and Technical Assessment Reports (STAR)**
- 7. Interagency Energy-Environment Research and Development**
- 8. "Special" Reports**
- 9. Miscellaneous Reports**

This report has been assigned to the INTERAGENCY ENERGY-ENVIRONMENT RESEARCH AND DEVELOPMENT series. Reports in this series result from the effort funded under the 17-agency Federal Energy/Environment Research and Development Program. These studies relate to EPA's mission to protect the public health and welfare from adverse effects of pollutants associated with energy systems. The goal of the Program is to assure the rapid development of domestic energy supplies in an environmentally-compatible manner by providing the necessary environmental data and control technology. Investigations include analyses of the transport of energy-related pollutants and their health and ecological effects; assessments of, and development of, control technologies for energy systems; and integrated assessments of a wide range of energy-related environmental issues.

EPA REVIEW NOTICE

This report has been reviewed by the participating Federal Agencies, and approved for publication. Approval does not signify that the contents necessarily reflect the views and policies of the Government, nor does mention of trade names or commercial products constitute endorsement or recommendation for use.

This document is available to the public through the National Technical Information Service, Springfield, Virginia 22161.

ENVIRONMENTAL ASSESSMENT OF A CRUDE-OIL HEATER USING STAGED AIR LANCES FOR NO_x REDUCTION

Volume II Data Supplement

By

R. DeRosier and B. DaRos
Acurex Corporation
Energy & Environmental Division
555 Clyde Avenue
P.O. Box 7555
Mountain View, California 94039

EPA Contract 68-02-3188

EPA Project Officer: Robert E. Hall
Industrial Environmental Research Laboratory
Research Triangle Park, North Carolina 27711

for

U. S. ENVIRONMENTAL PROTECTION AGENCY
Office of Research and Development
Washington, DC 20460

ABSTRACT

This volume of the report presents emission results obtained from field testing of a crude-oil process heater burning a combination of oil and refinery gas. The heater had been modified by the addition of a system for injection of secondary air to reduce NO_x emissions. One test was conducted with the staged air system (low-NO_x), and one was conducted without it (baseline). Tests included continuous monitoring of flue gas emissions and source assessment sampling system (SASS) sampling of the flue gas with subsequent laboratory analysis of samples utilizing gas chromatography (GC), and low resolution mass spectrometry (LRMS) for organics; and atomic absorption spectrometry (AAS) and spark source mass spectrometry (SSMS) for trace metals. Flue gas concentrations of NO_x were reduced 30 percent (from 83 to 56 ng/J) with the staged air system. Total organic emissions dropped from 17.1 to 3.4 mg/dscm from the baseline to the low-NO_x test. This was primarily due to a reduction in the C1 to C6 boiling point range compounds which constituted most of the organic emissions. GC/MS analysis identified 11 semivolatile priority pollutant compounds in both tests, most of them present in higher concentrations during the baseline test. LRMS analysis suggested the presence of eight compound categories in the organic emissions during the baseline test and four compound categories in the low-NO_x test. Biological tests indicated that the organic sorbent module extracts from both tests were of moderate toxicity and moderate-to-high mutagenicity.

TABLE OF CONTENTS

<u>Section</u>	<u>Page</u>
1 INTRODUCTION	1-1
2 PRELIMINARY EQUIPMENT CALIBRATION	2-1
3 HEATER OPERATING DATA	3-1
4 SAMPLING DATA SHEETS	4-1
4.1 KVB Continuous Emissions Monitoring Report	4-2
4.2 Operating Data for EPA Method 5	4-21
4.3 Operating Data for Controlled Condensation Train . .	4-34
4.4 Operating Data for SASS	4-41
5 ANALYTICAL RESULTS	5-1
5.1 Ultimate Analysis of Fuel Oil	5-2
5.2 Composition of Refinery Gas	5-4
5.3 SASS Particulate Emissions	5-7
5.4 EPA Method 5 Particulate Emissions	5-13
5.5 Sulfur Oxides Emissions by Turbidimetric Analysis .	5-23
5.6 Trace Element Analysis	5-28
5.7 Total Chromatographable Organics (TCO), Gravimetric Organics (Grav), Infrared (IR) Spectra, Gas Chromatography/Mass Spectrometry (GC/MS) and Low Resolution Mass Spectrometry (LRMS) of Total Sample Extracts	5-42
5.8 C ₁ -C ₆ Chromatograms	5-60
5.9 Radiometric Analysis Results	5-129
5.10 Bioassay Reports	5-131
Mutagenicity Evaluation of XAD Resin Extracts in the EPA Level 1 Ames Salmonella/Microsome Plate Test	5-132
Cytotoxic Evaluation of XAD Resin Extracts in the EPA Level 1 Rodent Cell (CHO) Clonal Toxicity Assay	5-168

SECTION 1

INTRODUCTION

The purpose of this data supplement is to document data in greater detail than was possible in Volume I (Technical Results) of this report. It is intended to provide sufficient detail for researchers to perform their own analysis of the data obtained. Readers are referred to the technical volume for objectives, description of source emission results, interpretation, and conclusions.

The remaining sections of this data supplement contain the following information:

Section 2 -- Preliminary Equipment Calibration Data

Section 3 -- Heater Operating Data: crude-oil temperatures and pressures, process rate, burner pressures, fuel flowrates

Section 4 -- Sampling Data Sheets: KVB continuous emissions monitoring report, operating data tables for EPA Method 5 (for particulate mass emissions), controlled condensation (for SO₂ and SO₃ sampling), and SASS (for trace element and organic sampling)

Section 5 -- Analytical Laboratory Results: ultimate analysis of the fuel oil, composition of reabsorber gas, SASS particulate emissions, sulfur oxides emissions by turbidometric analysis, trace element emissions by spark source mass spectrometry (SSMS) analysis and atomic absorption spectroscopy (AAS) analysis; total

chromatographable organic (TCO) and gravimetry (GRAV) results; infrared (IR) spectra; determination of organic compounds by gas chromatography/mass spectrometry (GC/MS); low resolution mass spectrometry (LRMS) of total sample extracts; C₁-C₆ chromatograms, radiometric analysis results, and biological assay reports on the SASS train samples.

SECTION 2
PRELIMINARY EQUIPMENT CALIBRATION

1-8-81

Date
Time 9:30 AM
Barometric Pressure 30.18
Ambient Temperature 64°

Orifice Meter
Orifice Magnehelic 80615 BW28
Primary Calibration Meter 433583
Control Module 084

Operators UNRUTH
Wet Bulb Temperature

METER CALIBRATION DATA

Orifice Manometer Δh_{ref} (in. w.g.)	Orifice Magnehelic Δh_0 (in. w.g.)	Primary Meter Δh_p (in. w.g.)	Dry Test Meter P_{dg}	Gas Volume Primary Meter V_p (ft. ³)	Gas Volume Dry Test Meter V_d (ft. ³)	Temperature						Time t (min.)	K_0		
						Primary Meter			Dry Test Meter						
						Inlet, T_{d1} (°F)	Outlet, T_{d2} (°F)	Avg., T_{da} (°F)	Inlet, T_{p1} (°F)	Outlet, T_{p2} (°F)	Avg., T_{pa} (°F)				
0.7				443.628	359.549	74	74	74	65	64	64	52	0	= .9960	
				461.882	377.375	64	68	70	63	65	64	52	0	$K_0 = .7167$	
0.4				18.754	17.966										= .4956
0.3				441.882	377.513	64	68	66.5	63	65	64	52	0	$K_0 = .7075$	
0.8				21.085	20.872										= .9983
1.0				482.467	348.347	67	67	67	63	65	65	33	0	$K_0 = .6943$	
				502.369	347.835	69	68	67.5	65	66	65.5	33	0	= 1.0006	
1.2				19.102	19.438										= .6860
1.6				502.564	348.835	69	65	67	62	64	66	31	0		
1.8				523.873	448.573	71	68	69	66	67	66	31	0		
2.0				20.868	20.682										

2.2						82	69								
2.4						533.000	450.010	82	69	66	67	30	0		= 1.0120
						591.052	505.313	64	76	80	67	68	67	0	$K_0 = 3.8005$
2.6						56.082	55.308								
2.8						591.082	505.313	94	76	83	74	68	67	28	0
						663.000	575.876	102	83	88	74	72	71	28	= 1.0193
3.0						71.918	72.578								= 3.6912
3.2						663.000	575.876	102	83	95	73	72	71	31	0
						253.457	669.561	107	88	95	73	72	71	31	= .9772
3.4						95.957	93.715								= 3.6244
3.6						133.387	669.561	747	98	103	73	72	73	17	0
						84.376	728.618	712	90	98	73	72	73	17	= 1.0283
3.8						60.439	59.057								= 3.5992
4.0															
4.2															
4.4															
4.6															
4.8															
5.0															

Averages: small $a = .9978$
 $K_0 = .7024$

large $a = 1.0092$
 $K_0 = 3.6803$

SECTION 3
HEATER OPERATING DATA

Process rate

Crude oil temperature and pressure

Inlet

Outlet

Reabsorber gas flow

Burner pressures

Oil

Steam

Gas

Crude API gravity

Tube temperatures

Notes

UNIT OPERATING DATA

Test No. 1 (BASELINE) Date 18 JUNE 1981 Location TOSCO Engr. R. DEROISIER
 Unit No. 11H13 Fuel Capacity 162.50 bbl/d, 55x10⁶ Btu/h
 Unit Type CRUDE HEATER Burner Type JOHN ZINK DBA-22
 Test Description: _____

Test number	1				
Time	11:56	12:50	1:50	3:50	4:50
Process Rate x 1847.7 bbl/d	6.3	6.3	6.3	6.3	6.3
Crude Tin (°F)	385	384	384	386	384
Crude Tout (°F) East	640	644	645	640	640
Crude Tout (°F) West	635	641	642	636	639
Crude Pin (psig) East	140	140	140	140	140
Crude Pin (psig) West	130	130	130	130	130
Crude Pout (psig)	--	--	--	34	33
Reabsorber gas in $\times 10^3$ scf/d)	2.1	2.1	2.2	2.1	2.0
Burner #1	44/80	50/79	50/80	50/80	52/80
Burner #2	44/80	44/80	44/82	44/80	44/80
Burner #3 Oil P/Steam P	--	--	--	--	--
Burner #4 (psig)	44/74	50/75	52/74	50/74	50/76
Burner #5	44/78	46/78	48/76	48/74	44/78
Burner #6	--	--	--	--	--
P to Burner #1 GAS	--	4.2	4.2	5.0	5.0
P to Burner #2	--	--	5.0	2.0	2.0
P to Burner #3	--	--	2.0	5.0	4.4
P to Burner #4	--	--	5.4	6.8	6.4
P to Burner #5	--	--	6.6	4.2	4.2
P to Burner #6	--	--	4.2	4.0	4.0
Air Registers (%open)	50	--	--	--	--
Crude API Gravity	22.5	22.5	22.5	22.5	22.5
Flame Observations	--	--	--	--	--
Tube 13 Pass A	710	763	765	766	762
Tube 13 Pass B	775	775	772	767	770
Tube 15 Pass A	735	733	736	739	732
Tube 15 Pass B	575	710	625	740	725

UNIT OPERATING DATA - CONTINUED

Test No. 1 Date 14 JUNE, 1981 Location TUSCO Engr. R. DROSIER
Unit No. 11H13 Fuel Capacity 16250 bbl/d, 55 x 10⁶ Btu/h
Unit Type CRUDE HEATER Burner Type JOHN ZINK DBA-22

Test Description:

UNIT OPERATING DATA

Test No. / Date 18 JUNE, 1981 Location TOSCO Engr. R. DEROSIER
 Unit No. 11H13 Fuel OIL/GAS Capacity 16250 bbl/d, 55×10^6 Btu/h
 Unit Type CRUDE HEATER Burner Type JOHN ZINK DBA-22

Test Description:

Test number	1			
Time	5:50	6:50	7:50	8:50
Process Rate x 1847.7 bbl/d	6.3	6.3	6.3	6.3
Crude Tin (°F)	388	383	383	383
Crude Tout (°F) East	635	642	643	643
Crude Tout (°F) West	635	638	636	632
Crude Pin (psig) East	140	140	140	140
Crude Pin (psig) West	130	130	130	130
Crude Pout (psig)	32	33	33	33
Reabsorber gas in 10^3 scf/d	2.4	2.0	2.2	2.2
Burner #1	52/80	50/80	52/80	50/80
Burner #2	43/81	44/80	43/80	43/81
Burner #3 Oil P/Steam P	--	--	--	--
Burner #4 (psig)	48/75	48/76	48/76	48/74
Burner #5	44/77	46/78	46/77	47/78
Burner #6	--	--	--	--
P to Burner #1	5.0	3.6	4.0	4.2
P to Burner #2	6.0	4.0	5.0	5.0
P to Burner #3 Gas P	2.2	2.0	1.8	2.0
P to Burner #4 (psig)	4.4	4.2	3.8	3.6
P to Burner #5	8.0	6.2	6.7	6.9
P to Burner #6	4.8	4.9	4.2	4.2
Air Registers (%open)	50	--	--	--
Crude API Gravity	22.5	--	--	--
Flame Observations	--	--	--	--
Tube 13 Pass A	760	765	765	762
Tube 13 Pass B	765	770	768	715
Tube 15 Pass A	732	735	735	735
Tube 15 Pass B	720	750	755	740

UNIT OPERATING DATA - CONTINUED

Test No. 1 Date 18 June, 1981 Location TOSCO Engr. R. DEROISIER

Unit No. 11H13 Fuel O/I GAS Capacity 16250 bbl/d, 55×10^6 Btu/h

Unit Type CRUDE HEATER Burner Type JOHN ZINK DBA-22

Test Description:

UNIT OPERATING DATA

Test No. 2 Date 19 JUNE, 1981 Location TOSCO Engr. R. DEROISIER
 Unit No. 11H13 Fuel OIL/GAS Capacity 16250 bbl/d, 55 x 10⁶ Btu/h
 Unit Type CRUDE HEATER Burner Type JOHN ZINK DBA-22
 Test Description: LOW-NOx - USING KUB LANCE

Test number	2				
Time	10:00	11:00	12:00	1:00	2:00
Process Rate x 1847.7 bbl/d	6.3	6.3	6.3	6.3	6.3
Crude Tin (°F)	385	383	384	384	384
Crude Tout (°F) East	643	645	642	644	639
Crude Tout (°F) West	639	644	642	646	641
Crude Pin (psig) East	140	140	140	140	140
Crude Pin (psig) West	130	130	130	130	130
Crude Pout (psig)	34	35	35	33	35
Reabsorber gas in $\times 10^3$ scf/d	2.2	2.1	2.1	2.1	2.0
Burner #1	50/80	50/80	50/80	50/80	52/80
Burner #2	44/80	42/80	43/80	43/80	44/80
Burner #3 Oil P/Steam P	--	--	--	--	--
Burner #4 (psig)	48/74	48/75	49/76	49/75	48/76
Burner #5	47/73	46/77	46/78	42/77	44/73
Burner #6	--	--	--	--	--
P to Burner #1	4.1	4.4	4.4	4.4	4.0
P to Burner #2	5.5	5.5	6.0	5.5	5.0
P to Burner #3 Gas P	2.0	2.0	2.0	2.0	2.0
P to Burner #4 (psig)	2.8	2.8	3.2	3.2	2.8
P to Burner #5	7.9	7.0	7.2	7.0	6.8
P to Burner #6	4.1	4.2	4.2	4.0	4.0
Air Registers (%open)	--	--	--	--	--
Crude API Gravity	23.0	23.0	23.0	23.0	23.0
Flame Observations	--	--	--	--	--
Tube 13 Pass A	756	767	760	767	757
Tube 13 Pass B	750	740	741	746	741
Tube 15 Pass A	726	729	731	734	728
Tube 15 Pass B	714	700	705	705	703

UNIT OPERATING DATA - CONTINUED

Test No. 2 Date 19 June 1981 Location TUSCO Engr. R DEROISIER

Unit No. 11413 Fuel Capacity 16250 bbl/d, 55x10⁶ Btu/h

Unit Type CRUDE HEATER Burner Type JOHN ZINK DBA-22

Test Description: Low-NO_x Using KUB Lance

UNIT OPERATING DATA

Test No. 2 Date 19 JUNE 1981 Location TOSCO Engr. K DEROSIER
 Unit No. 11H13 Fuel _____ Capacity 16250 bbl/d, 55x10⁶ Btu/h
 Unit Type CRUDE HEATER Burner Type JOHN ZINC DBA-22
 Test Description: LOW NOX

Test number	<u>2</u>	--	--	--
Time	<u>3:45</u>	--	--	--
Process Rate x 1847.7 bbl/d	<u>6.3</u>	--	--	--
Crude Tin (°F)	<u>386</u>	--	--	--
Crude Tout (°F) East	<u>638</u>	--	--	--
Crude Tout (°F) West	<u>640</u>	--	--	--
Crude Pin (psig) East	<u>140</u>	--	--	--
Crude Pin (psig) West	<u>130</u>	--	--	--
Crude Pout (psig)	<u>34.5</u>	--	--	--
Reabsorber gas in $\times 10^3$ scf/d)	<u>2.1</u>	--	--	--
Burner #1	<u>51/80</u>	--	--	--
Burner #2	<u>43/81</u>	--	--	--
Burner #3 Oil P/Steam P	<u>--</u>	--	--	--
Burner #4 (psig)	<u>49/77</u>	--	--	--
Burner #5	<u>48/77</u>	--	--	--
Burner #6	<u>--</u>	--	--	--
P to Burner #1	<u>4.2</u>	--	--	--
P to Burner #2	<u>5.0</u>	--	--	--
P to Burner #3 Gas P	<u>2.0</u>	--	--	--
P to Burner #4 (psig)	<u>3.0</u>	--	--	--
P to Burner #5	<u>6.6</u>	--	--	--
P to Burner #6	<u>4.0</u>	--	--	--
Air Registers (%open)	<u>--</u>	--	--	--
Crude API Gravity	<u>23.0</u>	--	--	--
Flame Observations	<u>--</u>	--	--	--
Tube 13 Pass A	<u>757</u>	--	--	--
Tube 13 Pass B	<u>738</u>	--	--	--
Tube 15 Pass A	<u>723</u>	--	--	--
Tube 15 Pass B	<u>700</u>	--	--	--

UNIT OPERATING DATA - CONTINUED

Test No. 2 Date 19 JUNE 1981 Location TOSCO Engr. R DEROISIER
Unit No. 11113 Fuel Capacity 16250 bbl/d, 55x10⁶ Btu/h
Unit Type CRUDE HEATER Burner Type JOHN ZINK DRA-22
Test Description: Low NO_x

Note --

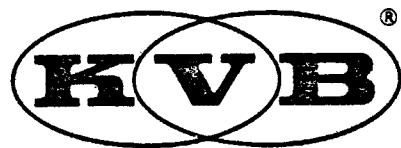
1. Gas flowmeter dropped from 3.3 to 2.1 (x205856 scfd) when cofiring with oil commenced.
2. During previous testing, KVB found the gas flowmeter to be reading high. A correction factor of 0.83 should thus be applied to all readings.

SECTION 4
SAMPLING DATA SHEETS

- 4.1 KVB Continuous Emissions Monitoring Report
- 4.2 Operating Data Tables for EPA Method 5
- 4.3 Operating Data Tables for Controlled Condensation Train
- 4.4 Operating Data Tables for SASS

4.1 KVB CONTINUOUS EMISSIONS MONITORING REPORT

**NO_x EMISSIONS ASSESSMENT:
GASEOUS EMISSIONS FROM A
REFINERY PROCESS HEATER IN
BASELINE AND LOW-NO_x
CONFIGURATIONS**



KVB11 47800-1284

**PREPARED FOR:
ACUREX CORPORATION
MOUNTAIN VIEW, CALIFORNIA**

**CONTRACT NO.
RB59313A**

**PREPARED BY:
R.J. TIDONA
KVB, INC.
RESEARCH AND ANALYSES DIV.
JULY 1981**

ABSTRACT

Gaseous emissions were measured on a natural draft refinery process heater in support of NO_x emissions assessment testing by Acurex Corporation. Species measured continuously by KVB were NO, CO, CO₂, O₂, and SO₂. Two test configurations, baseline and low-NO_x, were tested at a process rate of 77.11 m³/h (11,640 bbl/d) of crude feed. A mixture of refinery gas and No. 6 oil was fired for both tests. The low-NO_x configuration was obtained by staging approximately one-third of the total combustion air. The staged air was introduced through twenty-four lances (four per burner) at a height of 1.2m (4 ft) above the fuel injection plane. The average baseline NO emission was 172 ppm, dry at 3 percent O₂. At the low-NO_x conditions the average NO emissions dropped to 118 ppm, dry at 3 percent O₂ for a reduction of 31.4 percent. This result was in accordance with expectations based on previous tests taking into account differences in the relative proportions of No. 6 oil and refinery gas fired in the present tests. Emissions of the other continuously measured gaseous species at low-NO_x conditions were largely unchanged from their baseline levels.

CONTENTS

<u>Section</u>	<u>Page</u>
ABSTRACT	4-4
1.0 DESCRIPTION OF TESTS	4-6
2.0 GASEOUS EMISSIONS TEST METHODS AND INSTRUMENTATION	4-8
3.0 PRESENT TEST RESULTS	4-11
4.0 CONCLUSIONS	4-14
APPENDICES:	
A. GASEOUS EMISSIONS MEASUREMENTS TAKEN DURING SASS TESTS AT REFINERY PROCESS HEATER	4-15

SECTION 1.0

DESCRIPTION OF TESTS

KVB participated as a subcontractor to Acurex Corporation in the NO_x Emissions Assessment program (Contract No. RB59313A) at a refinery process heater from June 15, 1981 through June 19, 1981. During the test period KVB was responsible for continuously measuring NO_x, CO, CO₂, O₂, and SO₂ gaseous emissions. The emissions were measured on a natural draft refinery process heater firing a mixture of approximately one-third No. 6 oil (by heat input) and two-thirds refinery gas. The heater was equipped with staged air lances. One baseline test was conducted without staged combustion air and a second low-NO_x test was run with maximum staging at about the same or a slightly lower level of excess oxygen. Concurrent SASS, EPA Method 5, and wet chemical SO_x tests were run by Acurex Corporation.

Originally, plans had been made to operate the heater with a 50/50 mixture of gas and oil, however, the plant was unable to flare enough fuel gas to allow that much fuel oil to be burned. This situation occurs frequently during the summer when process offgases are more abundant. In addition, two oil guns were plugged during the tests. Due to scheduling problems with the plant maintenance department, the guns could not be cleaned in time to allow both SASS tests to be completed in the allotted time. It was nevertheless felt that the tests should be conducted and that appropriate allowances for the effect of oil/gas ratio on emissions could be made.

It is also important to note that the excess O₂ level (~4 percent) maintained in the low-NO_x test was considered to be the lowest continuous operating level by the plant, although previous tests by KVB had shown the system to be capable of operating over a short-term at 2 percent O₂. Problems in maintaining adequate combustion air flow had occurred during a 30-day test conducted on the same heater previously firing 100 percent refinery gas when 2 percent O₂ was the target operating condition. After an incident in which the unit nearly went down because of air starvation, the plant decided to

maintain 4 percent O₂ as the minimum operating level and this decision was carried over to the present tests with the oil/gas fuel combination. Thus, the NO_x reduction in the present low-NO_x test was due primarily to the staged air system. Lowered excess air was essentially not a factor in the NO_x reduction as it had been in some past tests.

SECTION 2.0

GASEOUS EMISSIONS TEST METHODS AND INSTRUMENTATION

All emission measurement instrumentation was carried in an 8 x 42 ft mobile laboratory trailer. The gaseous species measurements were made with analyzers located in the trailer.

The emission measurement instrumentation used was the following:

TABLE 1. EMISSION MEASUREMENT INSTRUMENTATION

Species	Manufacturer	Measurement Method	Model No
Carbon Monoxide	Beckman Instruments	IR Spectrometer	865
Oxygen	Teledyne	Polarographic	326A
Carbon Dioxide	Beckman Instruments	IR Spectrometer	864
Nitrogen Oxides	Thermo Electron Co.	Chemiluminescent	10A
Sulfur Dioxide	DuPont Instruments	UV Spectrometer	400

2.1 GAS SAMPLING AND CONDITIONING SYSTEM

A flow schematic of the flue gas sampling and analyzing system is shown in Figure 1. The sampling system uses one of three double-headed positive-displacement diaphragm pumps to continuously draw flue gas from the stack into the laboratory. The sample pumps pull from up to six unheated sample lines. Selector valves allow composites of up to six points to be sampled at one time. The probes are connected to the sample pumps with 0.95 cm (3/8") or 0.64 cm (1/4") nylon line. The positive displacement diaphragm sample pumps provide unheated sample gas to the refrigerated condenser (to reduce the dew point to 35°F), a rotameter with flow control valve, and to the O₂, NO, CO, and CO₂ instrumentation. Flow to the individual analyzers is measured and controlled with rotameters and flow control valves. Excess sample is vented to the atmosphere.

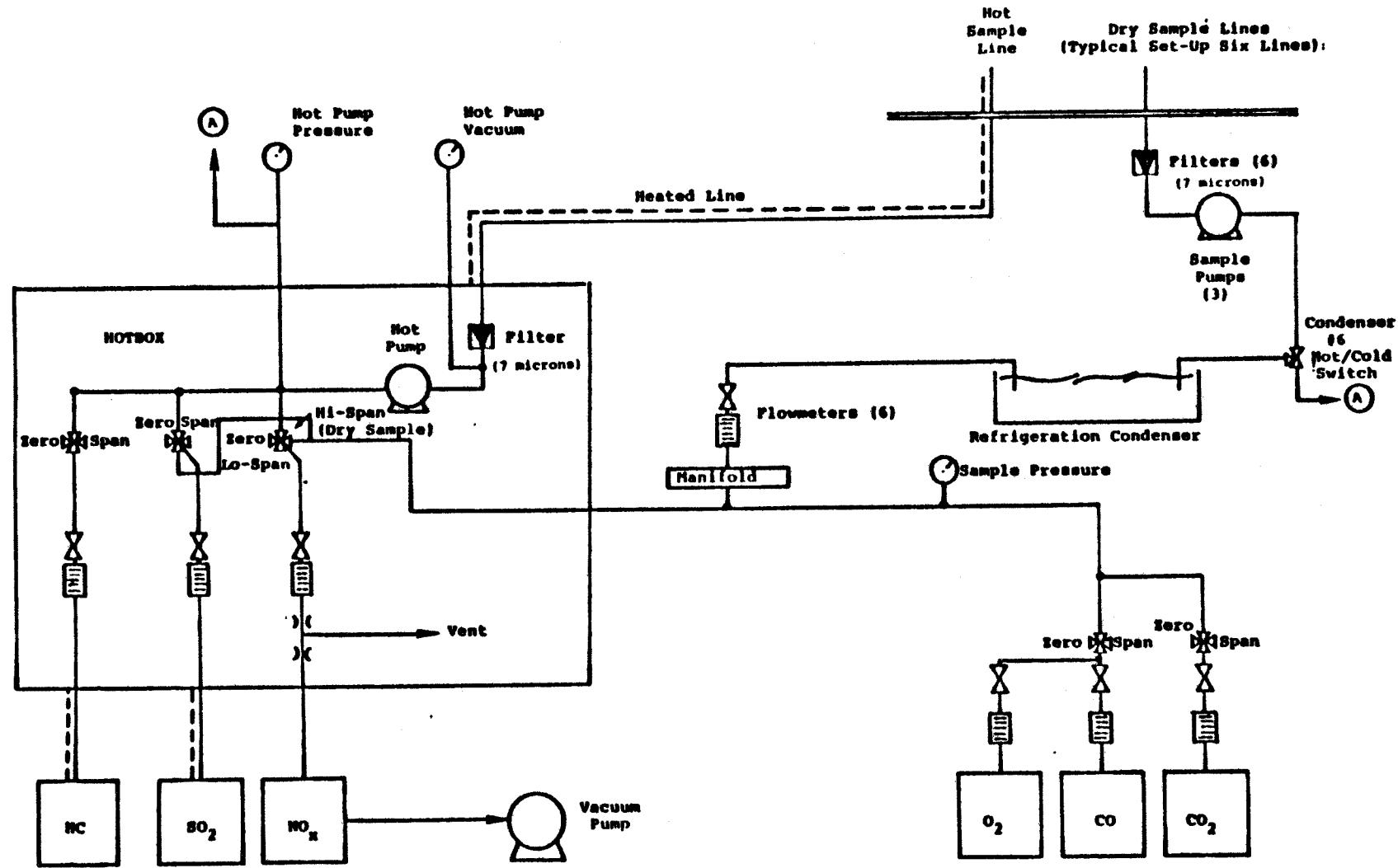


Figure 1. Flue gas sampling and analyzing system.

To obtain a representative sample for the analysis of NO_2 , SO_2 and hydrocarbons, the sample must be kept above its dew point, since heavy hydrocarbons may be condensable, and SO_2 and NO_2 are quite soluble in water. For this reason, a separate electrically heated sample line is used to bring the sample into the laboratory for analysis. The sample line is 0.95 cm (3/8-inch) Teflon line, electrically traced and thermally insulated to maintain a sample temperature of up to 400°F. A heated diaphragm pump provides hot sample gas to the hydrocarbon, SO_2 and NO_x analyzers and cold, dried gas to the other continuous analyzers via the condenser described above.

The laboratory trailer is equipped with the analytical instruments shown in Table 1 to continuously measure concentrations of NO , NO_2 , CO , CO_2 , O_2 , SO_2 , and hydrocarbons. All of the continuous monitoring instruments and sample handling system are mounted in the self-contained mobile laboratory. The instruments themselves are shock mounted on a metal console panel. The sample flow control measurement, and selection, together with instrument calibration are all performed from the console face. Three-pen recorders provide a continuous permanent record of the data taken. The sample gas is delivered to the analyzers at the proper condition and flow rate through the sampling and conditioning system described previously. A Monitor Laboratories Model 9300 Data Logger is normally used to record and average data from the analyzers and can be coupled with a Techtran Model 815 Datacassette recorder to provide digital storage of the data.

SECTION 3.0

PRESENT TEST RESULTS

The test unit was first run at a baseline operating condition at a process rate of 77.11 m³/h (11,640 bbl/d) of crude throughput. A sketch of the unit is shown in Figure 2. The secondary air registers were approximately 40 percent open and the excess oxygen at the stack was maintained at approximately four percent. For this baseline test the ratio of oil/gas by heat input was 37/63.

The gaseous emission measurements were taken by means of a single probe and heated sample line from the stack approximately five feet above the outlet of the transition section for both the baseline and the low-NO_x test which followed. Dry measurements of CO, CO₂, NO, and O₂ were made while a wet measurement was taken for SO₂.

The low-NO_x configuration on this process heater was achieved by means of staged air lances inserted through the heater floor. Roughly one-third of the total combustion air was introduced through the lances. This was the maximum staging capability of the system. An excess oxygen level of approximately 3-4 percent was maintained and the secondary air registers were adjusted to ten percent open as in previous tests conducted by KVB. The process rate was again 77.11 m³/hr of crude oil, but the oil/gas ratio changed slightly to 31/69.

(All numbers given for the operating parameters are approximate daily averages.)

The table below summarizes the emissions measured and the standard deviation in the emission measurements along with the range of values and number of measurements taken for both the baseline and the low-NO_x tests. SO₂ measurements are reported as wet values at measured excess O₂. The values for NO emissions are reported as dry at measured excess O₂. The average NO emissions were corrected to 3 percent O₂ using that average O₂ concentration

NATURAL DRAFT REFINERY PROCESS HEATER

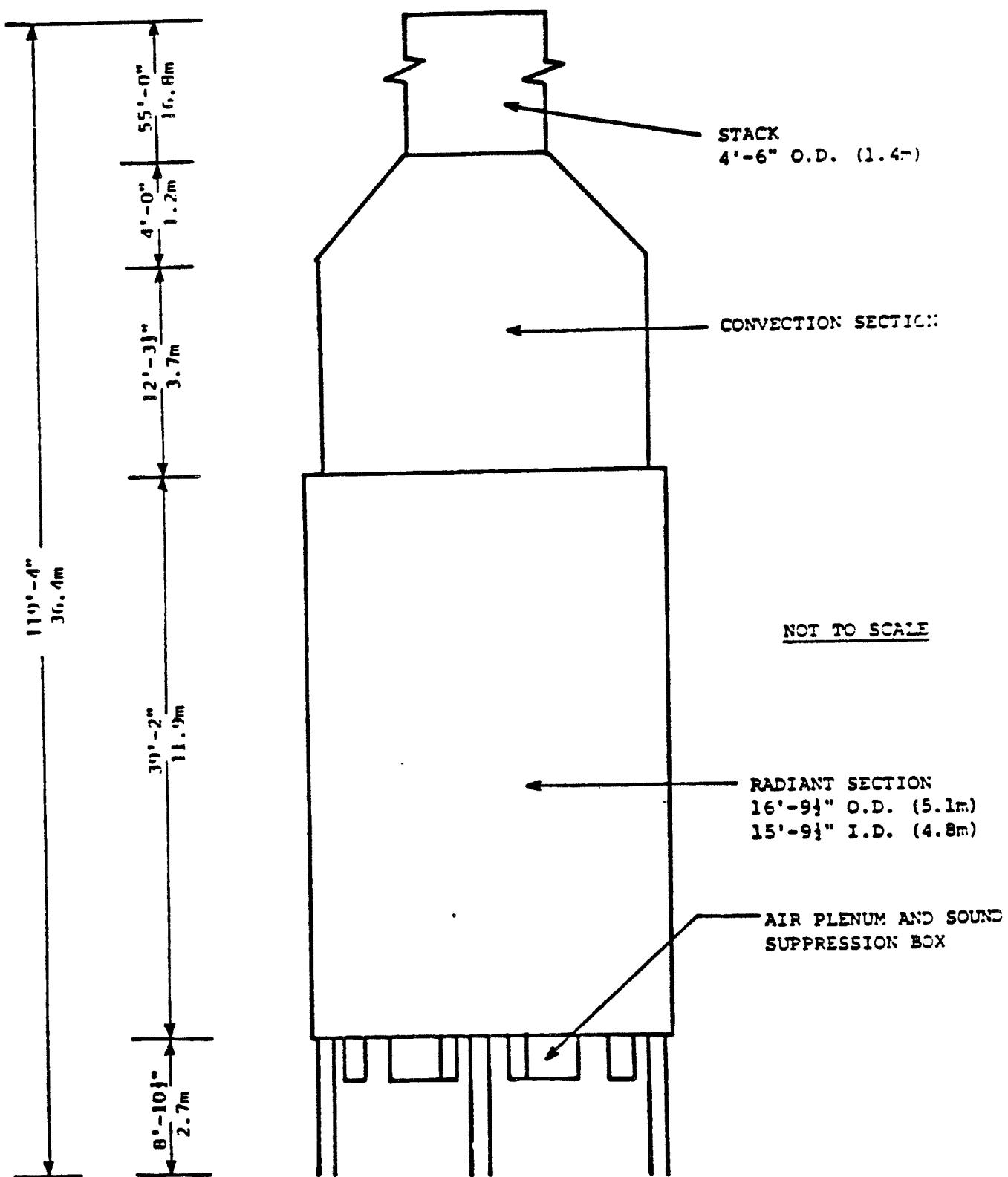


Figure 2. Sketch of the process heater tested.

both tests. The gaseous emission measurements were 15 minute averages as determined by manually interpreting the strip chart recordings for each analyzer. This procedure was made necessary by the failure of the electronic data logger just prior to the start of the tests.

TABLE 2. SUMMARY OF GASEOUS EMISSIONS DATA FROM
REFINERY PROCESS HEATER SASS TESTS

Species	<u>Baseline Test</u>				<u>Low NO_x Test</u>			
	\bar{C}	s	C_{min}/C_{max}	n	\bar{C}	s	C_{min}/C_{max}	n
O ₂ (% dry)	4.0	0.20	3.3/4.3	37	3.3	0.27	3.1/4.2	20
CO ₂ (% dry)	12.1	0.87	10.5/13.6	33	11.7	0.32	11.3/12.4	17
CO (ppm, dry)	3.8	2.9	0/10	38	7.3	3.4	0/10	20
SO ₂ (ppm, wet)	151	11.6	120/170	21	148	11.4	125/170	20
NO (ppm, dry)	162	6.0	152/174	37	116	3.8	108/125	20
NO (ppm, dry at 3% O ₂)	172	--	156/186	--	118	--	109/134	--

$$\text{\%NO reduction in low-NO}_x \text{ configuration} = \frac{172 - 118}{172} \times 100 = 31.4\%$$

The complete set of gaseous emission measurements is given in the Appendix. No significant NO₂ was measured during either test.

SECTION 4.0

CONCLUSIONS

The data show that an average NO emission reduction of 31.4 percent from the baseline emission of 172 ppm, dry at three percent O₂, was obtained in the low-NO_x configuration. The CO emissions remained very low in both tests. The high standard deviations reflect the fact that the CO levels were at the lower limits of detectability of the CO analyzer being used. SO₂ emissions were approximately the same in both baseline and low-NO_x configurations.

The percent reduction in NO emissions due to the staged air system was approximately the same as the reduction observed in previous tests when firing a 50/50 mixture of oil and gas fuels under roughly the same operating conditions. In those previous tests the heater process rate was 78.3 m³/h (11,800 bbl/d) and the overall excess O₂ was about 4 percent. In those tests, however, the baseline NO emission was 207 ppm, dry at three percent O₂.

The difference in baseline NO emissions between the present tests and the past tests is due to the difference in oil/gas ratio. From previous tests, it is known that the thermal NO_x alone at baseline conditions is about 125 ppm, dry at 3 percent oxygen. The baseline level of 172 ppm in the present tests for a 37/63 oil/gas mixture represents a 47 ppm increment above this emission level. One may predict an emission level for the 50/50 oil/gas mixture as follows (assuming a constant fuel nitrogen conversion efficiency):

$$\left(\frac{50}{37} \times 47\right) + 125 = 189 \text{ ppm}$$

This value is within 10 percent of the observed value of 207 ppm.

APPENDIX A

**GASEOUS EMISSIONS MEASUREMENTS TAKEN
DURING SASS TESTS AT REFINERY PROCESS HEATER**

KVB11-47800-1284

4-I5

BASELINE
SASS TEST I - 6-18-81
FROHOFER



$\text{SO}_2 \text{ ppm}$

Date	Time	NO ppm	O ₂ Percent	CO ₂ Percent	CO ppm	Load x 1000	Comment Codes
	0615						
	0630						
	0645						
	0700						
	0715						
	0730						
	0745						
	0800						
	0815						
	0830						
	0845						
	0900						
	0915						
	0930						
	0945						
	1000						
	1015						
	1030						
	1045						
	1100						
	1115						
	1130						
	1145	START OF SASS TEST		12.00			
	1200	162	4.2	13.6	10		148

Page 2

4-16

A Research-Cottrell Company

KVB 6017
9/80

BASELINE
SASS TEST I - 6-18-81
FROHOF



SO₂ ppm

Date	Time	NO ppm	O ₂ Percent	CO ₂ Percent	CO ppm	Load x 1000	Comment Codes
	1215	163	4.2	12.9	5		144
	1230	164	4.2	13.0	5		130
	1245	159	4.0	12.5	5		120
	1300	162	4.1	12.5	5		-0.5
	1315	170	4.2	12.5	5		005
	1330	169	4.3	12.8	5		005
	1345	167	4.1	13.3	5		005
	1400	168	4.1	13.6	5		005
	1415	170	4.2	13.3	5		005
	1430	172	4.2	13.3	5		005
	1445	174	4.2	13.3	5		005
	1500	173	4.3	12.8	5		005
	1515	Calibrate	4.1	12.8	5		005
	1530	162	Calibrate	12.5	5		005
	1545	162	4.0	Calibrate	5		Calibrate
	1600	163	3.9	11.7	5		Cal
	1615	163	3.9	11.7	5		Cal
	1630	163	4.0	11.7	5		Cal
	1645	164	4.0	11.7	5		Cal
	1700	164	3.9	11.6	5		Cal
	1715	157	Calibrate	11.3	Calibrate		Cal
	1730	160	3.8	Calibrate	5		170
	1745	Calibrate	3.9	11.3	5		160
	1800	170	4.2	10.5	10		170

Page 3

BASELINE
SASS TEST I 6-18-81
FROHOFF



$\text{SO}_2 \text{ ppm}$

Date	Time	NO ppm	O_2 Percent	CO_2 Percent	CO ppm	Load $\times 1000$	Comment Codes
6-18	1815	170	4.2	11.0	5		150
	1830	153	3.3	11.5	0		140
	1845	156	3.7	11.0	0		140
	1900	Calibrate	Calibrate	Calibrate	Cal		Cal
	1915	156	3.8	12.0	10		155
	1930	160	3.9	11.7	0		155
	1945	158	3.8	11.3	0		155
	2000	159	3.9	11.3 / Cal	6		155
	2015	157	3.8	11.7	0		155
	2030	158	3.8	11.7	0		155
	2045	158	4.0	11.0	0		155
	2100	157	4.1	—	0		155
	2115	155	4.1	Cal	0		155
	2130	153	3.9	—	0		155
	2145	152	4.0	—	0		155
	2200	—	END SASS TEST I				
	2215						
	2230						
	2245						
	2300						
	2315						
	2330						
	2345						
	2400						

SASS TEST II
Low NO_x - 6-19-81
FROHOFER - HTR II H13



Date	Time	NO ppm	O ₂ Percent	CO ₂ Percent	CO ppm	Load x 1000	Comment Codes
	0615						
	0630						
	0645						
	0700						
	0715						
	0730						
	0745						
	0800						
	0815						
	0830						
	0845						
	0900						
	0915						
	0930						
	0945		SASS TEST # II STARTED 10:00				
	1000						
	1015		—	Calibrate	—		
	1030		—	Calibrate	—		
	1045	125	4.2	11.3	0		145
	1100	120	3.9	11.3	0		145
	1115	116	3.5	11.7	5		145
	1130	114	3.4	11.7	5		145
	1145	117	3.4	11.7	10		140
	1150	118	3.4	11.7	10		130

NOTE* DATA POINTS ARE AVERAGES OF
PREVIOUS 15 MINUTES OF DATA

4-19

A Research-Cottrell Company

Page 2

KVB 6017
9/80

Sass TEST II
Low NO_x - 6-19-81
FRONOFF HTR 11413



50 ppm

Date	Time	NO ppm	O ₂ Percent	CO ₂ Percent	CO ppm	Load x 1000	Comment Codes
6-19	1215	115	3.2	11.9	5		125
	1230			CALIBRATE			
	1245	110	3.1	12.4	10		160
	1300	108	3.1	12.4	10		160
	1315	112	3.2	11.9	10		160
	1330	112	3.2	11.9	10		150
	1345	117	3.2	11.7	5		150
	1400	119	3.3	11.3	5		140
	1415	117	3.2	11.5	5		140
	1430	118	3.3	11.9	5		135
	1445			CALIBRATE			
	1500	117	3.2	—	10		170
	1515	118	3.2	—	10		160
	1530	118	3.2	Calibrate	10		160
	1545	118	3.2	11.7	10		150
	1600	119	3.2	11.7	10		150
	1615						
	1630						
	1645						
	1700						
	1715						
	1730						
	1745						
	1800						

NOTE * DATA POINTS ARE AVERAGES OF
PREVIOUS 15 MINUTES OF DATA

4-20

Page 3

4.2 OPERATING DATA FOR EPA METHOD 5

TRAVERSE POINT LOCATION FOR CIRCULAR DUCTS

PLANT TOSCO
 DATE 6/18/81
 SAMPLING LOCATION stack
 INSIDE OF FAR WALL TO
 OUTSIDE OF NIPPLE, (DISTANCE A) 58.5"
 INSIDE OF NEAR WALL TO
 OUTSIDE OF NIPPLE, (DISTANCE B) 12"
 STACK I.D., (DISTANCE A - DISTANCE B) 49" 46.5"
 NEAREST UPSTREAM DISTURBANCE Stack exit 72"
 NEAREST DOWNSTREAM DISTURBANCE top of heater 4.96"
 CALCULATOR Steiner

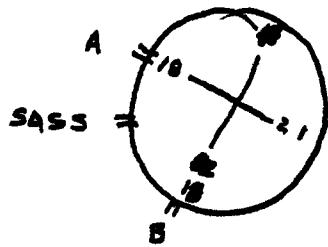
* adjust to 1" from stack wall

SCHEMATIC OF SAMPLING LOCATION

TRAVERSE POINT NUMBER	FRACTION OF STACK I.D.	STACK I.D.	PRODUCT OF COLUMNS 2 AND 3 (TO NEAREST 1/8 INCH)	DISTANCE B	TRAVERSE POINT LOCATION FROM OUTSIDE OF NIPPLE (SUM OF COLUMNS 4 & 5)
1	0.014	49	46.5 0.69*	12	13* 13.00
2	0.044	49	2.16 2.05	12	14.16 14.05
3	0.075	49	3.68 3.49	12	15.68 15.49
4	0.109	49	3.34 5.07	12	17.34 17.07
5	0.146	49	7.15 6.79	12	19.15 18.79
6	0.188	49	9.21 8.74	12	21.21 20.74
7	0.236	49	11.56 10.97	12	23.56 22.97
8	0.296	49	14.50 13.76	12	26.50 25.76
9	0.382	49	18.71 17.76	12	30.71 29.76
10	0.618	49	30.28 28.73	12	42.28 40.73
11	0.704	49	34.50 32.74	12	46.50 44.74
12	0.764	49	37.43 35.53	12	49.43 47.53
13	0.812	49	39.79 37.76	12	51.79 49.76
14	0.854	49	41.85 39.71	12	53.85 51.71
15	0.891	49	43.66 41.43	12	55.66 53.43
16	0.925	49	45.33 43.01	12	57.33 55.01
17	0.956	49	46.84 44.45	12	58.84 56.45
18	0.986	49	48.31* 46.85*	12	60* 57.00

PRELIMINARY VELOCITY TRAVERSE

PLANT TOSCO
 DATE 6/18/1
 LOCATION Stack
 STACK I.D. 46.5"
 BAROMETRIC PRESSURE, in. Hg 29.42
 STACK GAUGE PRESSURE, in. H₂O - 0.31
 OPERATORS Steiner / Sniffen



SCHEMATIC OF TRAVERSE POINT LAYOUT

TRAVERSE POINT NUMBER	VELOCITY HEAD (Δp_s), in.H ₂ O	STACK TEMPERATURE (T_s), °F
A-1	0.10	708
2	0.10	708
3	0.10	704
4	0.10	704
5	0.10	703
6	0.09	705
7	0.09	706
8	0.09	706
9	0.09	706
10	0.08	704
11	0.08	706
12	0.08	705
13	0.07	704
14	0.07	702
15	0.06	694
16	0.06	235*
17	0.04	191*
18	0.04	191*
A $\bar{p}_{\bar{v}_2} 1-15 = 704.3^{\circ}\text{F}$		
AVERAGE	0.08	708°F

EPA (Dur) 233
4/72

*NOT USED

TRAVERSE POINT NUMBER	VELOCITY HEAD (Δp_s), in.H ₂ O	STACK TEMPERATURE (T_s), °F
B-1	0.09	692
2	0.09	692
3	0.09	694
4	0.09	701
5	0.09	700
6	0.08	701
7	0.08	708
8	0.08	708
9	0.07	708
10	0.08	708
11	0.08	709
12	0.08	704
13	0.08	704
14	0.08	704
15	0.08	702
16	0.07	68*
17	0.06	570
18	0.06	570
B $\bar{p}_{\bar{v}_2} 1-15 = 680.61^{\circ}\text{F}$		
AVERAGE	.079	701.2

DRY MOLECULAR WEIGHT DETERMINATION

PLANT TOSCO
 DATE 6/18/81
 SAMPLING TIME (24-hr CLOCK) 8:07 am
 SAMPLING LOCATION stack
 SAMPLE TYPE (BAG, INTEGRATED, CONTINUOUS GRAB) Fyrite, grab
 ANALYTICAL METHOD
 AMBIENT TEMPERATURE 75°F
 OPERATOR Holm

COMMENTS:

RUN GAS	1		2		3		AVERAGE NET VOLUME	MULTIPLIER	MOLECULAR WEIGHT OF STACK GAS (DRY BASIS) M_d , lb/lb-mole
	ACTUAL READING	NET	ACTUAL READING	NET	ACTUAL READING	NET			
CO ₂		12		12		11.5	12.83	44/100	5.21
O ₂ (NET IS ACTUAL O ₂ READING MINUS ACTUAL CO ₂ READING)		4.5		4.5		5.0	4.66	32/100	1.49
CO (NET IS ACTUAL CO READING MINUS ACTUAL O ₂ READING)							0	28/100	\$
N ₂ (NET IS 100 MINUS ACTUAL CO READING)							83.51	28/100	23.38
								TOTAL	30.08

ISOKINETIC SAMPLING WORKSHEET

Plant TOSCOPerformed by SteinerDate 6/18/81Sample Location StackTest No./Type M5 #1

$$K = \frac{782.687 (C_p)^2 (1-B_{w0})^2 P_s M_d}{K_o^2 M_s P_m}$$

where: K = Constant of fixed and assumed parameters (dimensionless)

Pitot coefficient (dimensionless)	C_p	0.84
Water vapor in the gas stream (proportion by volume)	B_{w0}	0.12
Absolute stack gas pressure (in. Hg)	P_s	29.39
Molecular weight, stack gas dry (1b/lb-mole)	M_d	30.07
Orifice coefficient (dimensionless)	K_o	0.6697
Molecular weight, stack gas wet (1b/lb-mole) $M_d(1-B_{w0}) + 18(B_{w0})$	M_s	28.62
Absolute meter pressure (in. Hg)	P_m	29.42
<u>782.687 (.84)^2 (1-.12)^2 (29.39) (30.07)</u> <u>(.6697)^2 (28.62) (29.42)</u>	K	1000.85

ISOKINETIC NOZZLE CALCULATION
AND
SAMPLING RATE CALCULATION

Plant TOSCO
Date 6/18/81
Sample Location Stack
Test No./Type M5 #1

Performed by Steiner

$$N_d = \left(\frac{\Delta H}{K T_m \Delta P} \right)^{.25}$$

where: N_d = Nozzle diameter (inches)

Average pressure differential across the orifice meter (in. H ₂ O)	ΔH	<u>1.2</u>
Temperature stack gas, average (°F)	T_s	<u>700</u>
Temperature of gas meter, average (°F)	T_m	<u>105</u>
Stack gas velocity pressure (in H ₂ O)	ΔP	<u>0.08</u>
$\left(\frac{(1.2)(700 + 460)}{(1000.85)(105 + 460)(.08)} \right)^{.25}$	N_d	<u>0.4188</u>

$$\Delta H = K (N_d)^4 \frac{T_m}{T_s} (\Delta P)$$

where: ΔH = Pressure differential across the orifice meter (in H₂O)

Nozzle diameter, actual (inches)	N_d	<u>0.382</u>
Temperature of gas meter (°F)	T_m	
Temperature of stack gas (°F)	T_s	
Stack gas velocity pressure (in H ₂ O)	ΔP	
$\left(() ()^4 \left(\frac{+ 460}{+ 460} \right) () \right)$	ΔH	
Magic number <u>1000.85 (.382)⁴</u>	$K(N_d)^4$	<u>21.31</u>

FIELD DATA

Plant TOSCO, BAKERSFIELD, CADate 6-18-81Sample Location C. RUDIE HEATERSample Type M-SRun Number 1Operator J. STIENER / J. SNIFFENAmbient Temperature 75°F - 102°FBarometric Pressure 29.42Static Pressure, (H₂O) - 0.31 in.Filter Number(s) 96-34Leak Check: Initial at 16 " Hg, .002 CFM
Final at 14.5 " Hg, .002 CFMPitot Leak Check: OK

Impinger Volumes

Initial Final Net Gain

100 486.0 286.0
100 0 _____
0 _____ _____

Silica Gel

252.0 340.2 90.2
_____ _____ _____

SASS Condensate

N/A
_____Total Volume 376.2Probe Length and Type 5' - PYREXNozzel Size & I.D. .382 SN 907Pitot Coefficient & I.D. .84Assumed Moisture 12%Molecular Weight, Dry, (M_d) 30.07Meter Box Number Meter Coefficient $K_0 = 0.6697$ α Factor 1.010 $K = 1000.85$ $K(N_d)^4 = 1000 \times (.382)^4 = 21.31$

$$\Delta H = K(N_d)^4 \left(\frac{T_m}{T_s} \right) (P)$$

4-27

Traverse Point Number	Sampling Time, min	Clock Time (24-hr) Clock	Gas Meter Reading (V_m), ft ³	Velocity Head (ΔP_s), in. H_2O	Orifice Pressure Differential (ΔH), in. H ₂ O	Temperature °F						Pump Vacuum in. Hg	Avg. \sqrt{AP}			
						Desired	Actual	Stack	Probe	Impinger	Organic Module	Gas Meter				
												In	Out			
B-1	0	4:43 PM	Init. 685.342	0.13	1.31	732	250	96				200	110	110	6	.360
B-2	5		687.825	0.13	1.31	737	238	96				274	111	111	5	.360
B-3	10		691.525	0.13	1.32	737	330	96				246	112	111	5	.360
B-4	15		694.575	0.13	1.31	741	291	95				256	112	112	5	.360
B-5	20		697.615	0.13	1.32	741	282	97				251	112	111	5	.360
B-6	25		700.685	0.13	1.32	741	282	97				255	112	112	5	.346
B-7	30		703.680	0.12	1.21	743	277	94				252	112	112	5	.346
B-8	35		706.650	0.12	1.22	741	269	93				255	112	112	5	.346
B-9	40		709.435	0.10	1.01	740	263	94				257	112	112	5	.316
			712.150	0.09	0.91	745	257	95				257	112	112	4	.300

7602/5/81/Rev 1

Comments: 5 min/point

813795

4-28

Traverse Point Number	Sampling Time, min	Gas Meter Reading (V_m), ft ³	Velocity Head (ΔP_s), in. H ₂ O	Temperature °F					Pump Vacuum in. Hg	Avg. $\sqrt{\Delta P}$			
				Orifice Pressure Differential (ΔH), in. H ₂ O		Stack	Probe	Impinger	Organic Module	Oven	Gas Meter		
				Desired	Actual						In	Out	
B-10		714.620	0.09	0.91	739	258	94		230	112	112	4	.300
11		717.275	0.09	0.91	735	263	92		244	112	112	4	.300
12		719.730	0.09	0.91	742	248	89		264	111	111	4	.300
13		722.295	0.09	0.91	739	256	88		265	110	110	4	.300
14		724.870	0.09	0.91	736	262	89		255	110	110	4	.300
15		727.420	0.09	0.91	730	263	89		257	109	109	4	.300
16		730.000	0.09	0.92	716	264	90		262	110	110	4	.300
17		732.000	0.09	0.93	703	259	89		264	111	111	4	.300
18	6:13pm	735.174	0.09	0.94	697	259	88		262	111	111	4	.300
		735.174											
A-1	6:28	738.210	0.14	1.31	745	259	91		267	113	113	5	.374
2		741.380	0.14	1.42	743	306	88		276	115	115	5	.374
3		744.575	0.14	1.42	746	310	89		286	115	115	6	.374
4		747.815	0.14	1.43	735	271	93		241	115	115	6	.374
5		750.950	0.13	1.32	745	277	94		249	114	114	6	.360
6		754.075	0.13	1.32	744	271	93		246	113	113	6	.360
7		757.180	0.13	1.31	750	271	92		259	113	113	6	.360

Run No. #1

Date 6/18/81

Sampling Location Stack

Comments:

4-29

Traverse Point Number	Sampling Time, min	Clock Time (24-hr) Clock	Gas Meter Reading (V_m), ft ³	Velocity Head (ΔP_s), in. H ₂ O	Temperature °F					Pump Vacuum in. Hg	Avg. $\sqrt{\Delta P}$					
					Orifice Pressure Differential (ΔH), in. H ₂ O			Stack	Probe	Impinger	Organic Module	Oven	Gas Meter			
					Desired	Actual							In	Out		
A-8			760.285	0.13	1.31	747	250	92				251	112	112	6	.360
9			763.190	0.11	1.11	746	256	90				253	112	111	6	.331
10			765.930	0.10	1.01	743	259	89				251	110	111	5	.316
11			768.655	0.10	1.00	746	250	85				274	110	110	4.5	.316
12			771.375	0.10	1.01	740	252	83				279	110	110	4.5	.316
13			774.215	0.11	1.11	741	248	83				262	110	110	4.5	.331
14			777.055	0.11	1.11	742	253	82				271	110	109	5.0	.331
15			780.900	0.11	1.11	738	259	82				270	110	109	5.0	.331
16			782.675	0.10	1.04	705	258	80				274	110	109	5.0	.316
17			785.630	0.10	1.36	430	250	81				259	110	109	6.0	.316
18	7:58 pm		789.326	0.10	1.99	250	248	80				269	110	109	6.0	.316
180			103.984				1.16	714.7				111.5	111.2			.332
111.35																

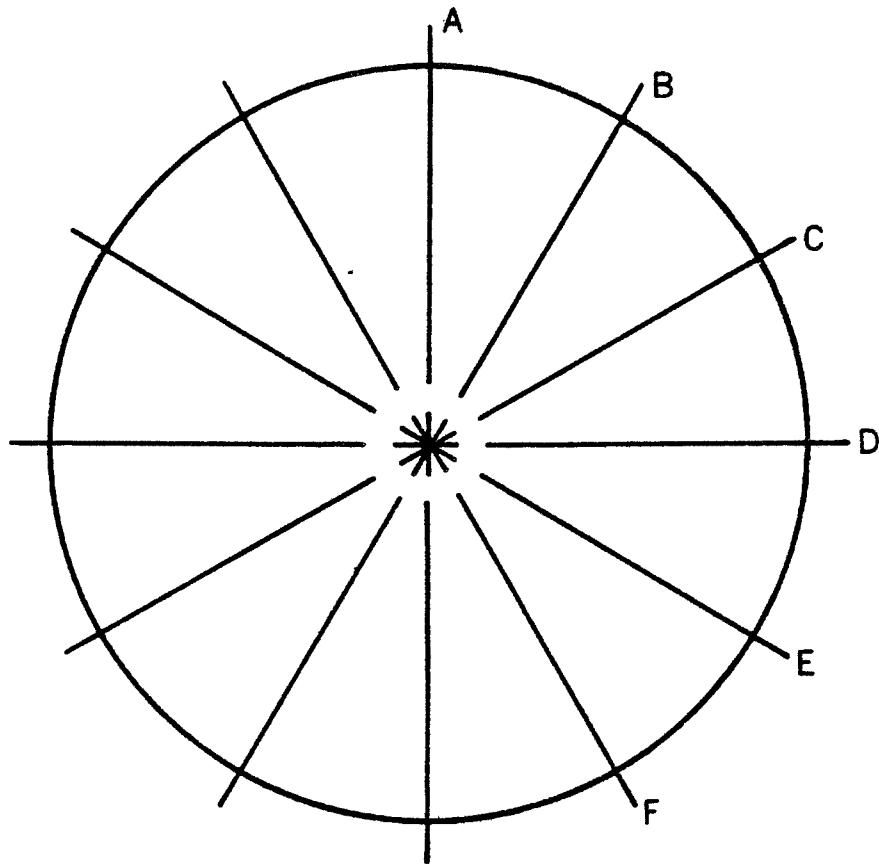
Run No. #1

Date 6/18/81

Sampling Location stack

Comments:

NOZZLE MEASUREMENT



DIAMETER
DIMENSION

A .380

B .383

C _____

D .384

E .382

F _____

AVG. .382

NOZZLE SERIAL

907

DATE
5-7-81

RECORDED BY
SFF

4-32

Traverse Point Number	Sampling Time, min (24-hr) Clock	Gas Meter Reading (V_m), ft ³	Velocity Head (ΔP_s), in. H ₂ O	Orifice Pressure Differential (ΔH), in. H ₂ O	Temperature °F					Gas Meter	Pump Vacuum in. Hg	Avg. $\sqrt{\Delta P}$				
					Desired	Actual	Stack	Probe	Impinger	Organic Module	Oven	In	Out			
A-10		818.245	0.09	0.88	728	250	88			254	110	107	2	.300		
11		820.755	0.09	0.88	736	254	88			267	110	107	2	.300		
12		823.285	0.09	0.88	732	253	82			268	111	107	2	.300		
13		825.670	0.08	0.89	728	254	85			240	112	110	2.	.282		
14		828.110	0.09	0.89	731	258	84			261	112	110	2	.300		
15		830.665	0.09	0.90	714	262	85			254	112	110	2	.300		
16		833.255	0.09	0.93	678	262	87			278	112	110	2	.300		
17		836.625	0.09	0.96	641	263	89			267	113	111	2	.300		
18*	12:06	840.175	0.07	0.75	641	263	89			267	113	111	2	.264		
		840.175														
B 1	12:20	843.042	0.12	1.19	728	258	94			278	118	116	2	.346		
2		845.910	0.12	1.20	731	302	93			280	118	117	2	.346		
3		848.880	0.12	1.20	727	285	96			271	118	117	2	.346		
4		851.820	0.11	1.09	736	258	94			280	118	117	2	.331		
5		854.675	0.11	1.10	734	286	93			278	118	116	2	.331		
6		857.520	0.10	1.00	729	271	92			266	118	118	2	.316		
7		860.140	0.10	1.00	735	261	92			272	117	115	2	.316		

Run No. M5 #2Date 6/19/81Sampling Location Stack

Comments: * Sampled A-18 for 9 mins instead of 5

Traverse Point Number	Sampling Time, min	Clock Time (24-hr) Clock	Gas Meter Reading (Vm), ft ³	Velocity Head (ΔPs), in. H ₂ O	Orifice Pressure Differential (ΔH), in. H ₂ O	Temperature °F						Pump Vacuum in. Hg	Avg. ΔP		
						Desired	Actual	Stack	Probe	Impinger	Organic Module	Oven	In	Out	
A-B			862.925	0.08	0.79	736	267	94			271	117	116	2	.282
9			865.375	0.08	0.80	731	259	96			269	116	115	2	.282
10			867.790	0.08	0.80	731	254	95			254	117	116	2	.282
11			870.210	0.08	0.80	729	267	94			259	116	115	2	.282
12			872.650	0.08	0.80	722	277	98			252	117	115	2	.282
13			874.040	0.08	0.80	722	258	95			248	117	117	2	.282
14			877.490	0.08	0.80	721	252	92			250	116	115	2	.282
15			879.900	0.07	0.80	575	261	94			242	115	114	2	.264
16			882.325	0.06	0.79	440	261	94			240	115	114	2	.264
17			884.800	0.06	0.79	440	252	92			244	116	116	2	.264
18			887.149	0.06	0.77	440	-	--			-	116	116	2	.264
180			97.64		.91	676.6						113.11	111.81	111.11	

11.2.21

= .307

Run No. 11.2.21Date 1.1.1981Sampling Location 117-1-C

Comments:

4.3 OPERATING DATA FOR CONTROLLED CONDENSATION TRAIN

CONTROLLED CONDENSATION SYSTEM (CCS)
FIELD CHECKPOINT SHEET

Checkpoint	Initials		Remarks
	Supervisor	QA Inspector	
LABORATORY PREPARATION			
• Inspect and clean CCC. Both filter holder and CCC are cleaned with hot chromic acid solution and D.I. H ₂ O.		✓	
• Rinse with acetone and air dry CCC.		✓	
• Place Tissuequartz filter in filter housing.		✓	
• Check seal between end of joint and filter.		✓	
• Do not use grease on joints.		✓	
• Inspect and clean all glass joints.		✓	
SITE SETUP			
• Rinse the inside of probe prior to run.		✓	
• Rinse probe with acetone until rinse solution is clear.		✓	
• Perform leak test.		✓	.0015 @ 21" Hg
• Leak rate must be less than 80 ml/min (0.003 cfm).		✓	
• Thermocouple leads attached to probe and filter.		✓	
• CCC water bath held at 60°C (140°F) <u>+10°C</u> .		✓	
• Leak test train.		✓	
• Probe temperature maintained at 316°C (600°F) <u>+17°C</u> .		✓	
• Gas temperature out of filter holder held at 228°C (550°F).		✓	
• Fresh solutions placed in impingers.		✓	
• Fresh absorbent replaced in final impinger.		✓	
• Adjust flowrate in system to 8 lpm.		✓	

CONTROLLED CONDENSATION SYSTEM (CCS)
FIELD CHECKPOINT SHEET -- Continued

Checkpoint	Initials		Remarks
	Supervisor	QA Inspector	
SAMPLING RUN		✓	
• Turn vacuum pump on just before inserting probe in stack.		✓	
• Check seal between probe and port to prevent any outside air from entering stack.		✓	
• Run test for 1 hour or until coils are frosted to 1/2 or 2/3 their length.		✓	
• After run, cap both ends of probe and lay in horizontal position.		✓	
• Rinse the CCC coils into the modified Erlenmeyer flask with a maximum of 40 ml D.I. H ₂ O.		✓	
• Was any of the solution lost (\checkmark ml estimated)?		✓	
• After probe has cooled, it is rinsed with a maximum of 40 ml D.I. H ₂ O into a 25-ml Erlenmeyer flask.		✓	
- Was any solution lost (\checkmark ml estimated)?		✓	
- Clean support equipment prior to next run.		✓	
- Save filter for titration.		✓	

Comments:

CONTROLLED CONDENSATION SYSTEM (CCS)
FIELD DATA SHEET

Plant TOSCO, BAKERSFIELD, CA
 Date 6-18-81
 Sample Location CRUDE HEATED
 Run No. 1 / CCS
 Operator B.C. DiaRos
 Ambient Temperature 104 °F
 Barometric Pressure 29.42 @ 1530
 Meter Box Number 684
 Meter Orifice Coefficient .7024
 Meter α Factor .998

Clock Time (24-hr) clock Sampling Time, Δ min	Gas Meter Reading (V _m), ft ³ Init. 240.680	Temperature (°F)							
		Stack	Probe	Filter		°C	Recirc Water	Dry Gas Meter	
				Skin 1	Out 2			Exit Coil 4	In
20	1600	845.510		543	1233	685	60	129	116
30	1610	848.010		560	1213	657	60	127	115
40	1620	850.900		561	1204	649	60	128	116
50	1630	853.800		564	1207	648	60	127	116
60	1640	856.500		569	1205	648	60	129	118
70	1650	859.940		570	1207	640	60	125	118
Average 70 min	19.260	714.7	561.2	1211.5	653.5	60	127.5	116.3	

↑ From M-5 / Run No 1 Data (average value)

TOSCO

CCS RUN NO 2 6-19-81

CONTROLLED CONDENSATION SYSTEM (CCS)
FIELD CHECKPOINT SHEET

Checkpoint	Initials		Remarks
	Supervisor	QA Inspector	
LABORATORY PREPARATION		✓	
• Inspect and clean CCC. Both filter holder and CCC are cleaned with hot chromic acid solution and D.I. H ₂ O.		✓	
• Rinse with acetone and air dry CCC.		✓	
• Place Tissuequartz filter in filter housing.		✓	
• Check seal between end of joint and filter.		✓	
• Do not use grease on joints.		✓	
• Inspect and clean all glass joints.		✓	
SITE SETUP		✓	
• Rinse the inside of probe prior to run.		✓	
• Rinse probe with acetone until rinse solution is clear.		✓	
• Perform leak test.		✓	.0018 dm ³ /s 18" Hg
• Leak rate must be less than 80 ml/min (0.003 cfm).		✓	
• Thermocouple leads attached to probe and filter.		✓	
• CCC water bath held at 60°C (140°F) <u>+10°C</u> .		✓	
• Leak test train.		✓	
• Probe temperature maintained at 316°C (600°F) <u>+17°C</u> .		✓	
• Gas temperature out of filter holder held at 228°C (550°F).		✓	
• Fresh solutions placed in impingers.		✓	
• Fresh absorbent replaced in final impinger.		✓	
• Adjust flowrate in system to 8 lpm.		✓	

TOSCO

CCS RUN NO 2

6-19-81

CONTROLLED CONDENSATION SYSTEM (CCS)
FIELD CHECKPOINT SHEET -- Continued

Checkpoint	Initials		Remarks
	Supervisor	QA Inspector	
SAMPLING RUN		✓	
• Turn vacuum pump on just before inserting probe in stack.		✓	
• Check seal between probe and port to prevent any outside air from entering stack.		✓	
• Run test for 1 hour or until coils are frosted to 1/2 or 2/3 their length.		✓	
• After run, cap both ends of probe and lay in horizontal position.		✓	
• Rinse the CCC coils into the modified Erlenmeyer flask with a maximum of 40 ml D.I. H ₂ O.		✓	
• Was any of the solution lost (ϕ ml estimated)?		✓	
• After probe has cooled, it is rinsed with a maximum of 40 ml D.I. H ₂ O into a 25-ml Erlenmeyer flask.		✓	
- Was any solution lost (ϕ ml estimated)?		✓	
- Clean support equipment prior to next run.		✓	
- Save filter for titration.		✓	

Comments:

CONTROLLED CONDENSATION SYSTEM (CCS)
FIELD DATA SHEET

Plant TOSCO, BAKERSFIELD, CA
Date 6-19-81
Sample Location CRIMSON HEATER
Run No. 2/CCS
Operator P.C. DeRos

Ambient Temperature 104°F
Barometric Pressure 29.40 "Hg
Meter Box Number 524
Meter Orifice Coefficient .7024
Meter α Factor .998

Clock Time (24-hr) clock	Gas Meter Reading (V _m), ft ³	Temperature (°F)								
		Stack	Probe	Filter		°C	Recirc Water	Exit Coil	Dry Gas Meter	
				Skin	Out				In	Out
Sampling Time, min	1220	Init.								
		860.800								
10	1230	864.600		589	1278	655	60	132	114	112
20	1240	867.800		681	1321	693	60	132	114	113
30	1250	870.950		599	1266	687	60	132	114	113
40	1300	874.100		605	1222	649	60	133	114	113
50	1310	877.300		601	1223	649	60	133	114	113
60	1320	880.200		588	1220	647	60	133	114	113
70	1330	882.125		548	1216	640	60	133	114	113
Average	70. min	21.325	696.6	601.6	1256.4	660.	60	132.5	113.4	

From M-5 / Run No 2 Data (average T_s)

4.4 OPERATING DATA FOR SASS

ISOKINETIC SAMPLING WORKSHEET

Plant TOSCO, BAKERSFIELD, CA Performed by J. HOLM

Date 6-19-81

Sample Location CRUDE HEATER

Test No./Type 1/SASS

$$K = \frac{782.687 (C_p)^2 (1-B_{wo})^2 P_s M_d}{K_o^2 M_s P_m}$$

where: K = Constant of fixed and assumed parameters (dimensionless)

Pitot coefficient (dimensionless)	C_p	0.84
Water vapor in the gas stream (proportion by volume)	B_{wo}	12% Assumed
Absolute stack gas pressure (in. Hg) $S_{STATIC} = -0.31$	P_s	29.397
Molecular weight, stack gas dry (1b/1b-mole)	M_d	30.07 Assumed
Orifice coefficient (dimensionless)	K_o	3.7601
Molecular weight, stack gas wet (1b/1b-mole) $M_d(1-B_{wo}) + 18(B_{wo})$	M_s	28.62 Assumed
Absolute meter pressure (in. Hg)	P_m	29.58
$\frac{782.687 (0.84)^2 (1-0.12)^2 (29.397)(30.07)}{(3.7601)^2 (28.62) (29.58)}$	K	29.52

FIELD DATA

Page 1 of 2

Plant TOSCO, BAKERSFIELD
 Date 6-18-81
 Sample Location CRUDE HEATER
 Sample Type SASS
 Run Number 1
 Operator J. HOLM
 Ambient Temperature 95°
 Barometric Pressure 29.42
 Static Pressure, $(H_2O) = 0.31$
 Filter Number(s) 211-003

Leak Check: Initial at 21" Hg., .021 CFM
 Final at 21" Hg., — CFM
 Pitot Leak Check: N/A

Impinger Volumes		
Initial	Final	Net Gain
<u>500</u>	<u>1580</u>	<u>1080</u>
<u>500</u>	<u>1095</u>	<u>595</u>
<u>500</u>	<u>820</u>	<u>915</u>
—	—	—
—	—	—
—	—	—
—	—	—
Silica Gel		
<u>750</u>	<u>873.7</u>	
<u>750</u>	<u>868.5</u>	<u>32.4</u>
	<u>+81.2</u>	
SASS Condensate		
<u>278 (PH = 4.0)</u>		
Total Volume		
<u>5099.4</u>		

Probe Length and Type 5 ft / Pyrex
 Nozzle Size & I.D. 1/122
 Pitot Coefficient & I.D.
 Assumed Moisture 1.2%
 Molecular Weight, Dry, (M_d) 30.07
 Meter Box Number 082
 Meter Coefficient 3.7602
 α Factor 1.0112
 $K = \frac{29.52}{x}$
 $K(N_d)^4 = \frac{x}{()^4} =$
 $4H = K(N_d)^4 \left(\frac{T_m}{T_s} \right) (P)$

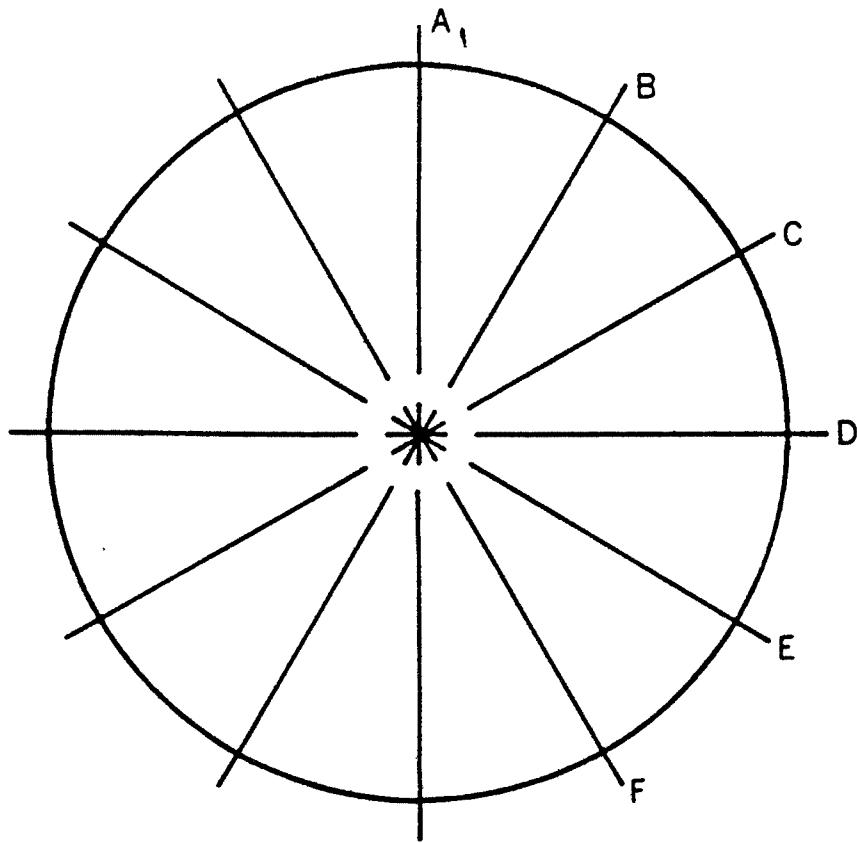
Traverse Point Number	Sampling Time, min	Gas Meter Reading (V_m), ft ³	Velocity Head (ΔP_s), in. H_2O	Orifice Pressure Differential (ΔH), in. H_2O		Temperature °F						Pump Vacuum in. Hg	Avg. \sqrt{AP}		
				Desired	Actual	Stack	Probe	Impinger	Organic Module	Oven	Gas Meter				
											In	Out			
0	16.35	Init. 567.354	—	2.1	2.3	732	695	85	72	290	96	95	11	Pump out 16.35 ft out 1642	
48"	5	588.050	—	2.2	2.3	732	699	83	70	290	99	98	15		
from inner wall to Nozzle E.	15	627.420	—	2.2	2.3	735	731	82	62	290	105	100	15		
30	17.10	687.9	—	2.2	2.3	735	731	82	62	292	107	101	15		
45	17.25	750.0	—	2.2	2.4	740	718	83	63	292	107	101	15		
60	17.40	812.6	—	2.2	2.4	740	713	83	64	295	105	101	15		
75	17.55	874.5	—	2.2	2.4	730	715	85	59	290	105	100	15		
90	18.14	932.9	—	2.2	2.4	720	705	86	59	289	102	100	16		
105	18.29	999.95	—	2.2	2.5	745	712	86	58	292	107	102	16		
120	18.44	1063.05	—	2.2	2.5	735	709	86	60	294	106	101	16		

82.5 min into test • DRAIN CONDENSATE

7602/5/81/Rev 1

Comments: Pitot tube Does not work! M-5 Velocity Profile at SASS location to be USED.
 1 Average AP = .08" H_2O (calculated from M-5 preliminary velocity traverse data)
 2/ Stack temp from M-5 data at near points

NOZZLE MEASUREMENT



DIAMETER DIMENSION

A 1.213
B 1.224
C 1.219
D 1.224
E 1.228
F 1.235

AVG. 1.224 1 7/32

NOZZLE SERIAL

DATE 11/13/80

RECORDED BY Frank

ISOKINETIC SAMPLING WORKSHEET

Plant TOSCO

Performed by J. HOLM

Date 19 JUNE 81

Sample Location CRUDE HTR

Test No./Type 2/SASS

$$K = \frac{782.687 (C_p)^2 (1-B_{wo})^2 P_s M_d}{K_o^2 M_s P_m}$$

where: K = Constant of fixed and assumed parameters (dimensionless)

$P_b = 29.40 @ 830 AM$

Pitot coefficient (dimensionless)	C_p	0.84
Water vapor in the gas stream (proportion by volume)	B_{wo}	13.5 % ASSUMED
Absolute stack gas pressure (in. Hg) <u>29.40 - 0.3 / 13.6</u>	P_s	29.377
Molecular weight, stack gas dry (1b/1b-mole)	M_d	30.07 Assumed.
Orifice coefficient (dimensionless)	K_o	3.7601
Molecular weight, stack gas wet (1b/1b-mole) $M_d(1-B_{wo}) + 18(B_{wo})$ <u>30.01 + 2.43</u>	M_s	28.44 Assumed
Absolute meter pressure (in. Hg). <u>29.40 + 0.0 / 13.6</u>	P_m	29.55
$\frac{0.7056 .7482}{782.687 (0.84)^2 (1-0.135)^2 (29.37) (30.07)}$ <u>(3.7601)^2 (1-0.135) (29.55)</u> <u>14.138</u>	K	30.724

FIELD DATA

Page 1 of 2

Plant TOSCO, BAKERSFIELD, CADate 6-19-81Sample Location CRUDE HEATERSample Type SASSRun Number 2Operator J. HolmAmbient Temperature 104°FBarometric Pressure 29.40 @ 0900Static Pressure, (H_2O) -0.31Filter Number(s) MN-211-002Leak Check: Initial at .06 " Hg, 0.068CFM @ ambient
Final at .26 " Hg, 0.048CFM @ 280°F

Pitot Leak Check: _____

Impinger Volumes		
Initial	Final	Net Gain
500	1040	540
500	760	260
500	525	285

Silica Gel		
750	1017.2	267.2
750	231.4	81.4
750	321.2	72.2

SASS Condensate 1925 @ pH = 2.5Total Volume 3170.8Probe Length and Type 5' PYREXNozzle Size & I.D. 1.224 NO I.D.Pitot Coefficient & I.D. 0.81Assumed Moisture 1.3%Molecular Weight, Dry, (M_d) 30.07Meter Box Number 082Meter Coefficient 3.7602 (K₀) α Factor 1.0112 $K = 30.724$ $K(N_d)^4 = x \left(\frac{P}{T} \right)^4$ $4H = K(N_d)^4 \left(\frac{T_m}{T_s} \right) (P)$

Traverse Point Number	Sampling Time, min	Clock Time (24-hr) Clock	Gas Meter Reading (V_m), ft ³	Velocity Head (A_P), in. H_2O	Orifice Pressure Differential (ΔH), in. H_2O	Temperature °F					Pump Vacuum in. Hg	Avg. VAP		
						Desired	Actual	Stack	Probe	Impinger	Organic Module	Oven	Gas Meter In	Gas Meter Out
0	1130	Init. 640.76				2.2	2.6	580	90	61	253	108	105	16
5	1135	664.050				2.2	2.6	671	91	54	260	117	108	16
15	1145	706.9				2.2	2.6	678	91	60	270	123	114	16
30	1200	768.44				2.2	2.6	676	87	60	270	118	112	16
45	1215	831.02				2.2	2.6	671	88	59	276	120	114	16½
60	1230	894.18				2.2	2.6	668	89	61	287	113	112	16½
75	1250	957.119				2.2	2.6	675	92	59	287	117	112	16½
90	1308	1020.85				2.2	2.6	680	92	61	290	110	105	16½
105	1323	1084.200				2.2	2.6	679	94	60	275	109	104	17
120	1351	1147.75				2.2	2.6							

INITIAL LEAK CHECK TO FILTER FRONT = 0.046 cfm @ 24" Hg; Entire train (inc. PROBE) = 0.068 cfm 7602/5/81/Rev 1 @ 36" Hg

TEST 1, PROBE WAS 37.75" INTO DUCT FROM INNER WALL TO NOZZLE.

TEST 2, SAME POSITION

65min: NOTICED A FEW XAD2 BEADS IN 1ST IMP. SOLUTION

11* 70min into test: DRAINED CONDENSATE, SILICA GEL, O, K NOTE: XAD2 PRESENT (SLIGHT AMOUNT) DEPLETED COND. CHANGED SO₂

21* 75MIN DRAINED PART OF 1ST IMP. INTO NEAT COND. SAMPLE BOULDER DUE TO EXCESSIVE FOAMING

105min into TEST:

Traverse Point Number	Clock Time (24-hr) Clock		Gas Meter Reading (V_m), ft ³	Velocity Head (ΔP_s), in. H ₂ O	Orifice Pressure Differential (ΔH), in. H ₂ O		Temperature °F					Gas Meter	Pump Vacuum in. Hg	Avg. VAP	
	Sampling Time, min	Init.			Desired	Actual	Stack	Probe	Impinger	Organic Module	Oven	In			
48"	135	1406	1210.9		2.2	2.6	673	94	61	289	114	106	17		
31	150	1421	1274.253		2.2	2.6	675	93	60	285	111	105	17		x
	165	1436	1312.15		2.2	2.6	674	93	60	284	112	106	17		
41	180	1451	1402.697		2.2	2.6	675	93	60	284	111	106	17 1/2		x
	195	1520	1466.50		2.2	2.6	673	92	59	283	111	105	17 1/2		
210	1535	1530.40	1530.40		2.2	2.6	673	93	58	290	112	106	17 1/2		
225	1550	1594.30	1594.30		2.2	2.6	672	95	59	290	112	106	17 1/2		
255		1722.075													
			1081.359		2.2	2.6	676.6					110.7	.309		
															5)

Run No. 2/BASSDate 6-19-81Sampling Location CRUDE HEATER

8) AT 150 MIN. INTOTEST COND. DRAINED. SiO₂ OK
 4) AT 180 MIN. ? CHANGED SILICAGEL.

Comments:

Ex from 2/M5 Test Data

SECTION 5
ANALYTICAL RESULTS

- 5.1 Ultimate Analysis of Fuel Oil
- 5.2 Composition of Refinery Gas
- 5.3 SASS Particulate Emissions
- 5.4 EPA Method 5 Particulate Emissions
- 5.5 Sulfur Oxides Emissions by Turbidometric Analysis
- 5.6 Trace Element Analyses
- 5.7 Total Chromatographable Organics (TCO), Gravimetric Organics (GRAV), Infrared (IR) Spectra, Gas Chromatography/Mass Spectrometry (GC/MS) and Low Resolution Mass Spectrometry (LRMS) of Total Sample Extracts
- 5.8 C₁-C₆ Chromatograms
- 5.9 Radiometric Analytical Results
- 5.10 Bioassays

5.1 ULTIMATE ANALYSIS OF FUEL OIL

Prelim. No. 7520
Lab. No. 81m135

RECEIVED 11/17/1981

Subcontract #659186A
Ketbase #3 ORIGINAL
November 16, 1981
Page 2 of 2

T_{TEST} 1 T_{TEST} 2

	#1, Code 813781			#2, Code 813782		
	1st Test	2nd Test	3rd Test	1st Test	2nd Test	3rd Test
Carbon (C), %	-----	87.37	--	--	86.66	--
Hydrogen (H), %	-----	10.47	--	--	10.98	--
Oxygen (O), by difference, %	-----	0.31	--	--	0.55	--
Nitrogen (N), %	-----	.71	.88	.90	.74	.90
Sulfur (S), %	-----	.92	.94	.96	.93	.90
Heating Value: BUT per Pound	-----	18,720	--	--	18,760	--
Gravity, API° @ 70°F	12.3	--	--	12.3	--	--
@ 60°F	11.8	--	--	11.8	--	--

Curtis Tompkins etc

5.2 COMPOSITION OF REFINERY GAS

**TOSCO CORPORATION
BAKERSFIELD REFINERY
GAS ANALYSIS REPORT**

LABORATORY:	OPERATOR:	DATE
Gas lab	J. McMahon	June 18, 1981
UNIT: Crude	Fuel Gas To 11H13	
SAMPLE DATE	6-18-81	
SAMPLE TIME	7 ¹⁵ PM	
PERCENT	Gas Vol.	
HYDROGEN	8.7	
NITROGEN	1.1	
OXYGEN		
CARBON MONOXIDE	0.4	
CARBON DIOXIDE		
HYDROGEN SULFIDE		
METHANE	22.7	
ETHANE	16.7	
ETHYLENE	3.8	
PROPANE	29.2	
PROPYLENE	3.3	
ISOBUTANE	1.1	
NORMAL BUTANE	1.7	
TOTAL BUTENES	0.5	
1, 3-BUTADIENE		
ISOPENTANE	0.2	
NORMAL PENTANE	0.1	
TOTAL PENTENES		
TOTAL C ₆ PLUS	0.5	
BTU/ft ³	1892	
Specific Gravity	1.1381	
DIST. GDD (2) JAK RWT	LDW P+I KVB	REMARKS: PREPARED BY James J. McMahon SUBMITTED BY T. R. Wynn

81 089 (9-80)

**TOSCO CORPORATION
BAKERSFIELD REFINERY
GAS ANALYSIS REPORT**

LABORATORY:	OPERATOR:	DATE:
Gas lab	Jim McMahon	June 22, 1981
UNIT: Crude	Fuel Gas 70/11H13	
SAMPLE DATE	6/19/81	
SAMPLE TIME	3 ³⁰ PM	
PERCENT	Gas Vol.	
HYDROGEN	13 9.3	
NITROGEN	9 0.8	
OXYGEN		
CARBON MONOXIDE	17 0.5	
CARBON DIOXIDE	21	
HYDROGEN SULFIDE	25 0	
METHANE	29 24.5	
ETHANE	33 16.5	
ETHYLENE	37 4.0	
PROPANE	41 28.5	
PROPYLENE	45 3.6	
ISOBUTANE	49 9.5	
NORMAL BUTANE	53 1.3	
TOTAL BUTENES	57 0.7	
1, 3-BUTADIENE	61	
ISOPENTANE	65 0.2	
NORMAL PENTANE	69 0.1	
TOTAL PENTENES	73	
TOTAL C ₆ PLUS	77 0.5	
BTU/ft ³	1842.6	
Specific Gravity	1.1041	
DST GDDC(2) L DW JAK P+I RWT KVB	REMARKS:	PREPARED BY: James A. McMahon SUBMITTED BY:

81089 (9-80)

5.3 SASS PARTICULATE EMISSIONS

ISOKINETIC PERFORMANCE WORKSHEET & PARTICULATE CALCULATIONS

 Plant MISCO, BAKERSFIELD, CA

 Performed by DeRos

 Date 6-18-81

 Sample Location CROSS HEATER

 Test No./Type 1 / SASS

Barometric Pressure (in. Hg)	P_b	29.42
Meter volume (std), $17.64 \left(\frac{V_m}{\alpha} \right) \left(\frac{P_b + \frac{\Delta H}{13.6}}{T_m + 460} \right)$ $17.64 \left(\frac{1061.306}{1.0112} \right) \left(\frac{(29.42) + \frac{(2.44)}{13.6}}{(102.9) + \frac{460}{460}} \right)$	V_m std	973.540
Volume of liquid collected (grams)	V_l_c	3099.4
Volume of liquid at standard condition (scf) $V_l_c \times 0.04707$	V_w std	145.887
Stack gas proportion of water vapor $\frac{V_w \text{ std}}{V_w \text{ std} + V_m \text{ std}}, \frac{(145.887)}{(973.540) + (145.887)}$	B_{wo}	0.1343
Molecular weight, stack gas dry (lb/lb-mole) $(\% \text{CO}_2 \times 0.44) + (\% \text{O}_2 \times 0.32) + (\% \text{N}_2 + \% \text{CO} \times 0.28)$ $(\underline{\quad} \times 0.44) + (\underline{\quad} \times 0.32) + (\underline{\quad} + \underline{\quad} \times 0.28)$	M_d	30.112
Molecular weight, stack gas wet (lb/lb-mole) $M_d(1-B_{wo}) + 18(B_{wo}), (30.112)(1-\underline{0.144}) + 18(\underline{0.144})$	M_s	29.35
Absolute stack pressure (in. Hg) $P_b + \frac{P_{stack} \text{ (in. H}_2\text{O)}}{13.6}, (\underline{\quad}) + \frac{(\underline{\quad})}{13.6}$	P_s	29.39

7602/5/81/Rev 1

TOSCO

1/SASS

6-18-81

Temperature stack gas, average ($^{\circ}$ F)	T_s	714.7
Stack velocity (fps) 85.49 (C_p) $(\sqrt{4P_s \text{ avg}})$	$\sqrt{\frac{T_s \text{ avg} + 460}{P_s M_s}}$	$v_s(\text{avg})$
85.49 (.84) $(\sqrt{29.39})$	$\sqrt{\frac{(714.7) + 460}{(29.39)(29.74)}}$	23.24
Total sample time (minutes)	θ	255
Nozzle diameter, actual (inches)	N_d	1.22
Percent isokinetic (%) 17.33 $(T_s + 460)(V_w \text{ std} + V_m \text{ std})$		
$\frac{17.33 (714.7 + 460)((45.88) + (973.540))}{(255)(27.55)(29.39)(1.22)^2}$	%I	74.15
Area of stack (ft^2) $\pi = 3.1416$ $\pi r^2 \div 144, \pi (\underline{\quad})^2 \div 144$	A_s
Stack gas volume at standard conditions (dscfm) 60 $(1 - B_{wo})V_s \text{ avg } A_s \left(\frac{528}{T_s \text{ avg} + 460} \right) \left(\frac{P_s}{29.92} \right)$	Q_s
$60 (1 - \underline{\quad})(\underline{\quad})(\underline{\quad}) \left(\frac{528}{\underline{\quad} + 460} \right) \left(\frac{(\underline{\quad})}{(29.92)} \right)$	
Particulate matter concentration, dry (gr/dscf) 15.432 $\frac{M_p \text{ (grams)}}{V_m \text{ std}}$, 15.432 $(\frac{\underline{\quad}}{\underline{\quad}})$	$C_s(\text{std})$	
Emission rate of particulate matter (lb/hr) 0.00857 (Q_s) $C_s(\text{std})$, 0.00857 $(\underline{\quad})(\underline{\quad})$	E_p	

7602/5/81/Rev 1

From M-5 Run No 1
5-9

ISOKINETIC PERFORMANCE WORKSHEET & PARTICULATE CALCULATIONS

 Plant TOSCO, BAKERSFIELD, CA

 Performed by DARes

 Date 6-19-81

 Sample Location CRUDE HEATER

 Test No./Type 2/SASS

Barometric Pressure (in. Hg)	P_b	29.40
Meter volume (std), $17.64 \left(\frac{V_m}{\alpha} \right) \left(P_b + \frac{\Delta H}{13.6} \right)$ $17.64 \left(\frac{1081.359}{1.011} \right) \left(\frac{29.70}{110.7} + \frac{2.6}{13.6} \right)$	V_m std	978.299
Volume of liquid collected (grams)	V_l_c	3170.8
Volume of liquid at standard condition (scf) $V_l_c \times 0.04707$	V_w std	149.25
Stack gas proportion of water vapor $\frac{V_w \text{ std}}{V_w \text{ std} + V_m \text{ std}}, \frac{(\text{_____})}{(\text{_____}) + (\text{_____})}$	B_{wo}	.132
Molecular weight, stack gas dry (1b/1b-mole) $(\% CO_2 \times 0.44) + (\% O_2 \times 0.32) + (\% N_2 + \% CO \times 0.28)$ $(\text{_____} \times 0.44) + (\text{_____} \times 0.32) + (\text{_____} + \text{_____} \times 0.28)$	M_d	30.00
Molecular weight, stack gas wet (1b/1b-mole) $M_d(1-B_{wo}) + 18(B_{wo}), (30.00)(1-.132) + 18(.132)$	M_s	28.42
Absolute stack pressure (in. Hg) $P_b + \frac{P_{stack} \text{ (in. H}_2\text{O)}}{13.6}, (\text{_____}) + \frac{(\text{_____})}{13.6}$	P_s	29.38

7602/5/81/Rev 1

TOSCO

2/SASS

6-19-81

Temperature stack gas, average ($^{\circ}$ F)	T_s	696.6 $\frac{1}{1}$
Stack velocity (fps) 85.49 (C_p) $(\sqrt{4P_s \text{ avg}})$	$V_s(\text{avg})$	26.12
$85.49 (\frac{1.84}{2.38})(\frac{309}{28.47}) \sqrt{\frac{(696.6) + 460}{(29.38)(28.47)}}$		
Total sample time (minutes)	θ	255
Nozzle diameter, actual (inches)	N_d	1.224
Percent isokinetic (%) 17.33 $(T_s + 460)(V_w \text{ std} + V_m \text{ std})$	%I	77.59
$\frac{17.33 (696.6 + 460)((1.225) + (\frac{978.299}{29.38}))}{(255)(26.12)(1.22)^2}$		
Area of stack (ft^2) $\pi = 3.1416$ $\pi r^2 \div 144, \pi (\underline{\quad})^2 \div 144$	A_s	
Stack gas volume at standard conditions (dscfm) 60 $(1 - B_{wo})V_s \text{ avg } A_s \left(\frac{528}{T_s \text{ avg} + 460} \right) \left(\frac{P_s}{29.92} \right)$	Q_s	
$60 (1 - \underline{\quad})(\underline{\quad})(\underline{\quad}) \left(\frac{528}{\underline{\quad} + 460} \right) \left(\frac{(\underline{\quad})}{(29.92)} \right)$		
Particulate matter concentration, dry (gr/dscf) 15.432 $\frac{M_p \text{ (grams)}}{V_m \text{ std}}$, 15.432 $\frac{(\underline{\quad})}{(\underline{\quad})}$	$C_s(\text{std})$	
Emission rate of particulate matter (lb/hr) 0.00857 $(Q_s) C_s(\text{std})$, 0.00857 $(\underline{\quad})(\underline{\quad})$	E_p	

 $\frac{1}{1}$ Data from 2/M-5 Test



DATA REPORTING FORM

CUSTOMER CMEA DATE
CUSTOMER CONTRACT NO. 307605.92 ACUREX CONTRACT NO. A81-07-011
RESULTS REPORT TO L. Waterland TELEPHONE
ADDRESS
TOSCO

SAMPLE ID (CUSTOMER)	I-Probe	II Probe	I-Filter	II-Filter	Filter Blk						
SAMPLE ID (LAB)	543	556	545	038	571						
PARAMETER											UNITS
Weight			3.1424	3.2191	2.9151						gram
Initial weight			2.9029	2.9939	2.9151						gram
Ash weight	0.0077	0.0453	0.2395	0.2252	<0.0004						gram

ANALYST _____

REVIEWER _____

5.4 EPA METHOD 5 PARTICULATE EMISSIONS

ISOKINETIC PERFORMANCE WORKSHEET & PARTICULATE CALCULATIONS

Plant TOSCO, BAKERFIELD, CA.Performed by DAResDate 6-18-81Sample Location CRUDE HEATERTest No./Type 1/M-5

Barometric Pressure (in. Hg)	P_b	29.42
Meter volume (std), $17.64 \left(\frac{V_m}{\alpha} \right) \left(P_b + \frac{\Delta H}{13.6} \right)$ $17.64 \left(\frac{103.984}{(1.010)} \right) \left(\frac{(29.42) + \frac{(1.16)}{13.6}}{(111.35) + \frac{460}{13.6}} \right)$	V_m std	93.787
Volume of liquid collected (grams)	V_l_c	376.2
Volume of liquid at standard condition (scf) $V_l_c \times 0.04707$	V_w std	17.708
Stack gas proportion of water vapor $\frac{V_w \text{ std}}{V_w \text{ std} + V_m \text{ std}}, \frac{(17.708)}{(17.708) + (93.787)}$	B_{wo}	.159
Molecular weight, stack gas dry $(1b/1b\text{-mole})$ $(\% CO_2 \times 0.44) + (\% O_2 \times 0.32) + (\% N_2 + \% CO \times 0.28)$ $(12.2 \times 0.44) + (4.0 \times 0.32) + (83.8 + \underline{\quad}) \times 0.28)$	M_d	30.112
Molecular weight, stack gas wet $(1b/1b\text{-mole})$ $M_d(1-B_{wo}) + 18(B_{wo}), (30.112)(1-.159) + 18(.159)$	M_s	28.19
Absolute stack pressure (in. Hg) $P_b + \frac{P_{stack} \text{ (in. } H_2O\text{)}}{13.6}, (29.42) + \frac{(-.31)}{13.6}$	P_s	29.39

✓ Molecular weight from B. DeRosier Field Data 7602/5/81/Rev 1
 (KUB - CM instrument var) 5-14

TOSCO

1/m-5

6-18-81

Temperature stack gas, average ($^{\circ}$ F)	T_s	714.7
Stack velocity (fps) 85.49 (C_p) ($\sqrt{4P_s \text{ avg}}$) $\sqrt{\frac{T_s \text{ avg} + 460}{P_s M_s}}$ 85.49 (.84) (.332) $\sqrt{\frac{(714.7) + 460}{(29.39)(28.39)}}$	$v_s(\text{avg})$	28.39
Total sample time (minutes)	θ	180
Nozzle diameter, actual (inches)	N_d	.382
Percent isokinetic (%) 17.33 $(T_s + 460)(V_w \text{ std} + V_m \text{ std})$ $\frac{\theta \quad v_s \quad P_s \quad N_d^2}{17.33 (714.7 + 460)((17.708) + (93.787))}$ $\frac{}{(180)(28.39)(29.39)(.382)^2}$	%I	103.57
Area of stack (ft^2) $\pi = 3.1416$ $\pi r^2 \div 144$, $\pi (\underline{\hspace{1cm}})^2 \div 144$	A_s	11.79
Stack gas volume at standard conditions (dscfm) 60 $(1 - B_{wo})v_s \text{ avg } A_s \left(\frac{528}{T_s \text{ avg} + 460} \right) \left(\frac{P_s}{29.92} \right)$ $60 (1 - \underline{\hspace{1cm}})(\underline{\hspace{1cm}})(\underline{\hspace{1cm}}) \left(\frac{528}{\underline{\hspace{1cm}} + 460} \right) \left(\frac{(\underline{\hspace{1cm}})}{(29.92)} \right)$	Q_s	7458
Particulate matter concentration, dry (gr/dscf) 15.432 $\frac{M_p \text{ (grams)}}{V_m \text{ std}}$, 15.432 $\frac{(\underline{\hspace{1cm}})}{(\underline{\hspace{1cm}})}$	$C_s(\text{std})$	
Emission rate of particulate matter (lb/hr) 0.00857 $(Q_s) C_s(\text{std})$, 0.00857 $(\underline{\hspace{1cm}})(\underline{\hspace{1cm}})$	E_p	

7602/5/81/Rev 1

ISOKINETIC PERFORMANCE WORKSHEET & PARTICULATE CALCULATIONS

 Plant Tosco, Bakersfield, CA

 Performed by DuRos

 Date 6-17-81

 Sample Location CRUDE HEATER

 Test No./Type 3/m-5

Barometric Pressure (in. Hg)	P_b	29.40
Meter volume (std), $17.64 \left(\frac{V_m}{\alpha} \right) \left(\frac{P_b + \Delta H}{T_m + 460} \right)$ $17.64 \left(\frac{(77.64)}{(1.010)} \right) \left(\frac{(29.40) + (.77)}{(113.2) + 460} \right)$	V_m std	87.833
Volume of liquid collected (grams)	V_l_c	367.7
Volume of liquid at standard condition (scf) $V_l_c \times 0.04707$	V_w std	17.308
Stack gas proportion of water vapor $\frac{V_w}{V_w + V_m}$ std , $\frac{(17.308)}{(17.308) + (87.833)}$	B_{wo}	.1646
Molecular weight, stack gas dry (lb/lb-mole) $(\% CO_2 \times 0.44) + (\% O_2 \times 0.32) + (\% N_2 + \% CO \times 0.28)$ $(11.7 \times 0.44) + (3.3 \times 0.32) + (3 \times 0.28)$	M_d	30.00
Molecular weight, stack gas wet (lb/lb-mole) $M_d(1-B_{wo}) + 18(B_{wo})$, $30.00(1-.1646) + 18(.1646)$	M_s	28.03
Absolute stack pressure (in. Hg) $P_b + \frac{P_{stack} \text{ (in. H}_2\text{O)}}{13.6}$, $(29.40) + \frac{(-.31)}{13.6}$	P_s	29.38

From B.Delosier field data KV3 - CMVAN 7602/5/81/Rev 1

Tosco

2/M-5

6-17-81

Temperature stack gas, average ($^{\circ}$ F)	T_s	696.6
Stack velocity (fps) 85.49 (C_p) ($\sqrt{4P_s \text{ avg}}$)	$v_s(\text{avg})$	26.3
$85.49 (\underline{.87})(\underline{.307}) \sqrt{\frac{(696.6) + 460}{(29.38)(28.03)}}$		
Total sample time (minutes)	θ	180
Nozzle diameter, actual (inches) (SN707)	N_d	.382
Percent isokinetic (%) 17.33 ($T_s + 460$) ($V_w \text{ std} + V_m \text{ std}$)	$\%I$	103.84
$\frac{\theta v_s P_s N_d^2}{17.33 (696.6 + 460)((17.508) + (\underline{\quad}))}$ $(180)(.382)(29.38)(.382)^2$		
Area of stack (ft^2) $\pi = 3.1416$ $\pi r^2 \div 144$, $\pi (\underline{\quad})^2 \div 144$	A_s	...?
Stack gas volume at standard conditions (dscfm) 60 (1 - B_{wo}) $V_s \text{ avg } A_s \left(\frac{528}{T_s \text{ avg} + 460} \right) \left(\frac{P_s}{29.92} \right)$	Q_s	...?.
$60 (1 - \underline{\quad})(\underline{\quad})(\underline{\quad}) \left(\frac{528}{\underline{\quad} + 460} \right) \left(\frac{(\underline{\quad})}{(29.92)} \right)$		
Particulate matter concentration, dry (gr/dscf) 15.432 $\frac{M_p \text{ (grams)}}{V_m \text{ std}}$, 15.432 $\frac{(\underline{\quad})}{(\underline{\quad})}$	$C_s \text{ (std)}$	
Emission rate of particulate matter (lb/hr) 0.00857 $(Q_s) C_s \text{ (std)}$, 0.00857 $(\underline{\quad})(\underline{\quad})$	E_p	

7602/5/81/Rev 1

7602.92

B.C. Daros

CMEA: TOSCO
BAKERSFIELD, CA

TEST I.D.	Vm(STD)	FRONT HALF		BACK HALF		IMPIINGER CONTENTS			
		PROBE & NOZZEL CATCH (Mg) <u>11</u>	M-5 FILTER CATCH (Mg) <u>11</u>	IMPIINGER RINSE (acetone) (Mg) <u>11</u>	IMPIINGER AQUEOUS PHASE (Mg) <u>11</u>				
M-5 TEST 1	93.787	33.81	48.81	2.02	2.08	20.84			
M-5 TEST 2	87.833	2.88	46.92	7.54	.30	.21			

11 Blank corrected value.

ACUREX ANALYTICAL REPORT

Sample of: Tesco

Sample Date: June 16 and 19, 1981

Requested By: Bruce Davis

I.D. Number: 2602.92 / CMEA

Analytical Method: EPA Method 5 Protocol - Ether/Chloroform Extractions
Date of Analysis: September 3, 1981 of Impinger Liquids

Lab I.D. Number	Component	Analytical Result	Unit
813795 - Test 1	175 mls		
- Organic Fraction Buck Hall		2.02	Net Gain milligrams
813579 - Test 2	140 mls		
- Organic Fraction Buck Hall		7.54	

Analysis By J. Sayer / L. Whitmer
5-19 Date September 15, 1981

ACUREX ANALYTICAL REPORT

Sample of: Tosco

Sample Date: June 16 and 19, 1981

Requested By: Bruce Davis

I.D. Number: 7602.92/CMEA

Analytical Method: EPA Method 5 Protocol - Ether / Chloroform Extraction
 Date of Analysis: Sept. 3, 1981 of Impinger Liquids

Lab I.D. Number	Component	Analytical Result	Unit
813549 - Test 1	484 mls		
- Aqueous Phase		3.57 - 1.49 = 2.08	
- Organic Phase		21.84 - .87 = 20.84	Net Gain milligrams
813578 - Test 2	485 mls		
- Aqueous Phase		1.79 - 1.49 = .30	
- Organic Phase		1.08 - .87 = .21	
813574 - Water Blank	218 mls		
- Aqueous Phase		0.67 .00307 mg/ml	
- Organic Phase		0.87	

5-20

Analysis By J. Sayer / G. Whiteman
 Date September 15, 1981

ACUREX ANALYTICAL REPORT

Sample of: Tesco

Sample Date: June 16 and 19, 1981

Requested By: Bruce Doros

I.D. Number: 760292/CINCA

Analytical Method: Gravimetric Analysis of Filters

Date of Analysis: July 31, 1981

Lab I.D. Number	Component	Analytical Result	Unit
813548 - Test #1	96-34	48.81	
813577 - Test #2	96-35	46.92	Net Gain milligrams

5-21

Analysis By

Date

J. Sayer / G. Whiteman
9/15/81

ACUREX ANALYTICAL REPORT

Sample of: Tesco

Sample Date: June 16 and 19, 1981

Requested By: Bruce Davis

I.D. Number: 7602.92/CMEA

Analytical Method: Densimetric Analysis of Acetone Probe Wash
Date of Analysis: September 1 and 2, 1981

Lab I.D. Number	Component	Analytical Result	Unit
813546 - Test 1	45 mls	$34.08 - .27$ $= 33.81$	Net gain milligrams
813576 - Test 2	110 mls	$3.54 - .66$ $= 2.88$	
813547 - Acetone Blank	100. -	0.60 .006 mg/ml	

5-22

Analysis By J. Sawyer / S. Whittemore
Date September 14, 1981

5.5 SULFUR OXIDES EMISSIONS BY TURBIDOMETRIC ANALYSIS

CONTROLLED CONDENSATION SYSTEM (CCS)
LABORATORY DATA SHEET

Plant CCSC, BAKERSFIELD, CA
Date 6-8-81
Sample Location CRUDE HEATER
Run No. 1-CCS

Analyst B.C. DiRosa
Date Lab Analysis Completed 10-21-81

Method BARIUM/THORIN Titrant Barium Sulfate Titration Data
Normality .0166 Indicator THORIN

Sample Description	Probe, Nozzle and Filter Rinse	G/R Coil Rinse	Impinger Contents and Rinse	H ₂ O Blank	3% H ₂ O ₂ Blank
Sample No.	813552	813553	813575	813793	813551
Vol. of Sample	80.0	66.0	224.0	UNITY	UNITY
Vol. of Aliquot	10.0	10.0	10.0	10.0	10.0
Vol. of Titrant Used	.10	.95	.95	.05	.05
Average Vol. of Titrant Used	—	.93	10.73	.05	.05

Calculations

Vol. of Gas Sampled (V_M) 19.26 ft³, Avg. Meter Temp (T_M) 116.3 °F,
Meter Pressure (P_M) 29.43 "Hg, Meter α Factor .998 dimensionless

$\frac{(.95 - .05) \cdot .0166 N}{10.0} = .0015 N$	$.0015 N \times 66.0 = .099 \text{ mg SO}_4^{\text{-}} / \text{Sample}$
$\frac{96 \times .099}{2} = 4.733 \text{ Mg SO}_4^{\text{-}}$	$48.15 \left(\frac{4.733}{96}, \text{MgSO}_4 \right) \left(\frac{116.3}{19.26}, T_M + 460 \right)$
$\text{PPM SO}_4 = \frac{48.15 \left(\frac{4.733}{96}, \text{MgSO}_4 \right) \left(\frac{116.3}{19.26}, T_M + 460 \right)}{64 \left(\frac{19.26}{V_M}, \frac{29.43}{P_M} \right) (.998, \alpha)}$	
ppm $\text{SO}_4 = 2.41$	
$\frac{(10.73 - .05) \cdot .0166 N}{10.0} = .0177 N$	$.0177 N \times 224.0 = 3.97 \text{ mg SO}_4^{\text{-}} / \text{Sample}$
$\frac{3.97 \times 96}{2} = 190.58 \text{ mg SO}_4^{\text{-}}$	$48.15 \left(\frac{127.04}{64}, \text{MgSO}_2 \right) \left(\frac{116.3}{19.26}, T_M + 460 \right)$
$\text{PPM SO}_2 = \frac{48.15 \left(\frac{127.04}{64}, \text{MgSO}_2 \right) \left(\frac{116.3}{19.26}, T_M + 460 \right)}{64 \left(\frac{19.26}{V_M}, \frac{29.43}{P_M} \right) (.998, \alpha)}$	
ppm $\text{SO}_2 = 96.98$	

✓ COLOR CHANGE UPON ADDITION OF INDICATOR

ISOKINETIC PERFORMANCE WORKSHEET & PARTICULATE CALCULATIONS

 Plant TOSCO, BAKERSFIELD, CA

 Performed by DiaRe

 Date 6-18-81

 Sample Location CRUDE HEATER

 Test No./Type 1/CCS

Barometric Pressure (in. Hg)	P_b	29.42
Meter volume (std), $17.64 \left(\frac{V_m}{\alpha} \right) \left(P_b + \frac{\Delta H}{13.6} \right)$ $17.64 \left(\frac{(19.26)}{(998)} \right) \left(\frac{(29.42)}{(116.3)} + \frac{(.1)}{13.6} \right)$	V_m std	17.38
Volume of liquid collected (grams)	V_l_c	
Volume of liquid at standard condition (scf) $V_l_c \times 0.04707$	V_w std	
Stack gas proportion of water vapor $\frac{V_w \text{ std}}{V_w \text{ std} + V_m \text{ std}}, \frac{(\text{_____})}{(\text{_____}) + (\text{_____})}$	B_{wo}	
Molecular weight, stack gas dry (lb/lb-mole) $(\% CO_2 \times 0.44) + (\% O_2 \times 0.32) + (\% N_2 + \% CO \times 0.28)$ $(\text{_____} \times 0.44) + (\text{_____} \times 0.32) + .(\text{_____} + \text{_____} \times 0.28)$	M_d	
Molecular weight, stack gas wet (lb/lb-mole) $M_d(1-B_{wo}) + 18(B_{wo}), (\text{_____})(1-\text{_____}) + 18(\text{_____})$	M_s	
Absolute stack pressure (in. Hg) $P_b + \frac{P_{stack} \text{ (in. H}_2\text{O)}}{13.6}, (\text{_____}) + \frac{(\text{_____})}{13.6}$	P_s	

7602/5/81/Rev 1

CONTROLLED CONDENSATION SYSTEM (CCS)
LABORATORY DATA SHEET

Plant TSC-2 BAKERSFIELD, CA
Date 6-19-81
Sample Location CRUDE HEATER
Run No. 2-CCS

Analyst B.C. Areos
Date Lab Analysis Completed 10-21-81

Method BARIUM THORIN Titrant Barium Iodide Normality .0166 Indicator THORIN

Sample Description	Probe, Nozzle and Filter Rinse	G/R Coil Rinse	Impinger Contents and Rinse	H ₂ O Blank	3% H ₂ O ₂ Blank
Sample No.	813790	813791	813792	813793	
Vol. of Sample	80.0	68.0	222.0	UNITY	UNITY
Vol. of Aliquot	10.0	10.0	10.0	10.0	10.0
Vol. of Titrant Used	1.75 1.80 1.75	1.75 1.80 1.75	14.25 14.20 14.20	.05 .05 —	.05 .05 —
Average Vol. of Titrant Used	—	1.767	14.22	.05	.05

Calculations

Vol. of Gas Sampled (V_M) 21.325 ft³, Avg. Meter Temp (T_M) 115.4°F,
Meter Pressure (P_M) 29.41"Hg, Meter α Factor .798 dimensionless

$\frac{(1.767 - .05) \cdot .0166N}{10.0} = .0012N$	$.0012N \times 68.0 = .0816 \text{ meq SO}_4/\text{sample}$
$\frac{96 \times .0816}{2} = 3.917 \text{ MgSO}_4$	$\text{PPM } \text{SO}_4 = \frac{48.15 (\frac{.917}{96}, \text{MgSO}_4)(115.4, T_M + 460)}{21.325, V_M(29.41, P_M) (.798, \alpha)}$
$\text{ppm SO}_4 = 1.8$	
$\frac{(14.22 - .05) \cdot .0166N}{10.0} = .0235N$	$.0235N \times 222.0 = 5.222 \text{ meq SO}_4/\text{sample}$
$\frac{5.222 \times 96}{2} = 250.65$	$\text{PPM } \text{SO}_2 = \frac{48.15 (\frac{167.1}{64}, \text{MgSO}_2)(115.4, T_M + 460)}{21.325, V_M(29.41, P_M) (.798, \alpha)}$
$250.65 \times \frac{64}{96} = 167.10$	
$\text{ppm SO}_2 = 114.7$	

ISOKINETIC PERFORMANCE WORKSHEET & PARTICULATE CALCULATIONS

 Plant TOSCO, BAKERSFIELD, CA

 Performed by DuRex

 Date 6-19-81

 Sample Location CRUDE HEATER

 Test No./Type 2/CCS

Barometric Pressure (in. Hg)	P_b	29.40
Meter volume (std), $17.64 \left(\frac{V_m}{\alpha} \right) \left(P_b + \frac{\Delta H}{13.6} \right)$ $17.64 \left(\frac{(21.32)}{(.998)} \right) \left(\frac{(29.40)}{(113.4)} + \frac{(.1)}{13.6} \right)$	V_m std	19.32
Volume of liquid collected (grams)	V_l_c	
Volume of liquid at standard condition (scf) $V_l_c \times 0.04707$	V_w std	
Stack gas proportion of water vapor $\frac{V_w \text{ std}}{V_w \text{ std} + V_m \text{ std}}, \quad \frac{(\underline{\quad})}{(\underline{\quad}) + (\underline{\quad})}$	B_{wo}	
Molecular weight, stack gas dry (lb/lb-mole) $(\% CO_2 \times 0.44) + (\% O_2 \times 0.32) + (\% N_2 + \% CO \times 0.28)$ $(\underline{\quad} \times 0.44) + (\underline{\quad} \times 0.32) + (\underline{\quad} + \underline{\quad} \times 0.28)$	M_d	
Molecular weight, stack gas wet (lb/lb-mole) $M_d(1-B_{wo}) + 18(B_{wo}), (\underline{\quad})(1-\underline{\quad}) + 18(\underline{\quad})$	M_s	
Absolute stack pressure (in. Hg) $P_b + \frac{P_{stack} \text{ (in. H}_2\text{O)}}{13.6}, (\underline{\quad}) + \frac{(\underline{\quad})}{13.6}$	P_s	

7602/5/81/Rev 1

5.6 TRACE ELEMENT ANALYSES

Fuel

Filter

Filter Blank

XAD

XAD Blank

Impinger 1

Impinger 1 Blank

Mercury, Antimony, Arsenic

COMMERCIAL TESTING & ENGINEERING CO.

Reply to

GENERAL OFFICES: 228 NORTH LA SALLE STREET, CHICAGO, ILLINOIS 60601 · AREA CODE 312 726-8434
INSTRUMENTAL ANALYSIS DIVISION: 14335 WEST 44TH AVENUE, GOLDEN, COLORADO 80401, PHONE: 303-278-9521

To: Mr. Roy Belletto
Acurex Corporation
485 Clyde Street
Mountain View, CA 94042



RECEIVED

Date: December 29, 1981

JAN U & REC'D

ACUREX

Analyst: J. Oldham

Release No. 7

P. O. No.:

Sample No.: 10-24-3
CTE 781

SPARK SOURCE MASS SPECTROGRAPHIC ANALYSIS

IAD No.: 97-H690-116-13

CONCENTRATION IN PPM WEIGHT

ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.
Uranium	<0.5	Terbium		Ruthenium		Vanadium	2
Thorium	<0.4	Gadolinium		Molybdenum		Titanium	6
Bismuth		Europium		Niobium		Scandium	<0.1
Lead	0.2	Samarium		Zirconium	0.8	Calcium	30
Thallium		Neodymium		Yttrium	0.2	Potassium	80
Mercury	NR	Praseodymium		Strontium	0.1	Chlorine	2
Gold		Cerium		Rubidium		Sulfur	>320
Platinum	2	Lanthanum		Bromine	0.5	Phosphorus	9
Iridium		Barium	2	Selenium		Silicon	45
Osmium		Cesium		Arsenic		Aluminum	15
Rhenium		Iodine	0.1	Germanium		Magnesium	10
Tungsten		Tellurium		Gallium	0.3	Sodium	<0.1
Tantalum		Antimony		Zinc	9	Fluorine	=0.2
Hafnium		Tin		Copper	7	Oxygen	NR
Lutetium		Indium	STD	Nickel	8	Nitrogen	NR
Ytterbium		Cadmium		Cobalt	<0.1	Carbon	NR
Thulium		Silver		Iron	13	Boron	0.1
Erbium		Palladium		Manganese	0.7	Beryllium	
Holmium		Rhodium		Chromium	0.6	Lithium	0.9
Dysprosium						Hydrogen	NR

STD - Internal Standard
NR - Not Reported
All elements not detected < 0.1 ppm
MC - Major Component
INT - Interference

Approved: *Robert L. Taylor*
for M.L.Taylor (2-9 Dec 81).

COMMERCIAL TESTING & ENGINEERING CO.

Reply to

GENERAL OFFICES: 228 NORTH LA SALLE STREET, CHICAGO, ILLINOIS 60601 - AREA CODE 312 726-8434
INSTRUMENTAL ANALYSIS DIVISION, 14335 WEST 44TH AVENUE, GOLDEN, COLORADO 80401. PHONE: 303-278-9521



To: Mr. Roy Belletto
Acurex Corporation
485 Clyde Street
Mountain View, CA 94042

Date: December 29, 1981

Release No. 7

Analyst: J. Oldham

P. O. No.:

Sample No.: 10-24-5
CTE 782

SPARK SOURCE MASS SPECTROGRAPHIC ANALYSIS

IAD No.: 97-H690-116-13

Taco 2 L-1
CONCENTRATION IN PPM WEIGHT

ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.
Uranium	<0.8	Terbium		Ruthenium		Vanadium	0.8
Thorium	<0.9	Gadolinium		Molybdenum		Titanium	10
Bismuth		Europium		Niobium		Scandium	
Lead	0.9	Samarium		Zirconium		Calcium	16
Thallium		Neodymium		Yttrium		Potassium	66
Mercury	NR	Praseodymium		Strontium	0.5	Chlorine	5
Gold		Cerium		Rubidium		Sulfur	33
Platinum	*11	Lanthanum		Bromine		Phosphorus	5
Iridium		Barium	1	Selenium		Silicon	16
Osmium		Cesium		Arsenic		Aluminum	13
Rhenium		Iodine		Germanium		Magnesium	12
Tungsten		Tellurium		Gallium	0.1	Sodium	6
Tantalum		Antimony		Zinc	3	Fluorine	=0.4
Hafnium		Tin		Copper	6	Oxygen	NR
Lutetium		Indium	STD	Nickel	8	Nitrogen	NR
Ytterbium		Cadmium		Cobalt	<0.2	Carbon	NR
Thulium		Silver		Iron	26	Boron	1
Erbium		Palladium		Manganese	0.5	Beryllium	
Holmium		Rhodium		Chromium	1	Lithium	2
Dysprosium			* Heterogeneous			Hydrogen	NR

STD — Internal Standard

NR — Not Reported

All elements not detected < 0.1ppm

MC — Major Component

INT — Interference

Approved: *Robert L. Taylor*
for M.L. Jacobs *29 Dec. 81*

COMMERCIAL TESTING & ENGINEERING CO.

Reply to

GENERAL OFFICES: 228 NORTH LA SALLE STREET, CHICAGO, ILLINOIS 60601 - AREA CODE 312 728-8434
INSTRUMENTAL ANALYSIS DIVISION, 14335 WEST 44TH AVENUE, GOLDEN, COLORADO 80401, PHONE: 303-278-9521



To: Mr. Roy Belletto
Acurex Corporation
485 Clyde Avenue
Mountain View, CA. 94042

Date: October 14, 1981

Release No. 6 Exhibit A
P. O. No.: Subcontract No. SW59159A

Analyst: J. Oldham

Sample No.: A81-07-011-545 SPARK SOURCE MASS SPECTROGRAPHIC ANALYSIS IAD No.: 97-H437-116-13

TOSCO 1 FILTER

CONCENTRATION IN $\mu\text{g}/\text{cm}^2$

ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.
Uranium		Terbium		Ruthenium		Vanadium	0.2
Thorium		Gadolinium		Molybdenum	0.004	Titanium	0.004
Bismuth		Europium		Niobium	<0.0001	Scandium	<0.0001
Lead	0.003	Samarium		Zirconium	0.005	Calcium	MC
Thallium		Neodymium		Yttrium	0.0001	Potassium	>0.4
Mercury	NR	Praseodymium	<0.0001	Strontium	0.001	Chlorine	0.008
Gold		Cerium	0.0002	Rubidium	0.0007	Sulfur	>0.2
Platinum		Lanthanum	0.0003	Bromine	0.003	Phosphorus	0.01
Iridium		Barium	0.009	Selenium	<0.0001	Silicon	MC
Osmium		Cesium		Arsenic	NR	Aluminum	>0.03
Rhenium		Iodine	0.0002	Germanium	<0.0001	Magnesium	MC
Tungsten		Tellurium	0.0001	Gallium	0.001	Sodium	>0.08
Tantalum		Antimony	NR	Zinc	0.05	Fluorine	≈0.1
Hafnium		Tin	<0.0001	Copper	0.004	Oxygen	NR
Lutetium		Indium	STD	Nickel	MC	Nitrogen	NR
Ytterbium		Cadmium	0.0002	Cobalt	0.01	Carbon	NR
Thulium		Silver	0.001	Iron	0.2	Boron	>0.6
Erbium		Palladium		Manganese	0.002	Beryllium	<0.001
Holmium		Rhodium		Chromium	0.004	Lithium	0.01
Dysprosium						Hydrogen	NR

STD — Internal Standard

NR — Not Reported

All elements not detected < 0.0001 $\mu\text{g}/\text{cm}^2$

MC — Major Component > 1 $\mu\text{g}/\text{cm}^2$

INT — Interference

Approved:

M. J. Beccals

COMMERCIAL TESTING & ENGINEERING CO.

GENERAL OFFICES: 228 NORTH LA SALLE STREET, CHICAGO, ILLINOIS 60601 • AREA CODE 312 726-8434
INSTRUMENTAL ANALYSIS DIVISION, 14335 WEST 44TH AVENUE, GOLDEN, COLORADO 80401, PHONE: 303-278-9521

Reply to



To: Mr. Roy Belletto
Acurex Corporation
485 Clyde Avenue
Mountain View, CA 94042

Date: October 14, 1981

Release No. 6 Exhibit A
P.O. No. Subcontract No. SW59159A

Analyst: J. Oldham

Sample No.: A81-07-011-038 SPARK SOURCE MASS SPECTROGRAPHIC ANALYSIS IAD No.: 97-H437-116-13

TOSCO 2 FILTER CONCENTRATION IN $\mu\text{g}/\text{cm}^2$

ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.
Uranium	0.0003	Terbium		Ruthenium		Vanadium	0.2
Thorium	0.0003	Gadolinium		Molybdenum	0.009	Titanium	0.02
Bismuth		Europium		Niobium	<0.0001	Scandium	<0.0001
Lead	0.006	Samarium		Zirconium	0.002	Calcium	MC
Thallium		Neodymium		Yttrium	0.0003	Potassium	>0.4
Mercury	NR	Praseodymium	<0.0001	Strontium	0.002	Chlorine	0.02
Gold		Cerium	0.0005	Rubidium	0.0003	Sulfur	>0.2
Platinum		Lanthanum	0.0004	Bromine	0.002	Phosphorus	0.02
Iridium		Barium	0.02	Selenium	0.0004	Silicon	MC
Osmium		Cesium		Arsenic	NR	Aluminum	>0.03
Rhenium		Iodine	0.0001	Germanium	0.0002	Magnesium	MC
Tungsten		Tellurium	<0.0001	Gallium	0.001	Sodium	>0.08
Tantalum		Antimony	NR	Zinc	0.05	Fluorine	=0.1
Hafnium		Tin	0.0001	Copper	0.01	Oxygen	NR
Lutetium		Indium	STD	Nickel	0.9	Nitrogen	NR
Ytterbium		Cadmium	0.0002	Cobalt	0.02	Carbon	NR
Thulium		Silver	0.01	Iron	0.3	Boron	0.1
Erbium		Palladium		Manganese	0.01	Beryllium	<0.0001
Holmium		Rhodium		Chromium	0.04	Lithium	0.002
Dysprosium						Hydrogen	NR

STD — Internal Standard

NR — Not Reported

All elements not detected < 0.0001 $\mu\text{g}/\text{cm}^2$

MC — Major Component > 1 $\mu\text{g}/\text{cm}^2$

INT — Interference

Approved:

COMMERCIAL TESTING & ENGINEERING CO.

GENERAL OFFICES: 228 NORTH LA SALLE STREET, CHICAGO, ILLINOIS 60601 · AREA CODE 312 728-8434
INSTRUMENTAL ANALYSIS DIVISION: 14335 WEST 44TH AVENUE, GOLDEN, COLORADO 80401, PHONE: 303-278-9521

Reply to

To: Mr. Roy Belletto
Acurex Corporation
485 Clyde Avenue
Mountain View, CA 94042

Date: October 14, 1981



Release No. 6 Exhibit A
P. O. No.: Subcontract No. SW59159A

Analyst: J. Oldham

Sample No A81-07-011-571 SPARK SOURCE MASS SPECTROGRAPHIC ANALYSIS IAD No.. 97-H437-116-13

TOSCO FILTER BLANK CONCENTRATION IN $\mu\text{g}/\text{cm}^2$

ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.
Uranium		Terbium		Ruthenium		Vanadium	0.0003
Thorium		Gadolinium		Molybdenum	0.0002	Titanium	0.1
Bismuth		Europium		Niobium	0.0001	Scandium	<0.0001
Lead	0.0007	Samarium		Zirconium	0.009	Calcium	MC
Thallium		Neodymium		Yttrium	<0.0001	Potassium	>0.5
Mercury	NR	Praseodymium	<0.0001	Strontium	0.005	Chlorine	MC
Gold	*0.0001	Cerium	0.0005	Rubidium	0.0008	Sulfur	0.06
Platinum		Lanthanum	0.0004	Bromine	0.001	Phosphorus	0.007
Iridium		Barium	0.004	Selenium		Silicon	MC
Osmium		Cesium		Arsenic	NR	Aluminum	>0.04
Rhenium		Iodine	<0.0001	Germanium	<0.0001	Magnesium	MC
Tungsten		Tellurium		Gallium	0.0006	Sodium	>0.09
Tantalum		Antimony	NR	Zinc	0.006	Fluorine	=0.3
Hafnium	0.0004	Tin		Copper	0.002	Oxygen	NR
Lutetium		Indium	STD	Nickel	0.01	Nitrogen	NR
Ytterbium		Cadmium	0.0001	Cobalt	0.0003	Carbon	NR
Thulium		Silver	*0.0001	Iron	0.02	Boron	0.2
Erbium		Palladium		Manganese	0.002	Beryllium	
Holmium		Rhodium		Chromium	0.0004	Lithium	0.0005
Dysprosium		*Heterogeneous				Hydrogen	NR

STD - Internal Standard

NR - Not Reported

All elements not detected < 0.0001 $\mu\text{g}/\text{cm}^2$

MC - Major Component > 1 $\mu\text{g}/\text{cm}^2$

INT - Interference

Approved:

COMMERCIAL TESTING & ENGINEERING CO.

GENERAL OFFICES: 228 NORTH LA SALLE STREET, CHICAGO, ILLINOIS 60601 • AREA CODE 312 728-8434
INSTRUMENTAL ANALYSIS DIVISION, 14335 WEST 44TH AVENUE, GOLDEN, COLORADO 80401, PHONE: 303-278-9521

Reply to



To: Mr. Roy A. Belletto
Aurex Corporation
485 Clyde Avenue
Mountain View, CA 94042

Date: October 12, 1981

Release No. 6 Exhibit A
P. O. No.: Subcontract No. SW59159A

Analyst: J. Oldham

Sample No. A81-07-011-569 SPARK SOURCE MASS SPECTROGRAPHIC ANALYSIS IAD No.: 97-H437-116-13

Tosco 1 XAD

CONCENTRATION IN PPM WEIGHT

ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.
Uranium		Terbium		Ruthenium		Vanadium	<0.1
Thorium		Gadolinium		Molybdenum	0.2	Titanium	1
Bismuth		Europium		Niobium	<0.1	Scandium	<0.1
Lead	0.2	Samarium		Zirconium	0.2	Calcium	36
Thallium		Neodymium		Yttrium		Potassium	39
Mercury	NR	Praseodymium		Strontium	4	Chlorine	4
Gold		Cerium		Rubidium	<0.1	Sulfur	8
Platinum	0.7	Lanthanum		Bromine	2	Phosphorus	1
Iridium		Barium	2	Selenium		Silicon	9
Osmium		Cesium	<0.1	Arsenic	NR	Aluminum	0.6
Rhenium		Iodine	0.1	Germanium		Magnesium	1
Tungsten		Tellurium		Gallium	<0.1	Sodium	11
Tantalum		Antimony	NR	Zinc	7	Fluorine	<0.1
Hafnium		Tin		Copper	3	Oxygen	NR
Lutetium		Indium	STD	Nickel	9	Nitrogen	NR
Ytterbium		Cadmium		Cobalt	<0.1	Carbon	NR
Thulium		Silver		Iron	6	Boron	<0.1
Erbium		Palladium		Manganese	0.3	Beryllium	
Holmium		Rhodium		Chromium	1	Lithium	<0.1
Dysprosium						Hydrogen	NR

STD - Internal Standard
NR - Not Reported
All elements not detected < 0.1 ppm
MC - Major Component
INT - Interference

Approved:

W. Jacobs

COMMERCIAL TESTING & ENGINEERING CO.

Reply to

GENERAL OFFICES: 228 NORTH LA SALLE STREET, CHICAGO, ILLINOIS 60601 - AREA CODE 312 726-8434
INSTRUMENTAL ANALYSIS DIVISION, 14335 WEST 44TH AVENUE, GOLDEN, COLORADO 80401, PHONE: 303-278-9521



To: Mr. Roy A. Belletto
Acurex Corporation
485 Clyde Avenue
Mountain View, CA 94042

Date: October 12, 1981

Release No. 6 Exhibit A
P. O. No.: Subcontract No. SW59159A

Analyst: J. Oldham

Sample No.: AB1-07-011-562 SPARK SOURCE MASS SPECTROGRAPHIC ANALYSIS IAD No.: 97-H437-116-13

TOSCO 2 XAO

CONCENTRATION IN PPM WEIGHT

ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.
Uranium		Terbium		Ruthenium		Vanadium	0.2
Thorium		Gadolinium		Molybdenum	0.9	Titanium	2
Bismuth		Europium		Niobium		Scandium	<0.1
Lead	0.2	Samarium		Zirconium	1	Calcium	150
Thallium		Neodymium		Yttrium		Potassium	250
Mercury	NR	Praseodymium		Strontium	0.5	Chlorine	15
Gold		Cerium		Rubidium	<0.1	Sulfur	74
Platinum	3	Lanthanum		Bromine	5	Phosphorus	8
Iridium		Barium	3	Selenium		Silicon	170
Osmium		Cesium	<0.1	Arsenic	NR	Aluminum	20
Rhenium		Iodine	0.3	Germanium		Magnesium	3
Tungsten		Tellurium		Gallium	<0.1	Sodium	>150
Tantalum		Antimony	NR	Zinc	4	Fluorine	=0.2
Hafnium		Tin		Copper	8	Oxygen	NR
Lutetium		Indium	STD	Nickel	1	Nitrogen	NR
Ytterbium		Cadmium	0.1	Cobalt	<0.1	Carbon	NR
Thulium		Silver		Iron	32	Boron	<0.1
Erbium		Palladium		Manganese	0.3	Beryllium	
Holmium		Rhodium		Chromium	5	Lithium	<0.1
Dysprosium						Hydrogen	NR

STD - Internal Standard
NR - Not Reported
All elements not detected < 0.1 ppm
MC - Major Component
INT - Interference

Approved:

COMMERCIAL TESTING & ENGINEERING CO.

Reply to

GENERAL OFFICES: 228 NORTH LA SALLE STREET, CHICAGO, ILLINOIS 60601 - AREA CODE 312 728-8434
INSTRUMENTAL ANALYSIS DIVISION, 14335 WEST 44TH AVENUE, GOLDEN, COLORADO 80401, PHONE: 303-278-9521



To: Mr. Roy Belletto
Acurex Corporation
485 Clyde Avenue
Mountain View, CA 94042

Date: October 15, 1981

Release No. 6 Exhibit A
P. O. No. Subcontract No. SW59159A

Analyst: J. Oldham

Sample No.: A81-07-011-580 SPARK SOURCE MASS SPECTROGRAPHIC ANALYSIS IAD No.: 97-H437-116-13

TOSCO XRD BLANK CONCENTRATION IN PPM WEIGHT

ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.
Uranium		Terbium		Ruthenium		Vanadium	<0.1
Thorium		Gadolinium		Molybdenum	0.3	Titanium	3
Bismuth		Europium		Niobium	<0.1	Scandium	<0.1
Lead	0.3	Samarium		Zirconium		Calcium	71
Thallium		Neodymium		Yttrium		Potassium	15
Mercury	NR	Praseodymium		Strontium	1	Chlorine	1
Gold		Cerium		Rubidium		Sulfur	4
Platinum	*44	Lanthanum	<0.1	Bromine	0.6	Phosphorus	1
Iridium		Barium	*1	Selenium		Silicon	5
Osmium		Cesium		Arsenic	NR	Aluminum	3
Rhenium		Iodine	<0.1	Germanium		Magnesium	1
Tungsten		Tellurium		Gallium	<0.1	Sodium	3
Tantalum		Antimony	NR	Zinc	*5	Fluorine	<0.1
Hafnium		Tin		Copper	2	Oxygen	NR
Lutetium		Indium	STD	Nickel	5	Nitrogen	NR
Ytterbium		Cadmium		Cobalt	<0.1	Carbon	NR
Thulium		Silver		Iron	3	Boron	<0.1
Erbium		Palladium		Manganese	0.2	Beryllium	
Holmium		Rhodium		Chromium	0.3	Lithium	0.1
Dysprosium		*Heterogeneous				Hydrogen	NR

STD — Internal Standard

NR — Not Reported

All elements not detected < 0.1 ppm

MC — Major Component

INT — Interference

Approved:

COMMERCIAL TESTING & ENGINEERING CO.

Reply to

GENERAL OFFICES: 226 NORTH LA SALLE STREET, CHICAGO, ILLINOIS 60601 • AREA CODE 312 728-8434
INSTRUMENTAL ANALYSIS DIVISION, 14335 WEST 44TH AVENUE, GOLDEN, COLORADO 80401, PHONE: 303-278-9521



To: Mr. Roy A. Belletto
Acurex Corporation
485 Clyde Avenue
Mountain View, CA 94042

Date: October 9, 1981

Release No. 6 Exhibit A
P. O. No-Subcontract No. SW59159A

Analyst: J. Oldham

Sample No.: A81-07-011-539 SPARK SOURCE MASS SPECTROGRAPHIC ANALYSIS IAD No.: 97-H437-116-13

TOSCO 4 IMP 1

CONCENTRATION IN $\mu\text{g/mL}$

ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.
Uranium	<0.005	Terbium		Ruthenium		Vanadium	0.001
Thorium	<0.006	Gadolinium		Molybdenum	0.006	Titanium	0.08
Bismuth		Europium		Niobium	0.004	Scandium	<0.001
Lead	0.006	Samarium		Zirconium	0.001	Calcium	0.8
Thallium		Neodymium		Yttrium		Potassium	0.8
Mercury	NR	Praseodymium		Strontium	0.005	Chlorine	4
Gold		Cerium		Rubidium	0.003	Sulfur	MC
Platinum		Lanthanum		Bromine	0.01	Phosphorus	0.1
Iridium		Barium	0.03	Selenium	<0.008	Silicon	9
Osmium		Cesium	0.002	Arsenic	NR	Aluminum	0.02
Rhenium		Iodine	0.003	Germanium		Magnesium	0.03
Tungsten		Tellurium		Gallium	0.002	Sodium	MC
Tantalum		Antimony	NR	Zinc	0.04	Fluorine	=2
Hafnium		Tin	0.003	Copper	0.04	Oxygen	NR
Lutetium		Indium	STD	Nickel	0.09	Nitrogen	NR
Ytterbium		Cadmium		Cobalt	<0.001	Carbon	NR
Thulium		Silver	*0.7	Iron	0.2	Boron	0.003
Erbium		Palladium		Manganese	0.01	Beryllium	
Holmium		Rhodium		Chromium	0.02	Lithium	<0.001
Dysprosium		*Heterogeneous				Hydrogen	NR

STD - Internal Standard

NR - Not Reported

All elements not detected $\leq 0.001 \mu\text{g/mL}$

MC - Major Component $> 10 \mu\text{g/mL}$

INT - Interference

Approved:

COMMERCIAL TESTING & ENGINEERING CO.

Reply to

GENERAL OFFICES: 226 NORTH LA SALLE STREET, CHICAGO, ILLINOIS 60601 • AREA CODE 312 726-8434
INSTRUMENTAL ANALYSIS DIVISION, 14335 WEST 44TH AVENUE, GOLDEN, COLORADO 80401, PHONE: 303-278-9521



To: Mr. Roy A. Belletto
Aurex Corporation
485 Clyde Avenue
Mountain View, CA 94042

Date: October 12, 1981

Release No. 6 Exhibit A
P. O. No.: Subcontract No. SW59159A

Analyst: J. Oldham

Sample No.: A81-07-011-554 SPARK SOURCE MASS SPECTROGRAPHIC ANALYSIS IAD No.: 97-H437-116-13

TOSCO 1 IMP Δ BLANK CONCENTRATION IN µg/mL

ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.
Uranium	<0.01	Terbium		Ruthenium		Vanadium	0.004
Thorium	<0.02	Gadolinium		Molybdenum	*0.02	Titanium	0.02
Bismuth		Europium		Niobium		Scandium	<0.001
Lead	0.009	Samarium		Zirconium	0.003	Calcium	1
Thallium		Neodymium		Yttrium		Potassium	1
Mercury	NR	Praseodymium		Strontium	0.004	Chlorine	*3
Gold		Cerium		Rubidium	<0.001	Sulfur	0.3
Platinum	0.008	Lanthanum		Bromine	0.04	Phosphorus	0.8
Iridium		Barium	0.009	Selenium		Silicon	*0.7
Osmium		Cesium		Arsenic	NR	Aluminum	0.09
Rhenium		Iodine		Germanium		Magnesium	≤0.02
Tungsten		Tellurium		Gallium		Sodium	0.2
Tantalum		Antimony	NR	Zinc	0.01	Fluorine	=0.002
Hafnium		Tin	0.02	Copper	0.01	Oxygen	NR
Lutetium		Indium	STD	Nickel	0.008	Nitrogen	NR
Ytterbium		Cadmium		Cobalt	<0.001	Carbon	NR
Thulium		Silver	0.8	Iron	0.05	Boron	<0.001
Erbium		Palladium		Manganese	0.001	Beryllium	
Holmium		Rhodium		Chromium	0.01	Lithium	<0.001
Dysprosium		*Heterogeneous				Hydrogen	NR

STD — Internal Standard

NR — Not Reported

All elements not detected < 0.002 µg/mL

MC — Major Component > 10 µg/mL

INT — Interference

Approved:

COMMERCIAL TESTING & ENGINEERING CO.

Reply to

GENERAL OFFICES: 228 NORTH LA'SALLE STREET, CHICAGO, ILLINOIS 60601 · AREA CODE 312 726-8434
INSTRUMENTAL ANALYSIS DIVISION, 14335 WEST 44TH AVENUE, GOLDEN, COLORADO 80401, PHONE: 303-278-9521



To: Mr. Roy Belletto
Acurex Corporation
485 Clyde Street
Mountain View, CA 94042

Date: October 14, 1981

Release No. 6 Exhibit A
Subcontract No. SW59159A

Analyst: J. Oldham

P. O. No.: Sample No.: A81-07-011-561 SPARK SOURCE MASS SPECTROGRAPHIC ANALYSIS IAD No.: 97-H437-116-13

TOSCO 2 TMP 1

CONCENTRATION IN $\mu\text{g/mL}$

ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.
Uranium		Terbium		Ruthenium		Vanadium	0.004
Thorium		Gadolinium		Molybdenum	0.009	Titanium	0.06
Bismuth		Europium		Niobium	0.02	Scandium	<0.002
Lead	0.009	Samarium		Zirconium	0.03	Calcium	1
Thallium		Neodymium		Yttrium	0.002	Potassium	1
Mercury	NR	Praseodymium		Strontium	0.02	Chlorine	0.1
Gold		Cerium	0.009	Rubidium	<0.02	Sulfur	MC
Platinum		Lanthanum	0.02	Bromine	<0.03	Phosphorus	0.008
Iridium		Barium	0.08	Selenium	<0.02	Silicon	3
Osmium		Cesium		Arsenic	NR	Aluminum	0.04
Rhenium		Iodine	<0.002	Germanium		Magnesium	0.008
Tungsten		Tellurium	<0.004	Gallium	0.003	Sodium	MC
Tantalum		Antimony	NR	Zinc	0.8	Fluorine	≈0.01
Hafnium		Tin	0.003	Copper	0.2	Oxygen	NR
Lutetium		Indium	STD	Nickel	0.2	Nitrogen	NR
Ytterbium		Cadmium		Cobalt	<0.001	Carbon	NR
Thulium		Silver	0.08	Iron	0.5	Boron	<0.001
Erbium		Palladium		Manganese	0.1	Beryllium	
Holmium		Rhodium		Chromium	0.05	Lithium	<0.001
Dysprosium						Hydrogen	NR

STD - Internal Standard

NR - Not Reported

All elements not detected < 0.001 $\mu\text{g/mL}$

MC - Major Component > 10 $\mu\text{g/mL}$

INT - Interference

Approved:

COMMERCIAL TESTING & ENGINEERING CO.

GENERAL OFFICES: 228 NORTH LA SALLE STREET, CHICAGO, ILLINOIS 60601 - AREA CODE 312 726-8434
INSTRUMENTAL ANALYSIS DIVISION, 14335 WEST 44TH AVENUE, GOLDEN, COLORADO 80401. PHONE: 303-278-9521

Reply to



To: Mr. Roy A. Belletto
Acurex Corporation
485 Clyde Avenue
Mountain View, CA 94042

Date October 12, 1981

Release No. 6 Exhibit A
P. O. No.: Subcontract No. SW59159A

Analyst: J. Oldham

Sample No.: A81-07-011-560 SPARK SOURCE MASS SPECTROGRAPHIC ANALYSIS IAD No.: 97-H437-116-13

TC550 2 IMP 1 BLINK CONCENTRATION IN µg/mL

ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.	ELEMENT	CONC.
Uranium	<0.01	Terbium		Ruthenium		Vanadium	0.001
Thorium	<0.01	Gadolinium		Molybdenum	0.008	Titanium	0.02
Bismuth		Europium		Niobium		Scandium	<0.001
Lead		Samarium		Zirconium	0.002	Calcium	0.9
Thallium		Neodymium		Yttrium		Potassium	*MC
Mercury	NR	Praseodymium		Strontium	0.003	Chlorine	2
Gold		Cerium		Rubidium	0.005	Sulfur	0.5
Platinum		Lanthanum		Bromine	0.01	Phosphorus	1
Iridium		Barium	0.04	Selenium		Silicon	5
Osmium		Cesium	0.006	Arsenic	NR	Aluminum	0.5
Rhenium		Iodine	0.002	Germanium		Magnesium	0.07
Tungsten		Tellurium		Gallium	0.003	Sodium	>4
Tantalum		Antimony	NR	Zinc	0.1	Fluorine	=0.05
Hafnium		Tin	0.01	Copper	0.02	Oxygen	NR
Lutetium		Indium	STD	Nickel	0.01	Nitrogen	NR
Ytterbium		Cadmium		Cobalt	<0.002	Carbon	NR
Thulium		Silver	0.03	Iron	0.09	Boron	0.001
Erbium		Palladium		Manganese	0.02	Beryllium	
Holmium		Rhodium		Chromium	0.01	Lithium	0.008
Dysprosium		*Heterogeneous				Hydrogen	NR

STD - Internal Standard

NR - Not Reported

All elements not detected < 0.001 µg/mL

MC - Major Component > 10 µg/mL

INT - Interference

Approved:



DATA REPORTING FORM

CUSTOMER CMEA DATE
 CUSTOMER CONTRACT NO. 307605.92 ACUREX CONTRACT NO. A81-07-011
 RESULTS REPORT TO L. Waterland TELEPHONE
 ADDRESS
TOSCO

SAMPLE ID (CUSTOMER)	Filter-1	XAD-1	Imp 1-1	Imp 2&3-1	Filter-2	XAD-2	Imp 1-2	Imp 2&3-2	NBS Ash*	Fly
SAMPLE ID (LAB)	545	569	539	555	038	562	561	564	033-6	
PARAMETER										UNITS
Hg Aliquot	3	<2	<1, <1	<1	2	<1	<1,<1	<1	10	ug/l
Hg blank Aliquot	4	3	<1	<1	4	3	<1	<1	<1	ug/l
Hg	<0.07	<0.5	<0.07	<0.07	<0.04	<0.2	<0.01	<0.07	0.10 ug/g	ug/dscm
As Aliquot	-	-	-	<20	-	-	-	<20	-	ug/l
As blank	-	-	-	<20	-	-	-	<20	-	ug/l
As	-	-	-	<1	-	-	-	<1	-	ug/dscm
Sb Aliquot	-	-	-	<20	-	-	-	<20	-	ug/l
Sb blank	-	-	-	<20	-	-	-	<20	-	ug/l
Sb	-	-	-	<1	-	-	-	<1	-	ug/dscm

*0.16 mg/g Hg

ANALYST _____

REVIEWER _____

5.7 TOTAL CHROMATOGRAPHABLE ORGANICS (TCO), GRAVIMETRIC ORGANICS (GRAV),
INFRARED (IR) SPECTRA, GAS CHROMATOGRAPHY/MASS SPECTROMETRY (GC/MS)
AND LOW RESOLUTION MASS SPECTROMETRY (LRMS) OF TOTAL SAMPLE EXTRACTS

ACUREX
M.S. 2-2260

September 21, 1981
Acurex ID #A81-07-011
#A81-07-033
Client P.O. 307605.92

Attention: L. Waterland

Sample: 2 SASS Train, received 7/8/81

The above referenced samples were analyzed per level 1 protocol. Arsenic and antimony were determined by furnace AAS.

Polynuclears were determined by a modified EPA method 625. 1 μ l of sample was injected onto a SE-54, J and W 30 meter capillary column using Grob injection. The column was held at 30°C, then ramped at 10°C per minute to 270°C.

Benzo (c) phenanthrene, dibenzo (c,g) carbozole, 7, 12-dimethylbenz (a) anthracene, 3-methyl chloranthene, and perylene were not detected (<40 ng/ μ l injection) in any sample analyzed by GC/MS.

The GCMS analysis of XAD-2 resin spiked with 100 mg of naphthalene, phenanthrene, and pyrene gave a recovery of 33% for naphthalene, 38% for phenanthrene, and <1% for pyrene.

Prepared by:

Greg Nicoll

Greg Nicoll
Program Director

Authorized by:

M. C. Ferguson

M. C. Ferguson
Sample Control Custodian

GN/MCF/kfh

DATA REPORTING FORM

CUSTOMER CMEA DATE _____
 CUSTOMER CONTRACT NO. 307605.92 ACUREX CONTRACT NO. A81-07-011
 RESULTS REPORT TO L. Waterland TELEPHONE _____
 ADDRESS _____
TOSCO

SAMPLE ID (CUSTOMER)	Filter-1	XAD-1 ^a	OMC-1	Filter-2	XAD-2 ^a	OMC-2	XAD-QC	XAD-1 ^b	XAD-2 ^b	
SAMPLE ID (LAB)	545	569	541	038	562	559	033-5	569	567	
PARAMETER										UNITS
GRAV Uncorrected	<6	21	<4	<8	18	6	-			mg
GRAV Blank	<10	10	<4	<10	10	4	-			mg
GRAV	<6	11	<4	<8	8	6				mg
TCO Uncorrected	-	39	<0.04	-	25	0.64	0.74	11	2.3	mg
TCO Blank	-	30	<0.04	-	30	<0.04	<0.02	0.5	0.5	mg
TCO	-	9	<0.04	-	<5	0.64	0.74*	10	1.8	mg
GRAV and TCO	<0.2	0.7	<0.1	<0.3	0.3	0.2	-	0.76	0.35	mg/dscm

*Spiked at 1.00 mg TCO

^aContaminated XAD, high blank

^bSample/blank contamination corrected by GC/MS

ANALYST _____

REVIEWER _____

IR REPORT

SAMPLE:

Tosco Filter Blank 571

Wave Number - (cm^{-1})	Intensity	Assignment	Comments	
		No Peaks		

IR REPORT

SAMPLE:

TOSCO XAD Blank

Wave Number cm^{-1}	Intensity	Assignment	Comments
		No Peaks	

DATA REPORTING FORM

CUSTOMER CMEA DATE _____
 CUSTOMER CONTRACT NO. 307605.92 ACUREX CONTRACT NO. A81-07-011
 RESULTS REPORT TO L. Waterland TELEPHONE _____
 ADDRESS _____
TOSCO

SAMPLE ID (CUSTOMER)	XAD-1	XAC-2	XAD-2 Dup		XAD-1	XAD-2	XAD-2 Dup			
SAMPLE ID (LAB)	569	562	562DS		569	562	562DS			
PARAMETER										UNITS
Phenol	26	6	14		1.0	0.2	0.55			
Naphthalene (Blank-corrected)	<1	9	20		<0.04	0.4	.78			
1,3-Dichlorobenzene	2	<1	<1		0.08	<0.04	<0.04			
1,4-Dichlorobenzene	1	1	2		0.04	0.04	0.08			
1,2-Dichlorobenzene	3	<1	<1		0.1	<0.04	<0.04			
Nitrobenzene	4	1	2		0.2	0.04	0.08			
2-Nitrophenol	<5	11	10		<0.2	0.43	0.39			
Diphenylamine	3	2	2		0.1	0.08	0.08			
Azobenzene	36	<1	1		1.4	<0.04	0.04			
Phenanthrene	30	3	4		1.2	0.1	0.2			
2,6-Dinitrotoluene	<1	3	<1		<0.04	0.1	<0.04			
Other Polynuclears	<1	<1	<1		<0.04	<0.04	<0.04			
Units	ng/uL	ng/uL	ng/uL		ug/dscm	ug/dscm	ug/dscm			

ANALYST _____

REVIEWER _____

ACUREX
Corporation
LYSIS LABORATORIES

DATA REPORTING FORM

CUSTOMER CMEA DATE
CUSTOMER CONTRACT NO. 307605.92 ACUREX CONTRACT NO. A81-07-011
RESULTS REPORT TO L. Waterland TELEPHONE
ADDRESS
TOSCO

SAMPLE ID (CUSTOMER)	Filter-1	Filter-2	Filter B1k	OMC-1	OMC-2	XAD B1k					
SAMPLE ID (LAB)	545	038	571	541	559	580					
PARAMETER											UNITS
Naphthalene	<1	<1	<1	<1	<1	20					ng/uL
Other Polynuclears	<1	<1	<1	<1	<1	<1					ng/uL
Naphthalene	<0.05	<0.08	-	<0.04	<0.04	-					ng/dscm
Other Polynuclears	<0.05	<0.08	-	<0.04	<0.04	-					ng/dscm

ANALYST _____

REVIEWER _____

ORGANICS ANALYSIS DATA SHEET

LABORATORY NAME Acurex

GCMS Detection Limits

ACID COMPOUNDS

	ng
21A 2,4,6-trichlorophenol	5
22A p-chloro-m-cresol	5
24A 2-chlorophenol	5
31A 2,4-dichlorophenol	5
34A 2,4-dimethylphenol	5
57A 2-nitrophenol	5
58A 4-nitrophenol	20
59A 2,4-dinitrophenol	20
60A 4,6-dinitro-o-cresol	20
64A pentachlorophenol	5
* 65A phenol	1

BASE/NEUTRAL COMPOUNDS

1B acenaphthene	1
5B benzidine	20
8B 1,2,4-trichlorobenzene	1
9B hexachlorobenzene	1
12B hexachloroethane	1
18B bis(2-chloroethyl)ether	1
20B 2-chloronaphthalene	1
25B 1,2-dichlorobenzene	1
26B 1,3-dichlorobenzene	1
27B 1,4-dichlorobenzene	1
28B 3,3'-dichlorobenzidine	5
33B 2,4-dinitrotoluene	1
36B 2,6-dinitrotoluene	1
37B 1,2-diphenylhydrazine (as azobenzene)	1
39B fluoranthene	1
40B o-chlorophenyl phenyl ether	1

BASE/NEUTRAL COMPOUNDS

	ng
41B 4-bromophenyl phenyl ether	1
42B bis(2-chloroisopropyl) ether	1
43B bis(2-chloroethoxy) methane	1
52B hexachlorobutadiene	1
53B hexachlorocyclopentadiene	1
54B isophorone	1
* 55B naphthalene	1
56B nitrobenzene	1
61B N-nitrosodimethylamine	NA
62B N-nitrosodiphenylamine	1
63B N-nitrosodi-n-propylamine	5
* 66B bis(2-ethylhexyl) phthalate	1
* 67B butyl benzyl phthalate	1
* 68B di-n-butyl phthalate	1
* 69B di-n-octyl phthalate	1
* 70B diethyl phthalate	1
* 71B dimethyl phthalate	1
72B benzo(a)anthracene	1
73B benzo(a)pyrene	1
74B 3,4-benzofluoranthene	1
75B benzo(k)fluoranthene	1
76B chrysene	1
77B acenaphthylene	1
78B anthracene	1
79B benzo(ghi)perylene	5
80B fluorene	1
81B phenanthrene	1
82B dibenzo(a,h)anthracene	5
83B indeno(1,2,3-cd)pyrene	5
84B pyrene	1

* Normally found in laboratory blanks



Energy & Environmental Division

CMEA/ACUREX

December 4, 1981

Acurex ID#: A81-10-011, A81-10-022

Client P.O.#: 307605

ATTENTION: L. Waterland

Samples: 9 extracts from Tosco and Ethan Allen

The above referenced samples were analyzed by direct probe mass spectrometry. Searches have been made for classes of compounds most likely to be found in the various LC fractions, according to procedures described in the "IERL-RTP Procedures Manual: Level 1 Environmental Assessment". The following fragment ions used for search are given below:

Compound Class	Fragment ions (m/e-)
Polycyclic aromatic hydrocarbons	178,202,216,228,252,276
Aliphatic hydrocarbons	57,71
Halogenated aliphatics	79,81,93,95,107,109,49,63
Aromatic hydrocarbons	50,51,77,78,79,91
Ethers	45,59,73
Alcohols	45,59,61,73,75
Phenols	51,77,94
Nitriles	54,68,82
Phthalate esters	61,59,71,87
Amines	44,58
Ketones	51,71
N-heterocyclics	117,167,129,179
Mercaptans, sulfides	47,61,75
Benzothiophenes	57,58,59,69,70,85,97,111,125
Carboxylic acids	60,73,149
Amides	58,72,86,100

To test the analysis procedure, a standard mixture containing ethers, amines, polycyclic aromatic hydrocarbons, nitrosamines, phenols, etc., was analyzed under identical conditions as the samples. Losses of the very volatile compounds such as naphthalene, bis(2-chloroethyl)ether, low molecular weight nitrosamines were observed, however the higher molecular weight compounds in a particular class were recovered.

Prepared by:

Greg Nicoll
Greg Nicoll
Program Director

Approved by:

Viorica Lopez-Avila
Viorica Lopez-Avila, Ph.D.
Technical Director

GN/VLA:es

LRMS REPORT

SAMPLE: Toxco I XAD 569

Major Categories

Intensity	Category	MW Range
100	Ethers	
100	Heterocyclic sulfur compounds	
100	Carboxylic acids	
10	Halogenated aliphatics	
10	Aromatic hydrocarbons	
10	Nitriles	

Sub-Categories, Specific Compounds

Intensity	Category	m/e	Composition

Other

LRMS REPORT

SAMPLE: Tosco I XAD 569 (cont)

Major Categories

Intensity	Category	MW Range
10	Alcohols	
10	Heterocyclic nitrogen compounds	

Sub-Categories, Specific Compounds

Intensity	Category	m/e	Composition

Other

LRMS REPORT

SAMPLE: Tesco II XAD 562 and OMC 599

Major Categories

Intensity	Category	MW Range
100	Aliphatic hydrocarbons	
100	Amines	
10	Aromatic hydrocarbons	
10	Carboxylic acids	

Sub-Categories, Specific Compounds

Intensity	Category	m/e	Composition

Other

5.8 C₁-C₆ CHROMATOGRAMS

TOSCO

Run	Time	Bulb	C ₁	C ₂	C ₃	C ₄	C ₅	C ₆
6/18/81								
Sample 7	1330	A	5.2	ND ^a	ND	ND	ND	ND
Sample 8	1335	B	0.9	ND	0.8	ND	ND	ND
Sample 9	1330	A	1.3	ND	3.0	4.9	0.2	ND
Sample 10	1335	B	ND	ND	1.2	0.2	ND	ND
Cal 11	1627	--	17.0	10.3	10.6	10.8	10.4	10.1
Blank 12	1649	B	2.0	0.3	ND	ND	ND	0.4
Blank 13	1702	A	2.1	ND	ND	ND	ND	ND
Sample 14	1740	B	1.2	0.3	1.5	ND	ND	ND
Sample 15	1750	A	ND	ND	1.4	ND	0.9	ND
Cal 16	1920	--	17.0	10.3	10.6	10.8	10.4	10.1
Sample 17	1945	A	2.5	7.8	5.6	7.0	0.7	ND
Sample 18	1950	B	0.9	8.0	5.6	7.3	ND	ND
Sample 19	1945	A	1.2	3.7	1.6	2.0	2.4	0.2
Sample 20	2055	A	2.3	4.3	1.4	0.9	ND	ND
Blank 24	2155	A	2.1	ND	ND	0.2	ND	ND
Blank 25	2205	B	2.0	ND	ND	ND	ND	ND
6/19/81								
Cal 27,28,29			17.0	10.3	10.6	10.8	10.4	10.1
Sample 30	1110	A	1.1	1.1	0.1	ND	ND	ND
Sample 31	1115	B	1.1	1.8	ND	ND	ND	ND
Sample 32	1115	B	1.1	1.6	ND	0.1	ND	ND
Sample 33	1400	A	1.3	1.8	ND	ND	ND	ND
Sample 34	1405	B	1.0	1.1	ND	ND	ND	ND
b								
Cal 2	1624	--	17.0	10.3	10.6	10.8	10.4	10.1
Sample 3	1645	A	1.5	3.0	ND	0.2	ND	ND
Sample 4	1650	B	0.7	3.0	ND	ND	ND	ND
b								
Cal 1	1621	--	17.1	10.3	10.6	10.8	10.4	10.1
Blank 2	1819	B	1.6	ND	1.4	ND	ND	ND
Blank 3	1831	A	1.3	ND	ND	ND	ND	ND

^aAll values \pm 10 percent, less than 0.5 ppm.

^bLost power

Note: All values calculated by averaging the before and after (where applicable) calibration amount/area. Values less than 0.1 ppm are ND. Tests run at 110°C, 26 psi through two Varian model 3700 GC packed with 6 ft of Super Q (see appendix A of Volume I for more detail).

TOSCO 6-18-81

	RT	Amount/Area	PPM
<u>Run 1 (Recal B) C₁-C₆</u>			
C ₁	0.26	9.1123 x 10 ⁻⁴	17.0
C ₂	0.42	3.4823 x 10 ⁻⁴	10.3
C ₃	0.72	2.2827 x 10 ⁻⁴	10.6
C ₄	1.46	1.6147 x 10 ⁻⁴	10.8
C ₅	3.28	1.3578 x 10 ⁻⁴	10.4
C ₆	7.88	1.1422 x 10 ⁻⁴	10.1
<u>Run 2 Blank B 1819</u>			
C ₁	0.22	1,728	1.6
C ₃	0.75	6,076	1.4
<u>Run 2 (Recalibrated B) C₁-C₆</u>			
C ₁	0.26	8.9310 x 10 ⁻⁴	17.0
C ₂	0.41	3.4392 x 10 ⁻⁴	10.3
C ₃	0.71	2.1631 x 10 ⁻⁴	10.6
C ₄	1.45	1.4770 x 10 ⁻⁴	10.8
C ₅	3.28	1.2880 x 10 ⁻⁴	10.4
C ₆	7.87	1.1383 x 10 ⁻⁴	10.1
<u>Run 3 Blank A 1831</u>			
C ₁	0.22	1,441	1.3
<u>Run 3 Sample A 1645</u>			
C ₁	0.23	1,649	1.5
C ₂	0.47	8,656	3.0
C ₃	0.96	1,610	0.2
<u>Run 4 Sample B 1650</u>			
C ₁	0.23	764	0.7
C ₂	0.47	8,548	3.0
C ₃	0.96	613	0.09

TOSCO 6-18-81 (Continued)

	RT	Amount/Area	PPM
<u>Run 6 Calibrated C₁-C₆</u>			
C ₁	0.24	9.2372 x 10 ⁻⁴	17.0
C ₂	0.41	3.4661 x 10 ⁻⁴	10.3
C ₃	0.70	2.2535 x 10 ⁻⁴	10.6
C ₄	1.42	1.5594 x 10 ⁻⁴	10.8
C ₅	3.22	1.3019 x 10 ⁻⁴	10.4
C ₆	7.74	1.0560 x 10 ⁻⁴	10.1
<u>Run 7 Sample A 1330</u>			
C ₁	0.22	5,888	5.2
<u>Run 8 Sample B 1335</u>			
C ₁	0.22	1,069	0.9
C ₃	0.47	4,126	0.8
<u>Run 9 Sample A 1330</u>			
C ₁	0.22	1,572	1.3
C ₃	0.46	15,299	3.0
C ₄	0.96	26,831	4.9
	1.33	10,875	
C ₅	2.89	1,579	0.2
<u>Run 10 Sample B 1335</u>			
C ₁	0.22	0	0
C ₃	0.46	6,388	1.2
C ₄	0.97	371	0.2
	1.34	867	
<u>Run 11 C₁-C₆ (Recalibrated)</u>			
C ₁	0.23	8.3600 x 10 ⁻⁴	17.0
C ₂	0.41	3.0853 x 10 ⁻⁴	10.3
C ₃	0.70	1.6758 x 10 ⁻⁴	10.6
C ₄	1.42	1.0577 x 10 ⁻⁴	10.8
C ₅	3.23	1.2584 x 10 ⁻⁴	10.4
C ₆	7.76	2.1048 x 10 ⁻⁴	10.1

TOSCO 6-18-81 (Continued)

	RT	Amount/Area	PPM
<u>Run 12 Blank Bulb B 1649</u>			
C ₁	0.22	2,286	2.0
C ₂	0.37	991	0.3
C ₆	3.82 4.99 5.62	1,231 1,188 274	0.4
<u>Run 13 Blank Bulb A 1702</u>			
C ₁	0.22	2,366	2.1
C ₄	0.91	57	0.01
<u>Run 14 Sample B 1740</u>			
C ₁	0.22	1,371	1.2
C ₂	0.31	951	0.3
C ₃	0.46	7,432	1.5
<u>Run 15 Sample A 1750</u>			
C ₁	0.23	0	0
C ₃	0.47	6,885	1.4
C ₅	1.75	6,903	0.9
LOST POWER			
<u>Run 16 (Recalibrated B) 1920</u>			
C ₁	0.25	9.2447 x 10 ⁻⁴	17.0
C ₂	0.42	3.5481 x 10 ⁻⁴	10.3
C ₃	0.71	2.3173 x 10 ⁻⁴	10.6
C ₄	1.45	1.5875 x 10 ⁻⁴	10.8
C ₅	3.27	1.3516 x 10 ⁻⁴	10.4
C ₆	7.86	1.2190 x 10 ⁻⁴	10.1
<u>Run 17 Sample A 1945</u>			
C ₁	0.24	2,748	2.5
C ₂	0.38	9,701	7.8
	0.46	12,396	
C ₃	0.69	24,034	5.6

TOSCO 6-18-81 (Continued)

	RT	Amount/Area	PPM
<u>Run 17 Sample A 1945 Cont.</u>			
C ₄	0.96	17,404	7.0
	1.41	26,507	
C ₅	2.30	196	0.7
	2.55	1,499	
	2.72	865	
	2.95	1,194	
	2.99	1,530	
C ₆	6.03	592	0.07
<u>Run 18 Sample B 1950</u>			
C ₁	0.22	1,000	0.9
C ₂	0.38	9,514	8.0
	0.45	13,108	
C ₃	0.68	24,088	5.6
C ₄	0.95	18,656	7.3
	1.37	27,033	
<u>Run 19 Sample A 1945</u>			
C ₁	0.21	1,312	1.2
C ₂	0.38	3,032	3.7
	0.45	7,373	
C ₃	0.69	7,114	1.6
C ₄	0.95	2,921	2.0
	1.43	9,483	
C ₅	2.14	8,550	2.4
	3.31	9,376	
C ₆	5.00	1,717	0.2
>C ₆	8.63	748	--
<u>Run 20 Sample A 2055</u>			
C ₁	0.20	2,526	2.3
C ₂	0.37	3,346	4.3
	0.46	8,645	
C ₃	0.69	6,058	1.4
C ₄	0.95	2,888	0.9
	1.37	2,537	
C ₆	4.0	0	0
<u>Run 21, 22, 23 Flame Out</u>			

TOSCO 6-19-81 (Continued)

	RT	Amount/Area	PPM
<u>Run 24 Blank A 2155 Zero Air</u>			
C ₁	0.22	2,283	2.1
C ₄	1.18	1,277	0.2
C ₅	2.38	658	0.09
<u>Run 25 Blank B 2205 Zero Air</u>			
C ₁	0.22	2,189	2.0
>C ₆	9.25	174	--
<u>Run: 27, 28, and 29 (Recalibrated B) C₁-C₆)</u>			
C ₁	0.25	9.6424 x 10 ⁻⁴	17.0
C ₂	0.40	3.5852 x 10 ⁻⁴	10.3
C ₃	0.69	2.3319 x 10 ⁻⁴	10.6
C ₄	1.43	1.5990 x 10 ⁻⁴	10.8
C ₅	3.26	1.3517 x 10 ⁻⁴	10.4
C ₆	7.82	1.1796 x 10 ⁻⁴	10.1
<u>Run 31 Sample B 1115</u>			
C ₁	0.22	1,220	1.1
C ₂	0.38	666	1.8
	0.47	4,480	
<u>Run 32 Sample B 1115</u>			
C ₁	0.22	1,220	1.1
C ₂	0.37	680	1.6
	0.47	3,754	
C ₄	0.96	665	0.1
<u>Run 33 Sample A 1400</u>			
C ₁	0.22	1,409	1.3
C ₂	0.46	5,096	1.8
<u>Run 34 Sample B 1405</u>			
C ₁	0.22	1,101	1.0
C ₂	0.32	331	1.1
	0.46	2,729	
LOST POWER			

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location Tesco, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 6-18-81, Time 3:55:57, Instrument ID VARIAN 3700
 Recorder/Printout Reference No. 7, Recorder ID HP3390A
 Purpose of Run C₁-C₆ HYDROCARBON ANALYSIS

Sample Description 1320 3124

GC CONDITIONS

Amount Injected 2.0ML, Inj. Port or Sample Loop Used 2.0ML LOOP
 Detector Used: FID X, ECD , FPD , TCD (Current)
 Detector Attenuation 4, Amplifier or Range 10⁻¹¹
 Column: Liquid Phase , Solid Phase PORAPAK Q,
 Length 6', O.D. 1/8", I.D. , Material S.S.
 Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C
 Temperature Program ISOTHERMAL

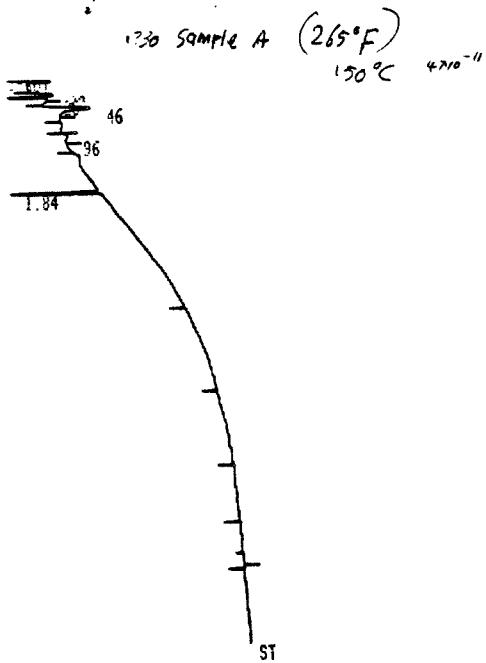
SAMPLE RUN

Sampling Method GRAB 500 ml S.S. BULB

RT	Area	Peak Height	Amount (ppm)	Component
<u>22</u>	<u>5588</u>		<u>5.2</u>	<u>C₁</u>

Name of Operator M.D. CHIPS, Date 9-1 1981

Sample A taken 1330 30PSI 312°F
 Sample B taken 1336 30PSI 275°F



Operator..M.D.C.H.P.S.	Date..6-14-81
Column No.....	Length...6'
Coating.....	Dia...1/8"
Support..PORAPAK Q	Concn.....
TEMP: Col: Init. 1.0 °C	Mesh 2/60
Rate..... °C/min.	Final. 1.0 °C
CARRIER GAS..N ₂	Inj. 1.20 °C
Pressures: Inlet.	Rate. 470 psi/ml.
Hydrogen 1.0 Psi.	Outlet.
DETECTOR E.C. T.C.	Air 45 psi/ml.
Scavenger.....	F.I.D. 2.10 ml/min.
Sens.....	Rate..... ml/min.
SAMPLE 1330 B.G.C.A.	Rec.Range. mV.
Solvent.....	Size 2.0 ml.
	Concn.....

RUN # 7 JUN/18/81 13:55:57

ESTD	RT	AREA	TYPE	CAL#	AMOUNT
	0.22	5888	BP	1R	5.439

TOTAL AREA= 5888
 MUL FACTOR= 1.0000E+00

RUN # 7 JUN/18/81 13:55:57

ESTD	RT	AREA	TYPE	CAL#	AMOUNT
	0.22	5888	BP	1R	5.439

TOTAL AREA= 5888
 MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location Tosco, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 6-18-81, Time 5:30:00, Instrument ID VARIAN 3700

Recorder/Printout Reference No. 8, Recorder ID HP3390A

Purpose of Run C₁-C₆ HYDROCARBON ANALYSIS

Sample Description 325 Case-B

GC CONDITIONS

Amount Injected 2.0 mL, Inj. Port or Sample Loop Used 2.0 mL Loop

Detector Used: FID X, ECD , FPD , TCD (Current)

Detector Attenuation 4, Amplifier or Range 10⁻¹¹

Column: Liquid Phase , Solid Phase PORAPAK Q,

Length 6', O.D. 1/8", I.D. , Material S.S.

Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C

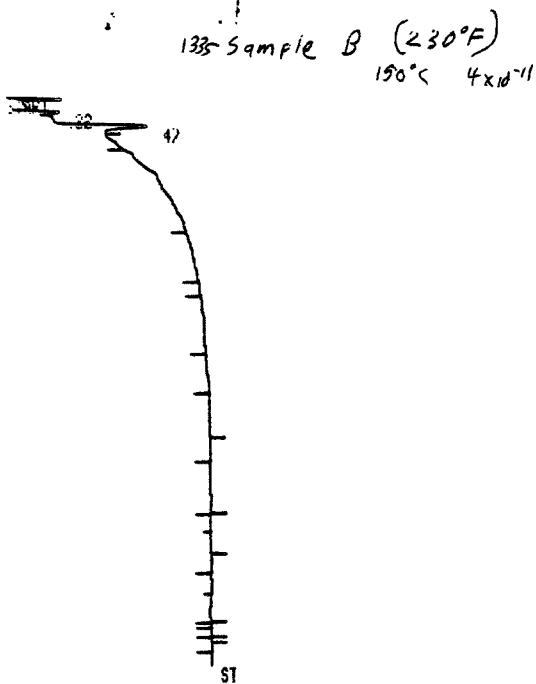
Temperature Program ISOTHERMAL

SAMPLE RUN

Sampling Method GRAB - 500 mL S.S. BULB

RT	Area	Peak Height	Amount (ppm)	Component
.22	1069		0.7	C ₁
.47	4136		0.8	C ₂

Name of Operator M.D. CHIPS, Date 9-1 1981



Operator	M.D.C.H.P.S.	Date	6-18-81
Column No.		Length	6'
Coating	P.D.R. ARALK.	Dia.	1/8"
Support	L	Concn.	Mesh. 60/80
TEMP:	Col: 150°C	Final I.	150°C
Rate	$^{\circ}\text{C}/\text{min.}$	Det.	250 $^{\circ}\text{C}$
CARRIER GAS	H_2	Inj.	1.20 $\mu\text{l.}$
Pressures:	Inlet	Rate	6.0. Psi. ml./min.
Hydrogen	4.0×10^{-11} ml./min.	Outlet	Air. 6.0×10^{-11} ml./min.
DETECTOR E.C.	T.C.	F.I.D.	4.0×10^{-11} mv.
Scavenger		Rate	
Sens.		Rec. Range	... ml./min.
SAMPLE	1335 BULL-B	Size	2.0 ml.
Solvent		Concn.	

RUN # 8 JUN/18/81 15:40:06

ESTD	RT	AREA	TYPE	CAL#	AMOUNT
	0.22	1069	PY	1R	0.988

TOTAL AREA= 1069
 MUL FACTOR= 1.0000E+00

OP # 6 @
 RPRT UNC PKS YES

RUN # 8 JUN/18/81 15:40:06

ESTD	RT	AREA	TYPE	CAL#	AMOUNT
	0.22	1069	PY	1R	0.988
	0.47	4126	VB		0.000

TOTAL AREA= 5195
 MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location Tosco, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 6-18-81, Time 1:12:30, Instrument ID VARIAN 3700
 Recorder/Printout Reference No. 7, Recorder ID HP3390A
 Purpose of Run C₁-C₆ HYDROCARBON ANALYSIS

Sample Description LIBC 700R-A

GC CONDITIONS

Amount Injected 2.0mL, Inj. Port or Sample Loop Used 2.0mL Loop
 Detector Used: FID X, ECD , FPD , TCD (Current)
 Detector Attenuation 4, Amplifier or Range 10⁻¹¹
 Column: Liquid Phase , Solid Phase PORAPAK Q,
 Length 6', O.D. 1/8", I.D. , Material S.S.
 Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C
 Temperature Program ISOTHERMAL

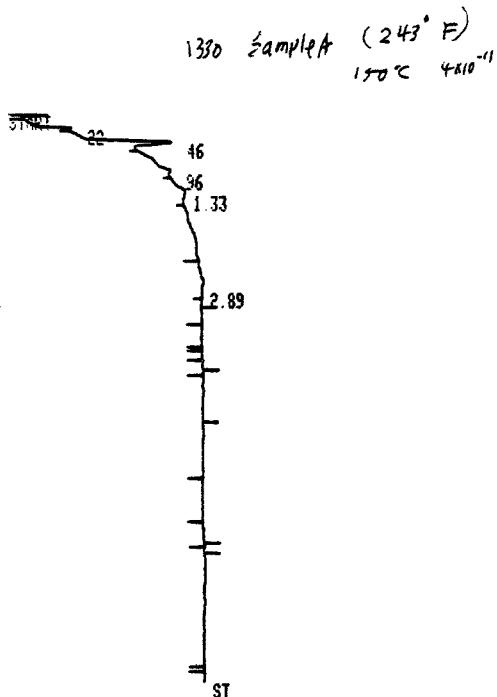
SAMPLE RUN

Sampling Method GRAB .500 ml S.S. BULB

RT	Area	Peak Height	Amount (ppm)	Component
.32	1572		1.3	C ₁
.46	15277		3.0	C ₂
.72	26931	\	4.7	C ₃
1.22	10875	/	/	C ₄
2.31	1277		0.2	C ₅

Name of Operator M.D. CHIPS, Date 9-1 1981

LIST: ZERO = 0, 0.0



Operator	M.D.Q.MIPS	Date	6-18-81
Column No.	6'	Length	6'
Coating	Conc.	Dia.	1/8"
Support	PORAPAK Q	Mesh	60/80
TEMP:	Col: 150°C	Init. Temp	1.0 °C
Rate:	Det. 25 °C	Final Temp	1.0 °C
CARRIER GAS	N ₂	Rate, N ₂ , psi	ml/min.
Pressures:	Inlet	Outlet	
Hydrogen, O ₂ , P _{S₂}	ml/min.	Air, N ₂ O, P _{S₂}	ml/min.
DETECTOR E.C.	T.C.	F.I.D. T.A.D.	A
Scavenger	Rate	ml/min.	
Sens.	Rec. Range	mV	
SAMPLE	1330	BULB-A	Size, 2.0mL
Solvent	Concn.		

RUN # 9 JUN/18/81 15:53:30

ESTD	RT	AREA	TYPE	CAL#	AMOUNT
	0.22	1572	PY	1R	1.452
	0.46	15299	VY		0.000
	0.96	26831	VY		0.000
	1.33	10875	VY	4R	1.696
	2.89	1579	VY		0.000

TOTAL AREA= 56156
MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location Tosco, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 6-18-81, Time 1655:17, Instrument ID VARIAN 3700

Recorder/Printout Reference No. 1D, Recorder ID HP3390A

Purpose of Run C₁-C₆ HYDROCARBON ANALYSIS

Sample Description 1225 - BULB-B

GC CONDITIONS

Amount Injected 2.0ml, Inj. Port or Sample Loop Used 2.0ml Loop

Detector Used: FID X, ECD , FPD , TCD (Current)

Detector Attenuation 4, Amplifier or Range 10⁻¹¹

Column: Liquid Phase , Solid Phase PORAPAK Q,

Length 6', O.D. 1/8", I.D. , Material S.S.

Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C

Temperature Program ISOTHERMAL

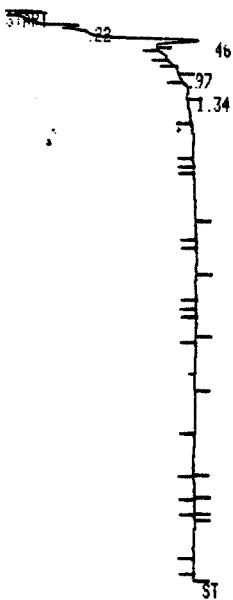
SAMPLE RUN

Sampling Method GRAB - 500 ml S.S. BULB

RT	Area	Peak Height	Amount (ppm)	Component
.22	0		0	C ₁
.46	6338		1.2	C ₃
.77	371	\	0.2	C ₂
1.34	1367	/		C ₄

Name of Operator M.D.Chips, Date 9-1 1981

1335 Sample B 243°F
1578 0.250-11



Operator...M.D.Chips.....	Date...6-18-81.....
Column No.....	Length...6'
Coating.....	Concn.....
Support...P.R.A.PAK. Q.....	Mesh 60/60
TEMP: Col: Init. 150 °C	Final 150 °C
Rate..... °C/min.	Det. 2.5D °C
CARRIER GAS...H ₂	Rate 40 P.S.I. ml/min.
Pressures: Inlet.....	Outlet.....
Hydrogen. 4.0 P.S.I. ml/min.	Air 1.0 P.S.I. ml/min.
DETECTOR E.C....T.C.....	F.I.D. 1.0 P.S.I.
Sens.....	Rate..... ml/min.
Scavenger.....	Rec. Range..... mv
SAMPLE. 1335.....	Size 2.0 mL
Solvent.....	Concn.....

RUN # 10 JUN/18/81 16:05:19

ESTD	RT	AREA	TYPE	CAL#	AMOUNT
	0.22	0	PP	1R	0.000
	0.46	6338	PB		0.000
	0.97	371	BB		0.000
	1.34	867	BB	4R	0.135

TOTAL AREA= 7576
MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location TOSCO, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 4-12-81, Time 10:47:27, Instrument ID VARIAN
3700

Recorder/Printout Reference No. 1, Recorder ID HP 3340A

Purpose of Run CALIBRATION

Sample Description SCOTT multi-component
MIXTURE UN-PARAFINS ± 10%

GC CONDITIONS

Amount Injected 2.0 ml, Inj. Port or Sample Loop Used 2.0 ml Loop

Detector Used: FID X, ECD , FPD , TCD (Current)

Detector Attenuation 4, Amplifier or Range 10⁻¹¹

Column: Liquid Phase , Solid Phase PORAPAK Q,

Length 6', O.D. 1/8", I.D. , Material S.S.

Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C

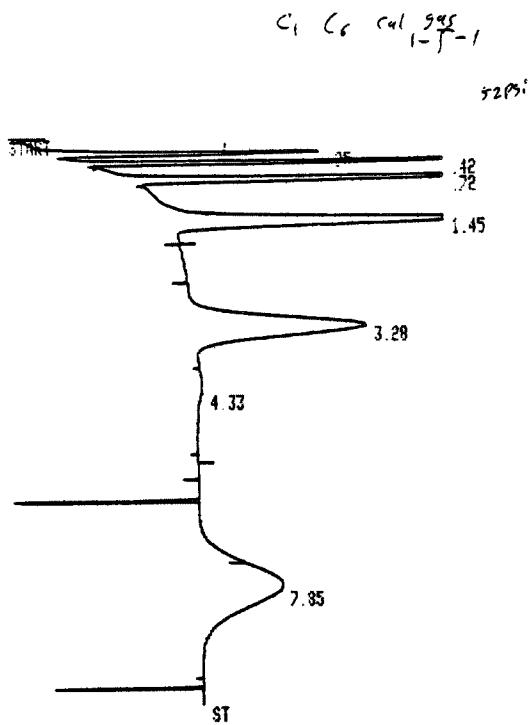
Temperature Program ISOTHERMAL

SAMPLE RUN

Sampling Method PURGE LOOP

RT	Area	Peak Height	Amount (ppm)	Component
1.2	6.3600	.	17.0	C ₁
1.4	2.0353	.	10.3	C ₂
1.5	4.6758	.	10.6	C ₃
1.7	1.0500	.	10.8	C ₄
3.23	1.2587	.	10.4	C ₅
7.7	2.5787	.	10.1	C ₆

Name of Operator M.D. CHIPIES, Date 9-1 1981



Operator...M.D.C.H.I.S.	Date...6-18-81
Column No.....	Length...6'
Coating.....	Dia...1/8"
Support...PORARAK Q	Concn.....
TEMP: Col: Init. 100 °C	Mesh. 60/80
Rate..... °C/min.	Final. 100 °C
CARRIER GAS N ₂	Inj. 120 °C
Pressures: Inlet.....	Rate 4.0 PSI
Hydrogen...N ₂ O ₂ /1.0 ml./min.	Outlet
DETECTOR E.C..... T.C.	Air 4.0 PSI
Scavenger.....	F.I.D. 2.0 ID
Sens.....	Rate .ml./min.
SAMPLE CAL. #11.....	Rec.Range .mv.
Solvent.....	Size 2.0 ml.
	Concn.....

RUN # 11 JUN/18/81 16:27:39

ESTD			
RT	AREA	TYPE	CAL#
0.25	29335	PY	1R
0.42	33384	VV	2R
0.72	63253	VV	3R
1.45	182110	VB	4R
3.28	82646	BV	5R
4.33	9846	VV	
7.85	47986	BP	6R

TOTAL AREA= 358760
MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location Tesco, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 6-18-81, Time 16:49:16, Instrument ID VARIAN 3700

Recorder/Printout Reference No. 12, Recorder ID HP3390A

Purpose of Run C₁-C₆ HYDROCARBON ANALYSIS

Sample Description BLANK

GC CONDITIONS

Amount Injected 2.0ML, Inj. Port or Sample Loop Used 2.0ML Loop

Detector Used: FID X, ECD , FPD , TCD (Current)

Detector Attenuation 4, Amplifier or Range 10⁻¹¹

Column: Liquid Phase , Solid Phase PORAPAK Q, Length 6', O.D. 1/8", I.D. , Material S.S.

Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C

Temperature Program ISOTHERMAL

SAMPLE RUN

Sampling Method N/A

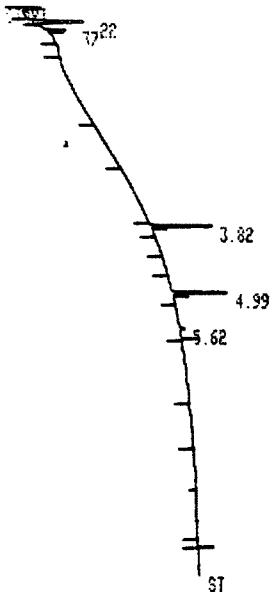
RT	Area	Peak Height	Amount (ppm)	Component
2.2	228		2.0	C ₁
3.7	971		0.3	C ₂
5.31	1231	\	-	"
7.77	1185	/	0.4	C ₃
5.62	274	/	-	"

Name of Operator M.D. CHIPS, Date 5-1 1981

Blank BulB B (236°F (63°C))

Zero Air pulse

150°C
 4×10^{-11}
 52psi



Operator. M. D. C. H. I. S.	Date. 6-18-81
Column No. 6'	Length. 6'
Coating. P.R.R.A.P.A.K. Q.	Dia. $\frac{1}{8}$ "
Support. P.R.R.A.P.A.K. Q.	Concn. Mesh 65/80
TEMP: Col: Init. 150 °C	Final 150 °C
Rate..... °C/min.	Det. 2.50 °C
CARRIER GAS. H ₂ S.	Inj. 120 °C
Pressures: Inlet.	Rate 60 psi. ml./min.
Hydrogen. 4.0. psi. ml./min.	Outlet. Air 60 psi. ml./min.
DETECTOR E.C. T.C.	F.I.D. X 10 ⁻¹¹
Scavenger.	Rate. ml./min.
Sens. Rec. Range mw.	
SAMPLE. BLANK. BULB-B	Size 2.0 mL
Solvent. TIME: 16:49	Concn.

RUN # 12

JUN/18/81 16:49:16

ESTD

RT	AREA	TYPE	CAL#	AMOUNT
0.22	2286	BP	1R	1.911
0.37	991	PB	2R	0.306
3.82	1231	0	PB	0.000
4.99	1188	D	BB	0.000
5.62	274	BB		0.000

TOTAL AREA= 5970
 MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location Tosco, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 6-18-81, Time 17:02:35, Instrument ID VARIAN 3700
 Recorder/Printout Reference No. 12, Recorder ID HP3390A
 Purpose of Run C₁-C₆ HYDROCARBON ANALYSIS

Sample Description RUBBER BULK-A

GC CONDITIONS

Amount Injected 2.0ML, Inj. Port or Sample Loop Used 2.0ML LOOP
 Detector Used: FID X, ECD , FPD , TCD (Current)
 Detector Attenuation 4, Amplifier or Range 10⁻¹¹
 Column: Liquid Phase , Solid Phase PORAPAK Q,
 Length 6', O.D. 1/8", I.D. , Material S.S.
 Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C
 Temperature Program ISOTHERMAL

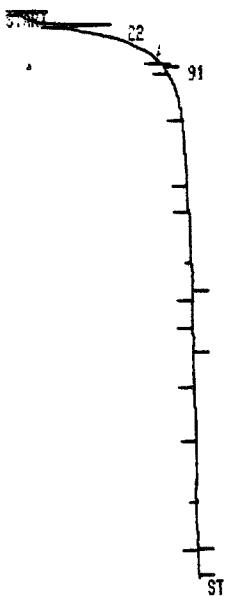
SAMPLE RUN

Sampling Method 1. Purge

RT	Area	Peak Height	Amount (ppm)	Component
.22	2366		2.1	C ₁
.71	57		.01	C ₄

Name of Operator M.D. CHIPS, Date 9-1 1981

BLANK BULB A (299°F)
 60PSI
 zero air
 150°C 4×10^{-4}



Operator.	M.D.Chips	Date.	JUN-18-81
Column No.	Length	6'	Dia. $\frac{1}{8}$ "
Coating	Concn.		
Support.	PtRAPAK Q	Mesh	60/80
TEMP. Col:	Inj. D. $^{\circ}\text{C}$	Final D. $^{\circ}\text{C}$	
Rate.....	$^{\circ}\text{C}/\text{min.}$	$^{\circ}\text{C}$	Det. 250 $^{\circ}\text{C}$ Inj. 120 $^{\circ}\text{C}$
CARRIER GAS.	H ₂	Rate	4.0 CPSI ml./min.
Pressures:	Inlet.	Outlet.	
Hydrogen	H_2 O.P.S. ml./min.	Air	O_2 O.P.S. ml./min.
DETECTOR E.C.	T.C.	F.I.D.	X 10 ⁴ mv.
Scavenger.....	Rate		ml./min.
Sens.	Rec. Range		
SAMPLE.	BLANK BULB-A	Size	2.0 ml.
Settent. Time:	1.702	Concn.	

RUN # 13 JUN/18/81 17:02:35

ESTD	RT	AREA	TYPE	CAL%	AMOUNT
	0.22	2366	PB	1R	1.979
	0.91	57	BB		0.000

TOTAL AREA= 2423
 MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location Tosco, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 6-18-81, Time 12:15 AM, Instrument ID VARIAN 3700

Recorder/Printout Reference No. 14, Recorder ID HP 3390A

Purpose of Run C₁-C₆ HYDROCARBON ANALYSIS

Sample Description 1740 BULB-R

GC CONDITIONS

Amount Injected 2.0 mL, Inj. Port or Sample Loop Used 2.0 mL Loop

Detector Used: FID X, ECD , FPD , TCD (Current)

Detector Attenuation 4, Amplifier or Range 10⁻¹¹

Column: Liquid Phase , Solid Phase PORAPAK Q,

Length 6', O.D. 1/8", I.D. , Material S.S.

Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C

Temperature Program ISOTHERMAL

SAMPLE RUN

Sampling Method GRAB .500 mL S.S. BULB

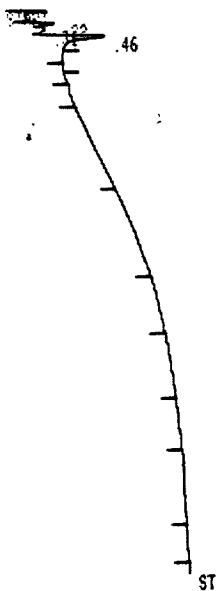
RT	Area	Peak Height	Amount (ppm)	Component
.22	1371		1.2	C ₁
.31	95.		0.3	C ₂
.46	7432		1.3	C ₃

Name of Operator M.D. CHIPS, Date 9-1 1981

1740 Sample B Taken 203°F
 Analyzed 250°F
 26 PSD

GC ATT = 4x6"

150°C
 1 cm/min



Operator..	M.D.C.H.P.S.	Date..	6-18-81
Column No.....	Length...6'	Dia.... $\frac{1}{8}$ "	
Coating.....	PORARAK Q	Mesh 60/80	Concn.
Support.....	Porarak Q	Init. I.D. °C	Final I.D. °C
TEMP: Col:	150°C	Rate..... °C/min.	Det. 250 °C Inj. 120 °C
CARRIER GAS...N ₂	Rate. 60 psi	ml./min.	
Pressures: Inlet.....	Outlet.....		
Hydrogen, N ₂ , Ar, etc. ml./min.	Air 60 psi	ml./min.	
DETECTOR E.C.....	T.C.....	F.I.D. 2 X 10 ⁻⁴	
Scavenger.....	Rate.....	ml./min.	
Sens.....	Rec.Range.....	mv.	
SAMPLE..1740.....	BULG: B	Size 2.0 ml	
Solvent.....	Concn.....		

RUN # 14

JUN/18/81 18:10:09

ESTD		AREA	TYPE	CAL#	AMOUNT
RT		1371	PY	1R	1.146
0.22		951	YP		0.000
0.31		7432	PB		0.000

TOTAL AREA= 9754
 MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location Tosco, Job No. 30 7602.91
BAKERSFIELD, CA

Injection Date 6-18-81, Time 11:21:54, Instrument ID VARIAN 3700

Recorder/Printout Reference No. 15, Recorder ID HP3390A

Purpose of Run C, -C₆ HYDROCARBON ANALYSIS

Sample Description 70 312-A

GC CONDITIONS

Amount Injected 2.0ml, Inj. Port or Sample Loop Used 2.0ml Loop

Detector Used: FID X, ECD , FPD , TCD (Current)

Detector Attenuation 4, Amplifier or Range 10⁻¹¹

Column: Liquid Phase , Solid Phase PORAPAK Q,
Length 6', O.D. 1/8", I.D. , Material S.S.

Temperature: Injector 120 °C, Oven 150 °C, Detector 350 °C

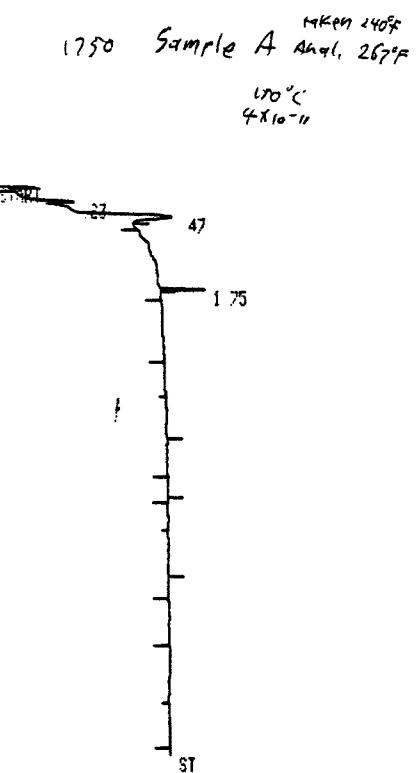
Temperature Program ISOTHERMAL

SAMPLE RUN

Sampling Method GRAB .500 ml S.S. BULB

RT	Area	Peak Height	Amount (ppm)	Component
1.22	6	—	—	C ₁
1.47	6825	1.4	2.0	C ₂
1.75	6702	0.9	2.5	C ₃

Name of Operator M.D. CHIPS, Date 9-1 1981



Operator...M.D.CHISS	Date...6-18-81
Column No.....	Length...6'..... Dia....1/8"
Coating.....	Concn.....
Support...FERRAPAK Q	Mesh. 60/80
TEMP: Col: Init. 1. D.	Final. 1. 2. °C
Rate..... °C/min.	Det. 250 °C Inj. 130 °C
CARRIER GAS...N ₂	Rate. 40 CPSI. ml./min.
Pressures: Inlet.....	Outlet.....
Hydrogen. H ₂ O. PSI. ml./min.	Air. N ₂ O. PSI. ml./min.
DETECTOR E.C. T.C.	FIDUX 1D-41
Scavenger.....	Rate..... ml./min.
Sens.....	Rec.Range..... mv.
SAMPLE 1750 Bulg-A	Size 2.0 MM.
Solvent.....	Concn.....

RUN # 15 JUN/18/81 18:21:54

ESTD	RT	AREA	TYPE	CAL#	AMOUNT
	0.23	0	PP	1R	0.000
	0.47	6885	PB		0.000
	1.75	6903	D BB		0.000

TOTAL AREA= 13788
MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location TOSCO, Job No. 307602-91
BAKERSFIELD, CA

Injection Date 6-18-81, Time 11:20:59, Instrument ID VARIAN 3700

Recorder/Printout Reference No. 1-, Recorder ID HP3390A

Purpose of Run CALIBRATION

Sample Description SCOTT multi-component
MIXTURE UN-PARAFINS ± 10%

GC CONDITIONS

Amount Injected 2.0 ml, Inj. Port or Sample Loop Used 2.0 ml loop

Detector Used: FID X, ECD , FPD , TCD (Current)

Detector Attenuation 4, Amplifier or Range 10⁻¹¹

Column: Liquid Phase , Solid Phase PORAPAK Q,
Length 6', O.D. 1/8", I.D. , Material S.S.

Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C

Temperature Program ISOTHERMAL

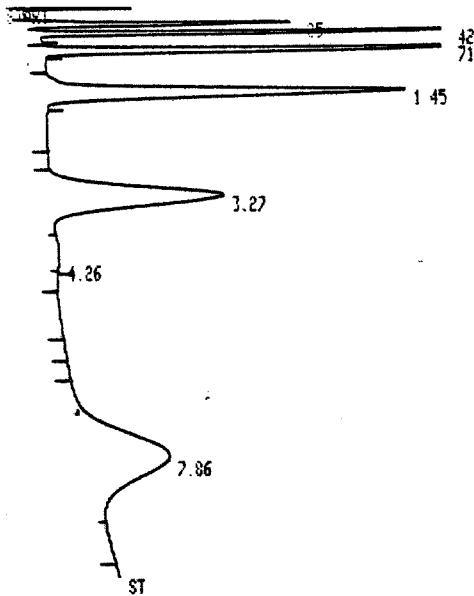
SAMPLE RUN

Sampling Method PURGE LOOP

RT	Area $\times 10^{-4}$	Peak Height	Amount ppm	Component
.25	7.2471	.	17.0	C ₁
.42	3.5743	.	10.3	C ₂
.71	2.3752	.	10.6	C ₃
1.45	1.5875	.	10.8	C ₄
2.27	1.1576	.	10.4	C ₅
7.86	12.40	.	10.1	C ₆

Name of Operator M. D. CHIPS, Date 9-1 1981

C₁-C₆ cul 995
1-T-1



Operator. M.D. CHIPS	Date. 6-18-81
Column No.	Length. 6'
Coating.	Dia. 1/8"
Support. PEGAPAK Q	Concen. Mesh 60/200
TEMP: Col: Init. 150 °C	Final. 120 °C
Rate. °C/min.	Det. 250 °C
CARRIER GAS. H ₂	Inj. 1.00 ml/min.
Pressures: Inlet.	Rate, ΔP_1 , ml/min.
Hydrogen. ΔP_2	Outlet, ΔP_2 , ml/min.
DETECTOR E.C.	T.C. F.I.D.
Scavenger.	Rate, ΔP_3 , ml/min.
Sens.	Rec.Range, mv.
SAMPLE, C ₁ -C ₆	Size, 2.0mL
Solvent.	Concn.

RUN # 16 JUN/18/81 19:28:39

ESTD		RT	AREA	TYPE	CAL#	AMOUNT
0.25	18389	BV	1R	15.373		
0.42	29030	V8	2R	8.957		
0.71	45742	BB	3R	7.666		
1.45	68033	PB	4R	7.196		
3.27	76947	BV	5R	9.683		
4.26	1719	VW		0.000		
7.86	82853	PB	6R	17.439		

TOTAL AREA= 322710
MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location Tesco, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 6-18-81, Time 20:01:10, Instrument ID VARIAN 3700

Recorder/Printout Reference No. 7, Recorder ID HP3390A

Purpose of Run C, -C₆ HYDROCARBON ANALYSIS

Sample Description 15:45 BULK-A

GC CONDITIONS

Amount Injected 2.0ML, Inj. Port or Sample Loop Used 2.0ML LOOP

Detector Used: FID X, ECD , FPD , TCD (Current)

Detector Attenuation 4, Amplifier or Range 10⁻¹¹

Column: Liquid Phase , Solid Phase PORAPAK Q,

Length 6', O.D. 1/8", I.D. , Material S.S.

Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C

Temperature Program ISOTHERMAL

SAMPLE RUN

Sampling Method GRAB .500 ML S.S. BULB

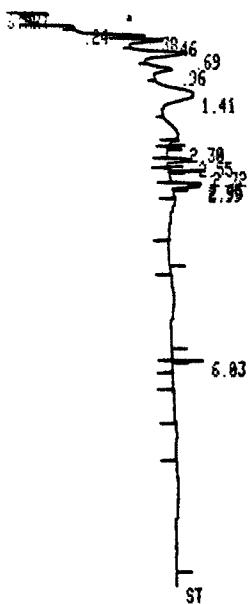
RT	My Area 10.1+	Peak Height	Amount (ppm)	Component
.24	2.748		3.5	C ₁
.33	9.701		7.2	C ₂
.42	1.2376		1.2	C ₃
.69	2.4031		5.6	C ₄
.76	1.7404		7.0	C ₅
1.41	2.6507		7.0	C ₆

→ continued on next page

Name of Operator M.D. CHIPS, Date 9-1 1981

RT	Area $\times 10^{-7}$	Peak Height	Amount (ppm)	Component
1.30	1196	1		
2.55	1479			
2.72	865		0.7	C ₂
2.75	1177	-		
2.77	1530			
6.53	572		0.07	C ₆

1945 Sample A 275°F
Anal 228°F



Operator, M. D. CHIPS	Date, 6-19-81
Column No.	Length, 6'
Coating,	Dia., $\frac{1}{8}$ "
Support, PORAPAK Q	Concn., Mesh 60/80
TEMP: Col: Init. 1.0 °C	Final, 1.0 °C
Rate, °C/min.	Det. RSD, °C
CARRIER GAS, H ₂	Inlet, 1.20 ml/min.
Pressures: Inlet,	Rate, L/D, psi
Hydrogen, H ₂ , psi	Outlet, ml/min.
DETECTOR E.C.	Air, L/D, psi
Sens.	ml/min.
Scavenger, Rate,	ml/min.
Solvent, Rec. Range, ml/min.	mv.
SAMPLE, 1945, BUL-A	Size, 2.0 ml
Solvent, Concn.	

RUN # 17 JUN/18/81 20:01:20

ESTD				
RT	AREA	TYPE	CAL#	AMOUNT
0.24	2748	BY	1R	2.540
0.38	9701	VV	2R	3.442
0.46	12396	VV		0.000
0.59	24834	VV	3R	5.570
0.96	17404	VV		0.000
1.41	26507	VB	4R	4.208
2.38	196	D BB		0.000
2.55	1499	BB		0.000
2.72	865	D PB		0.000
2.95	1194	BY		0.000
2.99	1530	D VB		0.207
6.83	592	D BB		0.000

TOTAL AREA= 98666
MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location Tosco, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 6-18-81, Time 20:12:58, Instrument ID VARIAN 3700
 Recorder/Printout Reference No. JK, Recorder ID HP 3390A
 Purpose of Run C, -C₆ HYDROCARBON ANALYSIS

Sample Description 1950 R12-B

GC CONDITIONS

Amount Injected 2.0mL, Inj. Port or Sample Loop Used 2.0mL Loop
 Detector Used: FID X, ECD , FPD , TCD (Current)
 Detector Attenuation 4, Amplifier or Range 10⁻¹¹
 Column: Liquid Phase , Solid Phase PORAPAK Q,
 Length 6', O.D. 1/8", I.D. , Material S.S.
 Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C
 Temperature Program ISOTHERMAL

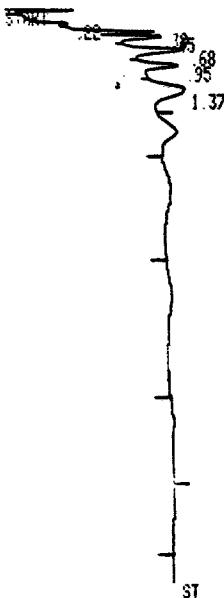
SAMPLE RUN

Sampling Method GRAB 500 mL S.S. BULB

RT	Area	Peak Height	Amount (ppm)	Component
.22	1050	•	0.9	-1
.30	7514	\	5.0	-2
.45	13103	/	5.6	-3
.68	24088	/	7.3	-4
.75	18656	\		
.87	27033	/		

Name of Operator M.D. CHIPS, Date 9-1 1981

1450 Sample B 295°F
Ave / 260°F



Operator...M.D.C.H.P.S.	Date...6-18-81
Column No.....	Length...6'
	Dia...1/8"
Coating.....	Concen.....
Support...P.D.R.A.P.A.K. Q	Mesh. 20/80
TEMP: Col: Init. 150 °C	Final. 170 °C
Rate..... °C/min.	Det. 25.0 °C
CARRIER GAS. H ₂ .	Inlet. Rate. 4.0 psi. ml/min.
Pressures: Inlet.	Outlet.
Hydrogen. 4.0 psi. ml/min.	Air. 4.0 psi. ml/min.
DETECTOR E.C. T.C.	F.I.D. 4.0 psi.
Sens.	Scavenger.
SAMPLE. 1450 BULB-B	Rec.Range.
Solvent.	Size. 2.0 ml.
	Concn.

RUN # 18 JUN/18/81 20:12:58

ESTD	RT	AREA	TYPE	CAL#	AMOUNT
	0.22	1000	PV	1R	0.925
	0.38	9514	VV		0.000
	0.45	13108	VV	2R	4.651
	0.68	24088	VV	3R	5.582
	0.95	18656	VV		0.000
	1.37	27033	VB	4R	4.291

TOTAL AREA= 93399
MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location TOSCO, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 6-18-81, Time 10:32:22, Instrument ID VARIAN 3700
 Recorder/Printout Reference No. 17, Recorder ID HP3390A
 Purpose of Run C₁-C₆ HYDROCARBON ANALYSIS

Sample Description 1945 Sample A

GC CONDITIONS

Amount Injected 2.0ML, Inj. Port or Sample Loop Used 2.0ML Loop
 Detector Used: FID X, ECD , FPD , TCD (Current)
 Detector Attenuation 4, Amplifier or Range 10⁻¹¹
 Column: Liquid Phase , Solid Phase PORAPAK Q,
 Length 6', O.D. 1/8", I.D. , Material S.S.
 Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C
 Temperature Program ISOTHERMAL

SAMPLE RUN

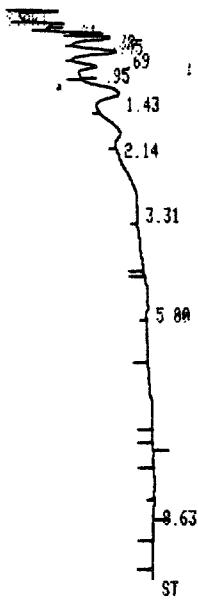
Sampling Method GRAB 500 ml S.S. BULB

RT	Area	Peak Height	Amount (ppm)	Component
.21	1312		1.2	C ₁
.36	3032		2.7	C ₂
.45	7373		2.7	C ₂
.67	2114		1.6	C ₃
.85	2721		2.2	C ₄
1.13	7483			

Name of Operator M.D. CHIPS, Date 9-1 1981

RT	Area	Peak Height	Amount (ppm)	Component
2.14	8550	1		
3.31	7370	.	5.	C ₅
5.05	17.7		.02	C ₆
8.63	748	-		> C ₆

1945 Sample A (repet)



Operator...M.D.C.H.P.S.	Date...6-18-81
Column No....	Length...6' Dia. $\frac{1}{8}$ "
Coating....	Concn.,
Support...PORARAK Q.	Mesh 60/62
TEMP: Col: Init. 1.0. °C Final. 1.0. °C	Rate. °C/min.
Rate.....	Det. RSD. °C Inj. 130 °C
CARRIER GAS H ₂	Rate. 6.0. PSI. ml/min.
Pressures: Inlet.....	Outlet.....
Hydrogen. 40. PSI. ml/min.	Air. 6.0. PSI. ml/min.
DETECTOR E.C. T.C. F.I.D. & I.O. ml/min.	
Scavenger.....	Rate.....
Sens.	Rec. Range.
SAMPLE...1945 BOLB-A	Size. 2.0. MM
Solvent.	Concn.

RUN # 19 JUN/18/81 20:32:22

ESTD

RT	AREA	TYPE	CAL#	AMOUNT
0.21	1312	PB	1R	1.213
0.39	3032	PV		0.000
0.45	7373	VV	2R	2.616
0.69	7114	VV	3R	1.649
0.95	2921	VB		0.000
1.43	9483	BV	4R	1.595
2.14	8550	VV		0.000
3.31	9376	VB	5R	1.267
5.88	1717	BB		0.000
8.63	748	PB		0.000

TOTAL AREA= 51626
MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location TOSCO, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 6-18-81, Time 21:09:12, Instrument ID VARIAN 3700

Recorder/Printout Reference No. 31, Recorder ID HP3390A

Purpose of Run C, -C₆ HYDROCARBON ANALYSIS

Sample Description 20-55 2122-A

GC CONDITIONS

Amount Injected 2.0ML, Inj. Port or Sample Loop Used 2.0ML Loop

Detector Used: FID X, ECD , FPD , TCD (Current)

Detector Attenuation 4, Amplifier or Range 10⁻¹¹

Column: Liquid Phase , Solid Phase PORAPAK Q,

Length 6', O.D. 1/8", I.D. , Material S.S.

Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C

Temperature Program ISOTHERMAL

SAMPLE RUN

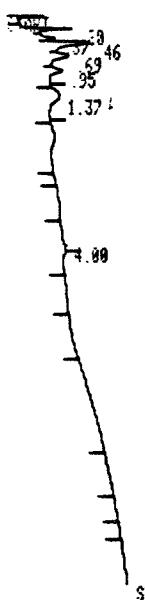
Sampling Method GRAB .500 ML S.S. BULB

RT	Area	Peak Height	Amount (ppm)	Component
.20	2526	✓	2.3	C ₁
.37	3346	✓	5.2	C ₂
.46	3645	✓		C ₂
.67	6058		1.4	C ₃
.75	2898	✓	5.4	C ₄
1.37	2557	✓	5.4	C ₇
4.02	0		0	C ₆

Name of Operator M.D. CHIPS, Date 9-1 1981

2055 Sample A (taken 280°F
Anal 270°F)
30 PSI

150°C
52 PSI
 4×10^{-11}



Operator...M.D.: Q.H.P.S.	Date 6-18-81
Column No.....	Length 6'
Coating.....	Dia. $\frac{1}{8}$ "
Support.....	Mesh 60/80
TEMP: Col:	Init. 1.0 °C Final 1.0 °C
Rate.....	°C/min. Det. 250 °C Inj. 120 °C
CARRIER GAS	He Rate. 60.0 PSI ml/min.
Pressures: Inlet	Outlet
Hydrogen...H ₂ PSI	ml/min. Air 60.0 PSI ml/min.
DETECTOR E.C.	T.C. F.I.D. 4×10^{-12}
Scavenger	Rate. Rec.Range. ml/min. mv.
Sens....	Size. 2.0 mV
SAMPLE...2055	Concn.
Solvent...	

RUN # 28 JUN/18/81 21:09:12

ESTD

RT	AREA	TYPE	CAL#	AMOUNT
0.20	2526	BV		0.000
0.37	3346	VV	2R	1.187
0.46	8645	VV		0.000
0.69	6058	VV	3R	1.404
0.95	2888	VB		0.000
1.37	2537	BB	4R	0.403
4.00	0	PB		0.000

TOTAL AREA= 26000
MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location Tosco, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 6-18-81, Time 21:55:42, Instrument ID VARIAN 3700
 Recorder/Printout Reference No. 34, Recorder ID HP3390A
 Purpose of Run C₁-C₆ HYDROCARBON ANALYSIS

Sample Description 2155 BULB-A

GC CONDITIONS

Amount Injected 2.0ML, Inj. Port or Sample Loop Used 2.0ML Loop
 Detector Used: FID X, ECD , FPD , TCD (Current)
 Detector Attenuation 4, Amplifier or Range 10⁻¹¹
 Column: Liquid Phase , Solid Phase PORAPAK Q,
 Length 6', O.D. 1/8", I.D. , Material S.S.
 Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C
 Temperature Program ISOTHERMAL

SAMPLE RUN

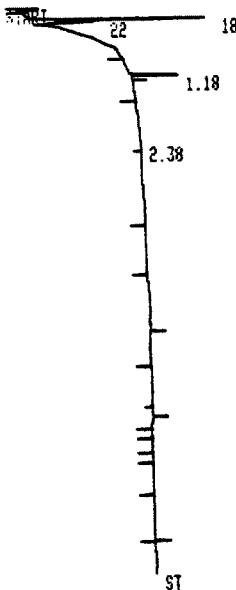
Sampling Method GRAB .500 ml S.S. BULB

RT	Area	Peak Height	Amount (ppm)	Component
1.22	3283	.	2.1	C ₁
1.18	1277	.	0.2	C ₄
2.38	6578	.	5.07	C ₅

Name of Operator M.D. CHIPS, Date 5-1 1981

Sample Pulp A 245°F
Zero Air Blank GOPSZ

LIST: ZERO = 0, 0.0



Operator: M.D.C. & P.S.	Date: 6-18-81	
Column No.: 1	Length: 6'	Dia. 1/8"
Coating: Porapak Q	Mesh: 60/80	Concn.:
Support: Porapak Q	Init. I.D. °C	Final I.D. °C
TEMP: Col:	Rate..... °C/min.	Inj. °C
CARRIER GAS: H ₂	Rate..... ml./min. ml./min.
Pressures: Inlet..... Outlet.....	
Hydrogen, %: Psi. ml./min.	Air, %: Psi. ml./min.	
DETECTOR E.C. T.C. F.I.D. N.I.C.	Rate..... ml./min.	
Sens. Rec.Range. mV.	
SAMPLE: 2.155 g. Bulb-A	Size: 2.0 mL	
Solvent.....	Concn.	

RUN #: 24

JUN/18/81 21:55:18

ESTD

RT	AREA	TYPE	CAL#	AMOUNT
0.18	3219	D PY		0.000
0.22	2283	YB	1R	2.111
1.18	1277	D BB		0.000
2.38	658	PB		0.000

TOTAL AREA= 7437
MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location TOSCO, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 6-18-81, Time 32:CE:57, Instrument ID VARIAN 3700

Recorder/Printout Reference No. 35, Recorder ID HP3390A

Purpose of Run C₁-C₆ HYDROCARBON ANALYSIS

Sample Description 2205 COB-B-13

GC CONDITIONS

Amount Injected 2.0ml, Inj. Port or Sample Loop Used 2.0ml Loop

Detector Used: FID X, ECD , FPD , TCD (Current)

Detector Attenuation 4, Amplifier or Range 10⁻¹¹

Column: Liquid Phase , Solid Phase PORAPAK Q,
Length 6', O.D. 1/8", I.D. , Material S.S.

Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C

Temperature Program ISOTHERMAL

SAMPLE RUN

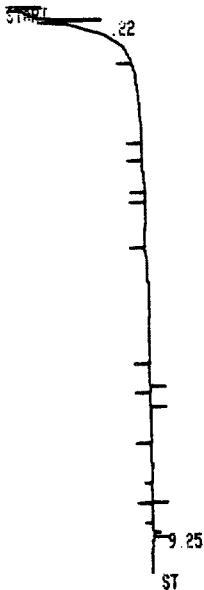
Sampling Method GRAB .500 ml S.S. BULB

RT	Area	Peak Height	Amount (ppm)	Component
.02	2.17	.	2.0	C ₁
7.25	174	.	—	>C ₆

Name of Operator M.D.Chips, Date 9-1 1981

Blank Sample BulB B 240°F
Zero Air 60 psi

LIST ZERO = 0.-0.1



Operator...M.D.SATIERS.....	Date...6-18-81.....
Column No.....	Length 6' Dia. 1/8"
Coating...PORARAK Q.....	Concn.....
Support...PORARAK Q.....	Mesh 10/60.....
TEMP: Col: Init. 120°C.....	Final. 120°C.....
Rate.....°C/min. Det. 250 °C.....	Inlet. 120 °C.....
CARRIER GAS...H2.....	Rate. 1.0 psi.....ml./min.
Pressures: Inlet.....	Outlet.....
Hydrogen. 1.0 psi...ml./min. Air & N2 1.0 psi...ml./min.	FID. Size 1/16".....
DETECTOR E.C.....T.C.....	Size 2.0 ml.....
Scavenger.....	Rate.....ml./min.
Sens.....	Rec.Range.....mv.
SAMPLE. 2.05 BulB-B.....	Size 2.0 ml.....
Solvent.....	Concn.....

RUN # 25 JUN/18/81 22:05:57

ESTD	RT	AREA	TYPE	CAL#	AMOUNT
	0.22	2189	PB	1R	2.024
	0.25	174	D PB		0.000

TOTAL AREA= 2363
MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location TOSCO, Job No. 307602-91
BAKERSFIELD, CA

Injection Date 6-17-81, Time 09:58:08, Instrument ID VARIAN 3700

Recorder/Printout Reference No. 27, Recorder ID HP3340A

Purpose of Run CALIBRATION

Sample Description SCOTT multi-component
MIXTURE C1-PARAFINS ± 10%

GC CONDITIONS

Amount Injected 2.0 mL, Inj. Port or Sample Loop Used 2.0 mL Loop

Detector Used: FID X, ECD , FPD , TCD (Current)

Detector Attenuation 4, Amplifier or Range 10⁻¹

Column: Liquid Phase , Solid Phase PORAPAK Q, Length 6', O.D. 1/8", I.D. , Material S.S.

Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C

Temperature Program ISOTHERMAL

SAMPLE RUN

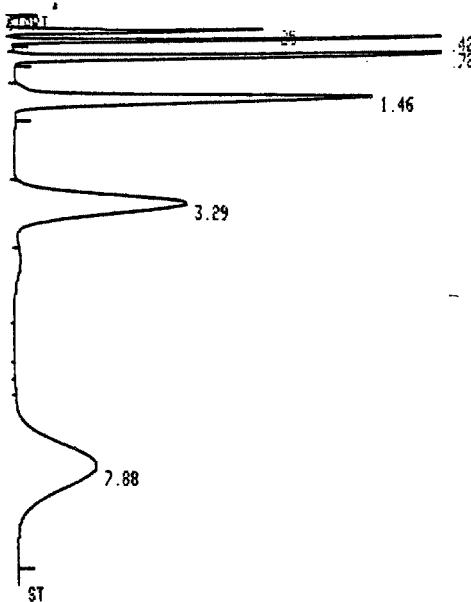
Sampling Method PURE LOOP

RT	MIN Area $\times 10^{-4}$	Peak Height	Amount ppm	Component
.25	7.6424	.	17.0	C ₁
.40	3.5352	.	10.3	C ₂
.69	2.33.7	.	10.6	C ₃
.73	1.5990	.	10.8	C ₄
1.36	1.3517	.	10.4	C ₅
1.82	1.1776	.	10.1	C ₆

Name of Operator M. D. CHIPS, Date 9-1 1981

Cal C₁-C₆ 1-T-1

LIST: ZERO = 0, 0.0



Operator. M.D. CHIPS	Date. 6-19-81
Column No.	Length. 6'
Coating... Support. PORARAK Q	Concn. Mesh.
TEMP: Col: Init. 100 °C	Final. 100 °C
Rate..... °C/min.	Det. 250 °C Int. 420 °C
CARRIER GAS..... H ₂	Rate. 1.0 psi. ml./min.
Pressures: Inlet. Hydrogen 40 psi. ml./min.	Outlet. Air 1.0 psi. ml./min.
DETECTOR E.C. T.C.	FID 4.0 x 10 ⁻⁴
Scavenger. Rate.	ml./min.
Sens. 0.758	Rec. Range. mv.
SAMPLE. CAL # 27	Size. 2.0
Solvent.	Concn.

RUN # 27 JUN/19/81 09:58:08

ESTD	RT	AREA	TYPE	CAL#	AMOUNT
	0.25	17870	BP	2R	0.000
	0.42	28720	PB	2R	10.190
	0.72	45643	BB	3R	10.577
	1.46	67818	BB	4R	10.766
	3.29	76621	BB	5R	10.356
	7.88	87186	BB	6R	10.628

TOTAL AREA= 323860
MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location TOSCO, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 4-19-81, Time 10:57:15, Instrument ID VARIAN 3700

Recorder/Printout Reference No. 2, Recorder ID HP 3390A

Purpose of Run CALIBRATION

Sample Description SCOTT multi-component
MIXTURE UN-PARAFINS ± 10%

GC CONDITIONS

Amount Injected 2.0 ml, Inj. Port or Sample Loop Used 2.0 ml loop

Detector Used: FID X, ECD , FPD , TCD (Current)

Detector Attenuation 4, Amplifier or Range 10⁻¹¹

Column: Liquid Phase , Solid Phase PORAPAK Q,

Length 6', O.D. 1/8", I.D. , Material S.S.

Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C

Temperature Program ISOTHERMAL

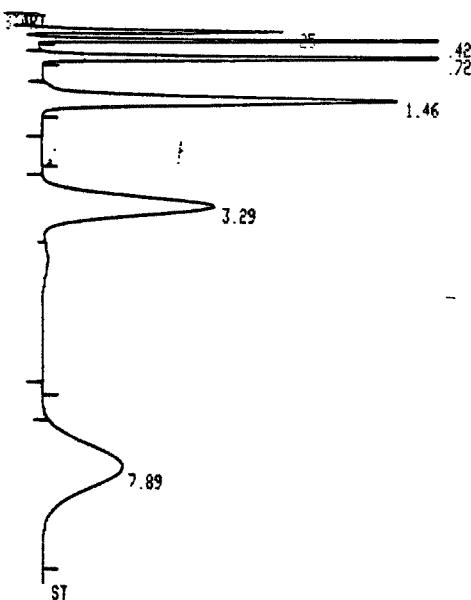
SAMPLE RUN

Sampling Method PURE LOOP

RT	Area % ⁺	Peak Height	Amount ppm	Component
2.25	2.2±27	.	17.0	C ₁
3.40	3.5±52	.	10.3	C ₂
4.67	2.2±17	.	10.6	C ₃
5.73	1.5±90	.	10.8	C ₄
7.26	1.3±17	.	10.4	C ₅
7.28	1.1±76	.	10.1	C ₆

Name of Operator M. D. CHIPS, Date 9-1 1981

c₁-c₆ cal



Operator.....	M.D. CHIPS.....	Date.....	6-19-81.....
Column No.....	6'	Length.....	6'
Coating.....	Concn.....	Mesh 60/40.....	Dia. 1/8"
Support.....	PORAPAK Q.....	Concn.....	Mesh 60/40.....
TEMP.....	Col: Init. 100°.....	Det. 100°.....	Final 100°.....
Rate.....	°C/min.	°C	°C
CARRIER GAS.....	N ₂	Rate: 60 psi.....	Inj. 120°.....
Pressures: Inlet.....	Outlet.....		
Hydrogen, H ₂ , P ₂	Air, O ₂ , P ₃	ml/min.	ml/min.
DETECTOR E.C.....	T.C.....	FID or KID.....	
Scavenger.....	Rate.....	ml/min.	
Sens.....	Rec. Range.....	mv.	
SAMPLE, C.A.L. #.....	10.5.....	Size 2.0 ml.....	
Solvent.....	Concn.....		

RUN # 28

JUN/19/81 18:15:15

ESTD

RT	AREA	TYPE	CAL#	AMOUNT
0.25	17459	P4	1	16.609
0.42	28710	V8	2R	18.296
0.72	45423	BB	3R	19.549
1.46	67497	BB	4R	19.749
3.29	76372	BB	5R	19.366
7.89	84110	BB	6R	9.744

TOTAL AREA= 319570
MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location TOSCO, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 6-7-81, Time 12:21:17, Instrument ID VARIAN 3700

Recorder/Printout Reference No. 29, Recorder ID HP 3390A

Purpose of Run CALIBRATION

Sample Description SCOTT MULTI-COMPONENT
MIXTURE UN-PARAFINS ± 10%

GC CONDITIONS

Amount Injected 2.0 ml, Inj. Port or Sample Loop Used 2.0 ml loop

Detector Used: FID X, ECD , FPD , TCD (Current)

Detector Attenuation 4, Amplifier or Range 10⁻¹¹

Column: Liquid Phase , Solid Phase PORAPAK Q,

Length 6', O.D. 1/8", I.D. , Material S.S.

Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C

Temperature Program ISOTHERMAL

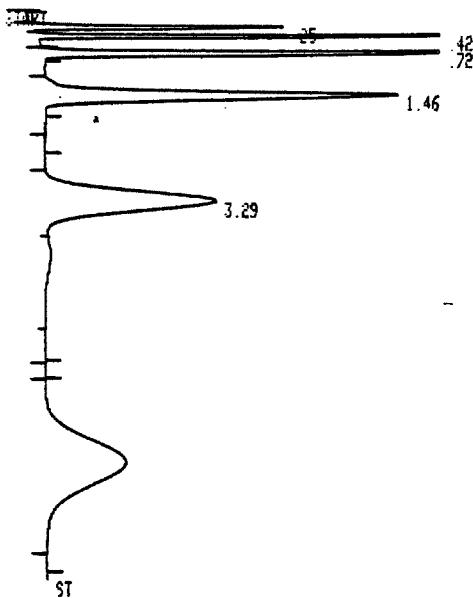
SAMPLE RUN

Sampling Method PURE LOOP

RT	Area	Peak Height	Amount (ppm)	Component
1.25	15.56	.	17.0	C ₁
1.42	28.757	.	10.3	C ₂
1.72	15.204	.	10.6	C ₃
1.75	67.3.2	.	10.8	C ₄
2.24	77.871	.	10.4	C ₅
—	—	.	10.1	C ₆

Name of Operator M.D. CHIPE, Date 9-1 1981

C₁ - C₆ C₇/



Operator...D. Q.M.R.S.	Date...6-19-81
Column No.....	Length...6'
Coating.....	Dia...1/8"
Support...PORAPAK Q	Concn.....
TEMP: Col: Init. 1.0 °C	Final. 1.0 °C
Rate..... °C/min.	Det. 250 °C
CARRIER GAS...N ₂	Inlet. Rate (O ₂) ml./min.
Pressures: Inlet.....	Outlet.....
Hydrogen... ml./min.	Air... ml./min.
DETECTOR E.C.T.C.....	F.I.D.
Scavenger.....	Rate..... ml./min.
Sens.....	Rec.Range..... mV.
SAMPLE Q.A. #. 29	Size 2.0 mL
Solvent.....	Concn.....

RUN # 29

JUN/19/81 10:27:17

ESTD

RT	AREA	TYPE	CAL#	AMOUNT
0.25	17567	PY	1	16.908
0.42	28757	V8	2R	10.315
0.72	45304	BB	3R	10.547
1.46	67312	BB	4R	10.745
3.29	77845	V9	5R	10.583

TOTAL AREA= 236790
MUL. FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location Tesco, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 6-11-81, Time 11:1400, Instrument ID VARIAN 3700
 Recorder/Printout Reference No. 30, Recorder ID HP 3390A
 Purpose of Run C₁-C₆ HYDROCARBON ANALYSIS

Sample Description L.D. 232-A

GC CONDITIONS

Amount Injected 2.0 mL, Inj. Port or Sample Loop Used 2.0 mL Loop
 Detector Used: FID X, ECD , FPD , TCD (Current)
 Detector Attenuation 4, Amplifier or Range 10⁻¹¹
 Column: Liquid Phase , Solid Phase PORAPAK Q,
 Length 6', O.D. 1/8", I.D. , Material S.S.
 Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C
 Temperature Program ISOTHERMAL

SAMPLE RUN

Sampling Method GRAB .500 ML S.S. BULB

RT	Area	Peak Height	Amount (ppm)	Component
.22	174		1.1	C ₁
.38	238		1.1	C ₂
.42	257			
.70	479		0.1	C ₃
.76	596		5.09	C ₄

Name of Operator M.D. CHIPS, Date 9-1 1981

1110 Sample A
 GC 20 150 250 74KPa 265°F
 470° 285°F



Operator... M.D. GRIER	Date... 6-19-81	
Column No.....	Length... 6'	Dia. 1/8"
Coating.....	Concn.....	
Support... P.A.R.P.A.K. Q	Mesh. 60/80	
TEMP: Col: Init. 1. 0 °C	Final. 1. 0 °C	
Rate..... °C/min.	Det. 250 °C	
CARRIER GAS... N ₂	Inlet. Rate. /cm. ³ /min.	
Pressures: Inlet.	Outlet	
Hydrogen. H ₂ P.i. ml./min.	Air. R. P.i. ml./min.	
DETECTOR E.C. T.C.	F.I.D. 4x 10 ⁻¹¹ mv	
Scavenger..... Rate..... ml./min.	Sens. Rec. Range. mv	
SAMPLE I.D. RUL-R-A	Size 2.0 m.s.	
Solvent.....	Concn.....	

RUN # 30 JUN/19/81 11:24:02

ESTD	RT	AREA	TYPE	CAL#	AMOUNT
	0.22	1194	PY	1	1.151
	0.38	738	WV	2R	0.265
	0.46	2459	VB		0.000
	0.78	479	BB	3R	0.112
	0.96	596	BB		0.000

TOTAL AREA= 5466
 MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location Tosco, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 6-1-81, Time 11:34:51, Instrument ID VARIAN 3700
 Recorder/Printout Reference No. 31, Recorder ID HP3390A
 Purpose of Run C₁-C₆ HYDROCARBON ANALYSIS

Sample Description 115 BULB

GC CONDITIONS

Amount Injected 2.0 mL, Inj. Port or Sample Loop Used 2.0 mL Loop
 Detector Used: FID X, ECD , FPD , TCD (Current)
 Detector Attenuation 4, Amplifier or Range 10⁻¹¹
 Column: Liquid Phase , Solid Phase PORAPAK Q,
 Length 6', O.D. 1/8", I.D. , Material S.S.
 Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C
 Temperature Program ISOTHERMAL

SAMPLE RUN

Sampling Method GRAB. 500 mL S.S. BULB

RT	Area	Peak Height	Amount (ppm)	Component
.32	1220		1.1	C ₁
.51	646			
.77	7480		1.18	C ₂

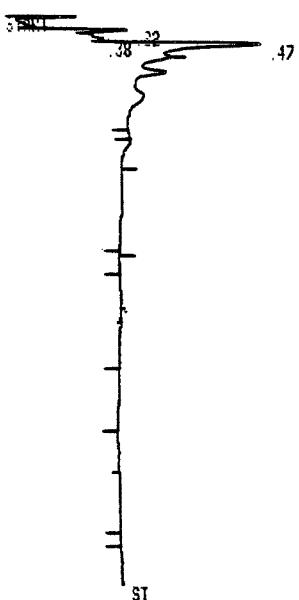
Name of Operator M.D. CHIPS, Date 9-1 1981

LIST: ATT 2† = 0

ATT 2† - 2 @
LIST: ATT 2† = -2

1115 Sample B
taken 30°C
Anal 287°F

Sample code
B-17



Operator.....	M. D. S. Hips.....	Date.....	6-19-81.....
Column No.....	6'	Length.....	6'
Coating.....	/g.	Dia.....	1/8"
Support.....	DORPAK Q.....	Concn.....	Mesh 20/60.....
TEMP: Col:	Init..... 1 °D..... °C	Final..... 2 °C..... °C	
Rate..... °C/min.	Det. RSP..... °C	Inj. 23.0..... °C	
CARRIER GAS.....	Rate (G.D. Psi)..... ml./min.		
Pressures: Inlet..... ml./min.	Outlet..... ml./min.		
Hydrogen..... ml./min.	Air..... G.C. Psi..... ml./min.		
DETECTOR E.G..... T.C.....	F.I.D. 4.2..... ml/min.		
Scavenger.....	Rate..... ml./min.		
Sens.....	Rec. Range..... MV.		
SAMPLE..... 1115.....	Size 3.0 mm.....		
Solvent.....	Concn.....		

RUN # 31 JUN/19/81 11:36:01

ESTD

RT	AREA	TYPE	CAL#	AMOUNT
0.22	1220	PY	1	1.176
0.38	666	VY	2R	0.239
0.47	4480	VB		0.000

TOTAL AREA= 6366
MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location Tosco, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 6-11-81, Time 12:17:55, Instrument ID VARIAN 3700

Recorder/Printout Reference No. 32, Recorder ID HP3390A

Purpose of Run C, - C₆ HYDROCARBON ANALYSIS

Sample Description 115 P.M.-C

GC CONDITIONS

Amount Injected 2.0 mL, Inj. Port or Sample Loop Used 2.0 mL Loop

Detector Used: FID X, ECD , FPD , TCD (Current)

Detector Attenuation 4, Amplifier or Range 10⁻¹¹

Column: Liquid Phase , Solid Phase PORAPAK Q,

Length 6', O.D. 1/8", I.D. , Material S.S.

Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C

Temperature Program ISOTHERMAL

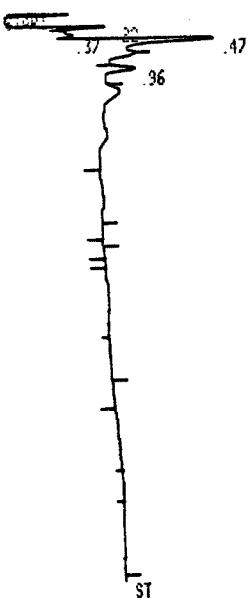
SAMPLE RUN

Sampling Method GRAB .500 mL S.S. BULB

RT	Area	Peak Height	Amount (ppm)	Component
.32	1220		1.1	C
.37	630		1.6	C ₂
.47	3754		1.6	C ₃
.49	665		0.1	C ₄

Name of Operator M.D. CHIPS, Date 9-1 1981

115 Sample B Repeat
and 257°F



Operator...M.D.C.HIPS	Date...6-19-81
Column No....	Length...6' Dia...1/4"
Coating...	Concn.
Support...PORARAK Q	Mesh 60/80
TEMP: Col: Init. 1 P. °C Final 10 °C	Inj. 130 °C
Rate.....°C/min.	Det. 3.30 °C
CARRIER GAS.....	Rate (L.D.P.S.I.) ml./min.
Pressures: Inlet.....	Outlet.....
Hydrogen. 4% Ar, ml./min.	Air 6.0 P.S.I. ml./min.
DETECTOR E.C..... T.C..... F.I.D. 420°C	Rate..... ml./min.
Scavenger.....	Rec. Range..... mv.
Sens.....	
SAMPLE...115 BULB-B	Size 2.0 ml.
Solvent.....	Concn.....

RUN # 32 JUN/19/81 12:17:55

ESTD RT	AREA	TYPE	CAL%	AMOUNT
0.22	1220	PY	1	1.176
0.37	680	VP	2R	0.244
0.47	3754	PB		0.000
0.96	665	BB		0.000

TOTAL AREA= 6319
MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location Tosco, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 6-81, Time 1:26:12, Instrument ID VARIAN 3700
 Recorder/Printout Reference No. 33, Recorder ID HP3390A
 Purpose of Run C₁-C₆ HYDROCARBON ANALYSIS

Sample Description 1-02 BULK-A

GC CONDITIONS

Amount Injected 2.0ML, Inj. Port or Sample Loop Used 2.0ML Loop
 Detector Used: FID X, ECD , FPD , TCD (Current)
 Detector Attenuation 4, Amplifier or Range 10⁻¹¹
 Column: Liquid Phase , Solid Phase PORAPAK Q,
 Length 6', O.D. 1/8", I.D. , Material S.S.
 Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C
 Temperature Program ISOTHERMAL

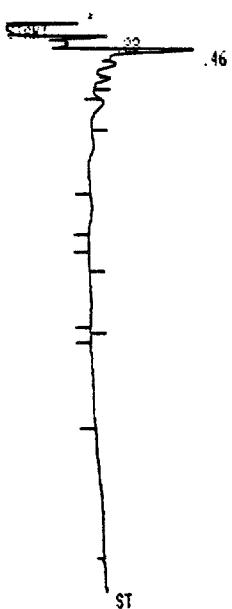
SAMPLE RUN

Sampling Method GRAB .500 ml S.S. BULB

RT	Area	Peak Height	Amount (ppm)	Component
<u>1.22</u>	<u>1409</u>		<u>1.3</u>	<u>C₁</u>
<u>1.46</u>	<u>5070</u>		<u>1.8</u>	<u>C₂</u>

Name of Operator M.D. CHIPS, Date 9-1 1981

1400 sample taken 290°F
at 210°F



Operator...	M. S. Hines	Date...	6-19-81
Column No...	6'	Length...	6"
Coating...	Q	Dia...	1/8"
Support...	P.D.R. ADAMS	Concn...	
TEMP:	Col: Init. 120 °C	Mesh 60/80	Final. 120 °C
Rate.....	°C/min.	Det. 250 °C	Inj. 120 °C
CARRIER GAS...	H ₂	Rate: 60 ml./min.	ml./min.
Pressures:	Inlet	Outlet	
Hydrogen: 2.0	ml./min.	Air: 2.0	ml./min.
DETECTOR E.C.	T.C.	F.I.D. X 1.0	
Scavenger...	Rate	ml./min.	
Sens.	Rec. Range	mV	
SAMPLE...	Buyl-A	Size 2.0 mm	
Solvent...	Concn		

RUN # 33 JUN/19/81 14:26:42
NO CALIB PEAKS FOUND

AREA%	RT	AREA TYPE	AR/HT	AREA%
	0.22	1409 PY	0.054	21.668
	0.46	5096 WV	0.112	78.340

TOTAL AREA= 6505
MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location Tesco, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 6-1-81, Time 14:39:09, Instrument ID VARIAN 3100
 Recorder/Printout Reference No. 37, Recorder ID HP 3310A
 Purpose of Run C₁-C₆ HYDROCARBON ANALYSIS

Sample Description HGC RUB-R

GC CONDITIONS

Amount Injected 2.0ml, Inj. Port or Sample Loop Used 2.0ml Loop
 Detector Used: FID X, ECD , FPD , TCD (Current)
 Detector Attenuation 4, Amplifier or Range 10⁻¹¹
 Column: Liquid Phase , Solid Phase PORAPAK Q,
 Length 6', O.D. 1/8", I.D. , Material S.S.
 Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C
 Temperature Program INTERNAL

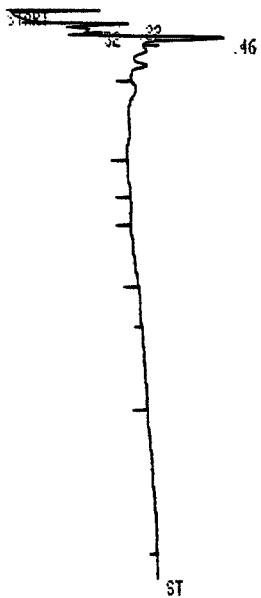
SAMPLE RUN

Sampling Method GRAB 500 ml SS. 25LBS

RT	Area	Peak Height	Amount (ppm)	Component
.22	1101		1.0	1
.32	331	✓		1
.46	2729	✓	1.1	2

Name of Operator M.D. CHIPS, Date 9-1 1981

14-05 sample B taken 25°F
Area 28°F



Operator...	M.D. CHIPS	Date...	6-19-81
Column No.....	L.	Length.....	6'
Coating.....	1/8"	Dia.....	1/8"
Support.....	PORAPAK Q	Concn.....	
TEMP: Col:	Init. 142 °C	Final. 7 °C	Mesh 60/80
Rate.....	°C/min.	Det. 250 °C	Inj. 420 °C
CARRIER GAS	H ₂	Rate 60 ml/min.	
Pressures:	Inlet	Outlet	
Hydrogen. 40 psi. ml/min.	Air 40 psi. ml/min.		
DETECTOR E.C.	T.C.	F.I.D. X 10 ⁻¹¹	
Scavenger	Rate	ml/min.	
Sens.	Rec. Range	mV	
SAMPLE... 14-05	BOLB-B	Size 2.0 mm	
Solvent.....	Concn.....		

RUN # 34 JUN/19/81 14:39:09
NO CALIB PEAKS FOUND

AREA%		RT	AREA	TYPE	AR/HT	AREAZ
0.22	1101	0.22	4161	PY	0.044	26.468
0.32	331	0.32		VB	0.066	7.955
0.46	2729	0.46		PB	0.078	65.585

TOTAL AREA= 4161
MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location TOSCO, Job No. 307602-91
BAKERSFIELD, CA

Injection Date 6-15-81, Time 10:21:11, Instrument ID VARIAN 3700

Recorder/Printout Reference No. 2, Recorder ID HP 3390A

Purpose of Run CALIBRATION

Sample Description SCOTT multi-component
MIXTURE UN-PARAFINS ± 10%

GC CONDITIONS

Amount Injected 2.0 ml, Inj. Port or Sample Loop Used 2.0 ml Loop

Detector Used: FID X, ECD , FPD , TCD (Current)

Detector Attenuation 4, Amplifier or Range 10⁻¹

Column: Liquid Phase , Solid Phase PORAPAK Q,
Length 6', O.D. 1/8", I.D. , Material S.S.

Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C

Temperature Program ISOTHERMAL

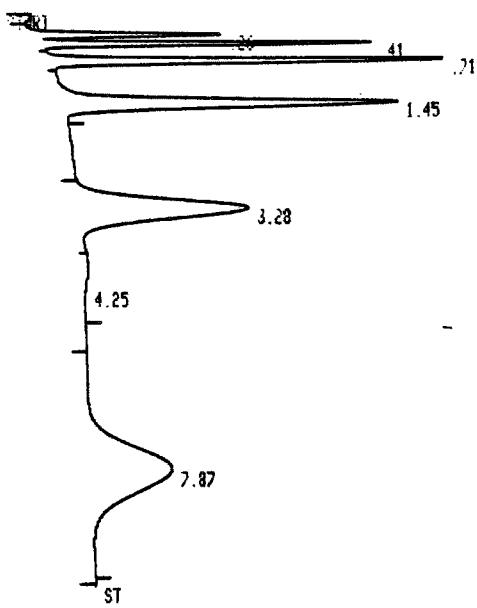
SAMPLE RUN

Sampling Method PULSE LOOP

RT	Area	Peak Height	Amount (ppm)	Component
1.16	37310	.	17.0	C ₁
1.71	37372		10.3	C ₂
1.74	21431		10.6	C ₃
1.75	14770		10.8	C ₄
1.86	12380		10.4	C ₅
2.87	11353		10.1	C ₆

Name of Operator M.D. CHIPS, Date 9-1 1981

C₁ - C₆ ref



Operator: M.J.C.H.P.S.	Date: 6-19-81
Column No.: 6'	Length: 6'
Coating: Porapak Q	Dia.: 1/8"
Support: Porapak Q	Concn.: %/g
TEMP: Col: Int. 100 °C	Final. 170 °C
Rate: 10 °C/min.	Det. & SD: 10 °C
CARRIER GAS: He	Inj. 120 °C
Pressures: Inlet	Rate 10 ml/min.
Hydrogen: 10 ml/min.	Outlet
DETECTOR E.C.: T.C.	Air fact. 1.0 ml/min.
Scavenger:	T.C. Rate 10 ml/min.
Sens.	Rec. Range: mv
SAMPLE: C ₆ Alk. #2	Size: 3.0 ml
Solvent:	Concn.: %/g

RUN #: 2 JUN/19/81 16:42:11

AREA%				
RT	AREA	TYPE	AR/HT	AREA%
0.26	19034	D P4	0.885	5.488
0.41	29949	D VV	0.875	8.635
0.71	49004	D VV	0.896	14.130
1.45	73116	V8	0.184	21.882
3.28	80746	V8	0.400	23.282
4.25	6241	V8	0.722	1.890
7.87	88727	V8	0.930	25.583

TOTAL AREA= 346820
MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location TOSCO, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 6-5-81, Time 7:45A, Instrument ID VARIAN 3700
 Recorder/Printout Reference No. 2, Recorder ID HP3390
 Purpose of Run C₁-C₆ HYDROCARBON ANALYSIS

Sample Description 1-72 RULB-A

GC CONDITIONS

Amount Injected 2.0 ml, Inj. Port or Sample Loop Used 2.0 ml Loop
 Detector Used: FID X, ECD , FPD , TCD (Current)
 Detector Attenuation 4, Amplifier or Range 10⁻¹¹
 Column: Liquid Phase , Solid Phase PORAPAK Q,
 Length 6', O.D. 1/8", I.D. , Material S.S.
 Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C
 Temperature Program ISOTHERMAL

SAMPLE RUN

Sampling Method GRAB .500 ml S.S. BULB

RT	Area	Peak Height	Amount (ppm)	Component
.23	10-7		1.5	C
.47	30-50		2.0	C ₂
.72	10-5		0.2	C ₃

Name of Operator M.D. CHIPS, Date 9-1 1981

1645 SAMPLE A SAMPLE
240°F
30 PSI ANALYSIS
303 OF



Operator...M.D. CHIASS.	Date...6-19-81
Column No.....	Length, 6'
Coating.....	Dia. $\frac{1}{8}$ "
Support...P.R.R.A.K. Q.	Concn.....
TEMP: Col: Init. 100 °C	Mesh, 60/80
	Final 100 °C
Rate..... °C/min.	Int. 2.50 °C
CARRIER GAS...H ₂	Inlet 1.00 BAR
Pressures: Inlet	Rate, 60 ml/min.
Hydrogen, H ₂ , ml/min.	Outlet
DETECTOR E.C.T.C.F.I.D. X,N,H ₂	ml/min.
Scavenger.....	Rate..... ml/min.
Sens.....	Rec.Range..... mv.
SAMPLE 1643	Size 2.0 ml.
Solvent.....	Concn.....

RUN # 3 JUN/19/81 17:14:08
NO CALIB PEAKS FOUND

AREA%			
RT	AREA	TYPE	AR/HT
0.23	1649	PV	0.081
0.47	8656	VV	0.169
0.96	1618	VB	0.184
			13.840
			72.648
			13.512

TOTAL AREA= 11915
MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location TOSCO, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 6-17-81, Time 2:26:02, Instrument ID VARIAN 3700
 Recorder/Printout Reference No. 4, Recorder ID HP339D
 Purpose of Run C₁-C₆ HYDROCARBON ANALYSIS

Sample Description TEST BULB-B

GC CONDITIONS

Amount Injected 2.0 ml, Inj. Port or Sample Loop Used 2.0 ml Loop
 Detector Used: FID X, ECD , FPD , TCD (Current)
 Detector Attenuation 4, Amplifier or Range 10⁻¹¹
 Column: Liquid Phase , Solid Phase PORAPAK Q,
 Length 6', O.D. 1/8", I.D. , Material S.S.
 Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C
 Temperature Program ISOTHERMAL

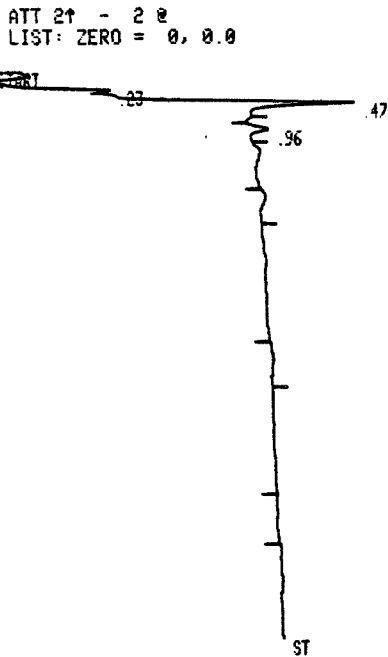
SAMPLE RUN

Sampling Method GRAB 50 ml S.S. BULB

RT	Area	Peak Height	Amount (ppm)	Component
.43	70+	.	0.1	C
.47	15+	.	3.0	C ₂
.50	513	.	0.07	C ₃

Name of Operator M.D. CHIPS, Date 9-1 1981

1650 Sample B Sample 249°F
Anal 229°F



Operator. M.D. CHIPS	Date 6-19-81
Column No.	Length 6'
Coating	Dia. 1/8"
Support. P.T.R.A.PAK Q	Concen.
TEMP: Col: Init. 150 °C	Final. 100 °C
Rate..... °C/min.	Det. 350 °C
CARRIER GAS H ₂	Inj. 120 °C
Pressures: Inlet	Rate (psi) ml/min.
Hydrogen, N ₂ , Ar	Outlet
DETECTOR E.C. T.C.	Air (psi) ml/min.
Sens. 1.0	F.I.D. 4.5 x 10 ⁻¹¹ mv.
Scavenger	Rate .ml./min.
SAMPLE. 1650 BULG-B	Rec. Range .mv.
Solvent.	Concn.

RUN # 4 JUN/19/81 17:26:12
NO CALIB PEAKS FOUND

AREA%	RT	AREA	TYPE	AR/HT	AREA%
0.23		764	PV	0.050	7.698
0.47		8548	VB	0.144	86.126
0.96		613	PB	0.106	6.176

TOTAL AREA= 9925
MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CHEM, Location TORONTO, Job No. 2070-2-71
BAKERSFIELD, CA

Injection Date 4-5-71, Time 14:30:18, Instrument ID VARIAN
 Recorder/Printout Reference No. 1, Recorder ID HP3392A
 Purpose of Run CALIBRATION

Sample Description TEST MIXTURE
100% PROPANE IN AIR

GC CONDITIONS

Amount Injected 2.0 μl, Inj. Port or Sample Loop Used 2.0 μl
 Detector Used: FID x, ECD , FPD , TCD (Current)
 Detector Attenuation 1, Amplifier or Range 10-1
 Column: Liquid Phase , Solid Phase POLYAPAT,
 Length 6', O.D. 1/8", I.D. , Material S.S.
 Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C
 Temperature Program TEST PROGRAM

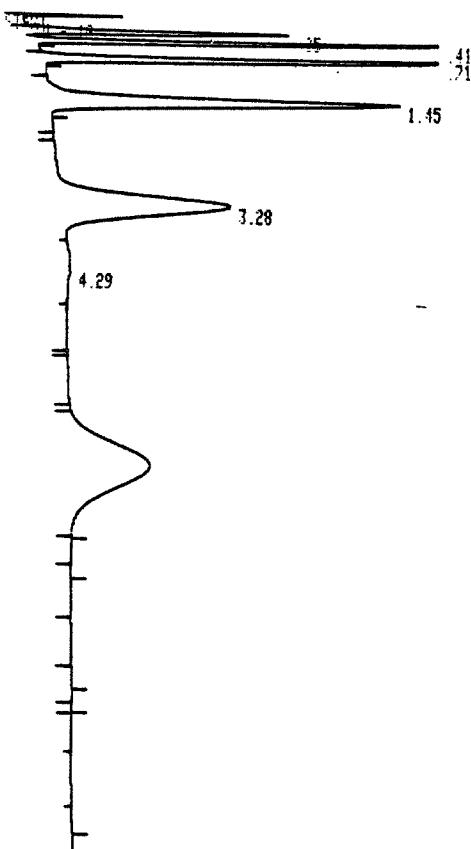
SAMPLE RUN

Sampling Method PURGE LOOP

RT	Area % RT	Peak Height	Amount (ppm)	Component
1.36	7.1123		17.0	C ₁
1.42	3.1823		10.3	C ₂
1.72	2.2127		10.6	C ₃
1.80	1.6047		10.8	C ₄
3.20	1.3572		10.4	C ₅
7.66	1.1422		10.1	C ₆

Name of Operator M.A. CHIESI, Date 9-1 1971

LIST: ZERO = 0, 0.0



STOP

Operator: M.D.CHAPES	Date: 6-19-81
Column No.: 1	Length: 6'
Coating: PORAPAK Q	Concn.: 1/4"
Support: PORAPAK Q	Mesh: 60/80
TEMP: Col: Init. 1.0 °C	Final 1.0 °C
Rate: 1.0 °C/min.	Det. 250 °C
Rate: Inj. 130 °C	
CARRIER GAS: H ₂	Rate: 140 ml/min.
Pressures: Inlet:	Outlet:
Hydrogen, N ₂ , Ar, He, ml/min.	Air, N ₂ , He, ml/min.
DETECTOR E.C.: T.C.: FID: X (L)	
Scavenger:	Rate: ml/min.
Sens.:	Rec. Range: mv.
SAMPLE C.A.L. #: 1	Size: 2.0 M.R.
Solvent:	Concn.:

RUN #: 1 JUN/19/81 16:21:18

AREA%	RT	AREA	TYPE	AR/HT	AREA%
0.25	18155	BY	0.059	7.462	
0.41	28629	YB	0.049	11.768	
0.71	44871	BB	0.071	18.444	
1.45	66644	PB	0.162	27.393	
3.28	79336	BY	0.394	32.619	
4.29	5653	YB	0.725	2.324	

TOTAL AREA= 243290
MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location TOSCO, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 6-7-81, Time 14:19:22, Instrument ID VARIAN 3700
 Recorder/Printout Reference No. 2, Recorder ID HP339D
 Purpose of Run C₁-C₆ HYDROCARBON ANALYSIS

Sample Description BLANK BULB-R

GC CONDITIONS

Amount Injected 2.0 ml, Inj. Port or Sample Loop Used 2.0 ml Loop
 Detector Used: FID X, ECD , FPD , TCD (Current)
 Detector Attenuation 4, Amplifier or Range 10⁻¹¹
 Column: Liquid Phase , Solid Phase PORAPAK Q,
 Length 6', O.D. 1/8", I.D. , Material S.S.
 Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C
 Temperature Program ISOTHERMAL

SAMPLE RUN

Sampling Method PURGE N2

RT	Area	Peak Height	Amount (ppm)	Component
.22	.728	.	1.6	C ₁
.75	6070	.	1.4	C ₃

Name of Operator M.D. CHIPS, Date 9-1 1981

Blank BULB 60PSI
20°C
216°F



Operator. M. D. CHIRES	Date. 6-19-81
Column No.....	Length... 6'
Coating.....	Dia.... 1/8"
Support. P.T.R.A.P.A.K. Q	Mesh. 100/120
TEMP: Col: Init. 1. 2. °C	Final. 1. 6. °C
Rate..... °C/min.	Detect. 50. °C Inj. 120. °C
CARRIER GAS. He.....	Rate (psi) 1.0 ml/min.
Pressures: Inlet.....	Outlet.....
Hydrogen (psi) 2.4 ml/min.	Air (psi) 2.4 ml/min.
DETECTOR E.C. T.C. F.I.D. He X 10 ⁻⁴	
Scavenger..... Rate.....	ml/min.
Sens..... Rec. Range.....	mv.
SAMPLE. BLANK. BULB-B	Size 2.0 ml.
Solvent. Time: 18.9	Concn.....

RUN # 2 JUN/19/81 18:19:32

ESTD	RT	AREA	TYPE	CAL#	AMOUNT
	0.22	1728	PY		0.000
	0.75	6076	VB	3R	1.387

TOTAL AREA= 7884
MUL FACTOR= 1.0000E+00

GAS CHROMATOGRAPH OPERATING CONDITIONS AND FIELD LOG

Client CMEA, Location TOSCO, Job No. 307602.91
BAKERSFIELD, CA

Injection Date 6-19-81, Time 18:21:47, Instrument ID VARIAN 3700
 Recorder/Printout Reference No. ?, Recorder ID HP 3590
 Purpose of Run C₁-C₆ HYDROCARBON ANALYSIS

Sample Description R. TV RULB-A

GC CONDITIONS

Amount Injected 2.0 ml, Inj. Port or Sample Loop Used 2.0 ml Loop
 Detector Used: FID X, ECD , FPD , TCD (Current)
 Detector Attenuation 4, Amplifier or Range 10⁻¹¹
 Column: Liquid Phase , Solid Phase PORAPAK Q,
 Length 6', O.D. 1/8", I.D. , Material S.S.
 Temperature: Injector 120 °C, Oven 150 °C, Detector 250 °C
 Temperature Program ISOTHERMAL

SAMPLE RUN

Sampling Method Purge N₂

RT	Area	Peak Height	Amount (ppm)	Component
•22	1-+1		1.3	C ₁

Name of Operator M.D. CHIPS, Date 9-1 1981

841B A 81mK (refect)

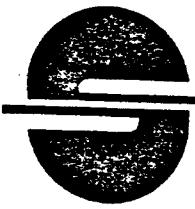
Operator...M.D.CARIPS..... Date...6-19-81
Column No....Length...6'..... Dia...1/8".....
Coating.....
Support...PORARAK Q.....Concn..... Mesh 60/80
TEMP: Col: Init...1.0 °C Final...1.0 °C
Rate..... °C/min. Det.R. 50 °C Inj. 120 °C
CARRIER GAS...H₂..... Rate 60 ft.⁻¹ ml/min.
Pressures: Inlet..... Outlet.....
Hydrogen H₂ F.S.I. ml/min. Air F.S.I. ml/min.
DETECTOR E.C. T.C. FID. X 10⁻¹
Scavenger..... Rate..... ml/min.
Sens..... Rec.Range..... mv.
SAMPLE BULGAR..... Size R.P.....
Solvent. TOME: 1.234 Concn.....

RUN # 3 JUN/19/81 18:31:47
NO CALIB PEAKS FOUND

AREA%	RT	AREA	TYPE	AR/HT	AREA%
	0.22	1441	PB	0.052	100.000

TOTAL AREA= 1441
MUL FACTOR= 1.0000E+00

5.9 RADIOMETRIC ANALYSIS RESULTS

**SAFETY SPECIALISTS, Inc.**

3194 De La Cruz Blvd., Suite 15 • Santa Clara, California 95050 • Telephone (408) 988-1111

ASSAY REPORT

Acurex Corporation
485 Clyde Avenue
Attn: Ms. Linda Bohannon, M/S 0-1212
Mountain View, California 94042

Date: December 28, 1981

Date Samples Received: 11/11/81

Customer Order No. RB59185A, Rel. 20

Activity*

<u>SSI No.</u>	<u>Client Description</u>	<u>Gross Beta pCi/Filter</u>	<u>Gross Gamma pCi/Filter</u>
81388A	#1, A81-07-011-038, TOSCO 2 Filter	60.6 ± 7.8	≤ 143
B	#2, A81-07-011-545, TOSCO 1 Filter	75.0 ± 11.3	≤ 143
C	#3, A81-07-011-571, TOSCO Filter Blank	99.6 ± 13.6	≤ 142

Gross Alpha
pCi/liter

<u>SSI No.</u>	<u>Client Description</u>	<u>Gross Alpha pCi/liter</u>
81388A	No. 1, A81-07-011-38, TOSCO 2 Filter	15 ± 12
B	No. 2, A81-07-011-545, TOSCO 1 Filter	≤ 18
C	No. 3, A81-07-011-571, TOSCO Filter Blank	≤ 19

Fariba Danesh
Analyst: Fariba DaneshApproved: P. C. Noble, Director
Safety & Health Services Division

*The ± values are the two sigma Poisson standard deviation of the counting error.

The ≤ values are equal to or less than three sigma of the counting error.

5.10 BIOASSAY REPORTS

GENETICS ASSAY NOS. 6153-6156
LBI SAFETY NOS. 7535-7538

MUTAGENICITY EVALUATION OF
XAD RESIN EXTRACTS
IN THE
EPA LEVEL 1
AMES SALMONELLA/MICROSOME
PLATE TEST

FINAL REPORT

SUBMITTED TO:

ACUREX CORPORATION
485 CLYDE AVENUE
MOUNTAIN VIEW, CALIFORNIA 94042

SUBMITTED BY:

LITTON BIONETICS, INC.
5516 NICHOLSON LANE
KENSINGTON, MARYLAND 20895

LBI PROJECT NO. 22064

REPORT DATE: MARCH 1982



BIONETICS

5-132

PREFACE

This assay conforms to the standard EPA Level 1 procedure for the Ames Salmonella/microsome mutagenesis assay as described in "IERL-RTP Procedures Manual: Level 1 Environmental Assessment Biological Tests"¹. The data were evaluated and formatted as recommended in "Level 1 Biological Testing Assessment and Data Formatting"².

The Ames Salmonella/microsome mutagenesis assay has been shown to be a sensitive method for detecting mutagenic activity for a variety of chemicals representing various chemical classes³. This assay is one of several recommended by EPA to identify, categorize and rank the pollutant potential of influent and effluent streams from industrial and energy-producing processes. This assay has been well validated with a wide range of positive and negative control chemicals and complex environmental samples.

All procedures and documents pertaining to the receipt, storage, preparation, testing and evaluation of the test material shall conform to Litton Bionetics, Inc. standard operating procedures, the U.S. Food and Drug Administration's Good Laboratory Practices Regulations of 1979⁴ and the proposed U.S. Environmental Protection Agency's Good Laboratory Practice Guidelines^{5,6}. Deviations from standard procedure shall be fully documented and noted in the report.

All test and control results in this report are supported by fully documented raw data which are permanently maintained in the files of the Department of Molecular Toxicology or in the archives of Litton Bionetics, Inc., 5516 Nicholson Lane, Kensington, Maryland 20895. Copies of raw data will be supplied to the sponsor upon request.



BIONETICS

TABLE OF CONTENTS

	<u>Page No.</u>
PREFACE	i
I. SUMMARY	1
II. OBJECTIVE	2
III. TEST MATERIAL	3
A. Description	3
B. Handling and Preparation	3
IV. MATERIALS	5
A. Indicator Microorganisms	5
B. Media	6
C. Activation System	6
1. S9 Homogenate	6
2. S9 Mix	6
V. EXPERIMENTAL DESIGN	7
A. Dose Selection	7
B. Mutagenicity Test	7
1. Nonactivation Assay	7
2. Activation Assay	8
C. Control Compounds	8
D. Recording and Presenting Data	10
VI. RESULTS	11
A. Interpretations	11
1. TOSCO 2 XAD + OMC	11
2. TOSCO 1 XAD	12
3. MOHAWK 1 XAD	12
4. MOHAWK 2 XAD	13
B. Tables	14
VII. ASSAY ACCEPTANCE AND EVALUATION CRITERIA	27
A. Surviving Populations	27
B. Dose-Response Phenomena	28
C. Control Tests	28
D. Evaluation Criteria for Ames Assay	28
1. Strains TA-1535 and TA-1537	29
2. Strains TA-98 and TA-100	29
3. Pattern	29
4. Reproducibility	29
E. Relation Between Mutagenicity and Carcinogenicity	30
F. Criteria for Ranking Samples in the Ames Assay	30
VIII. REFERENCES	32



BIONETICS

I. SUMMARY

Four resin extract samples supplied by Acurex Corporation were tested and evaluated for their mutagenic activities in the EPA Level 1 Ames Salmonella/microsome mutagenesis assay. The samples, supplied in methylene chloride, were solvent exchanged to 2.0 ml of dimethylsulfoxide (DMSO) prior to testing. The samples were identified as TOSCO 2 XAD + OMC (A81-07-011-562,559), TOSCO 1 XAD (A81-07-011-569), MOHAWK 1 XAD (A81-09-007-488) and MOHAWK 2 XAD (A81-09-007-495).

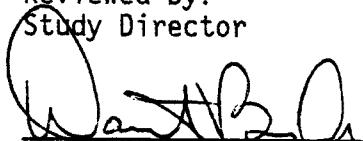
In order to preserve the maximum quantity of test material, it was decided to test each sample in only two strains, TA-98 and TA-100 using a single plate per dose. Tests were conducted in duplicate both with and without S9 metabolic activation mix. Even with these modifications, it was impossible to reach the Maximum Applicable Dose (MAD) for the Ames assay which is 5000 µg/plate. In the Mohawk 2 XAD sample no activity was observed in the highest dose, but based on the limited organic content, a Non Detectable classification could not be assigned. The sample was ranked as having undetermined mutagenicity, but moderate or less. The TOSCO 2 XAD sample was the most mutagenic with a Minimum Effective Concentration (MEC) of 30 µg/plate, placing the sample in the high mutagenicity category. Mohawk 1 XAD and TOSCO 1 XAD, were the next most active samples with MEC values of 75 µg/plate (moderate mutagenicity) and 85 µg/plate (moderate mutagenicity), respectively.

Insufficient quantities of test material prevented standard Level 1 Ames testing of the four samples. While three of the samples were found to be mutagenic and were ranked by EPA Level 1 evaluation criteria the MEC of those samples should be viewed with some caution. Adequate retesting was not possible to confirm the accuracy of the reported MEC values.

Submitted by:

Cynthia Rabenold 4/7/82
Cynthia Rabenold, B.S.
Submammalian Genetics,
Department of Molecular
Toxicology

Reviewed by:
Study Director

David J. Brusick, Ph.D. 4/7/82
Director,
Department of Molecular
Toxicology



BIONETICS

II. OBJECTIVE

The objective of this study was to determine and rank the genetic activity of four XAD resin extract samples in the EPA Level 1 Ames Salmonella mutagenesis assay with and without the addition of mammalian metabolic activation preparations. The samples were solvent exchanged into dimethylsulfoxide (DMSO) before Ames testing was initiated. The genetic activity of each sample was measured in these assays by its ability to revert the Salmonella indicator strains from histidine dependence to histidine independence. The degree of genetic activity of a sample was reflected in the number of revertants that were observed on the histidine-free medium. Standard EPA Level 1 mutagenicity evaluation criteria for the Ames mutagenesis assay were used to rank the mutagenic potential of each test material.



BIONETICS

III. TEST MATERIAL

A. Description

Four samples were supplied by Acurex Corporation, Mountain View, California. The samples were assigned LBI safety numbers and LBI assay numbers upon receipt. The Acurex code numbers, sample identification, LBI safety numbers and LBI assay numbers are identified below. All laboratory documentation used the LBI assay number to identify samples.

The four test materials were received as clear, yellow solutions of organic material in methylene chloride except for the TOSCO 1 XAD sample. That sample was an amber-colored, clear solution with a few suspended particles. The quantity of organic material in each sample, as determined by Acurex Corporation, is identified below. No information on sampling parameters (such as the equivalent volume of stack gas represented by the sample) was provided.

Acurex Corp. Code	Sample Identification	Quantity (mg organic)	LBI Safety No.	LBI Assay No.
A81-07-011-562, 559	TOSCO 2 XAD + OMC	12	7537	6153
A81-07-011-569	TOSCO 1 XAD	17	7538	6154
A81-09-007-488	MOHAWK 1 XAD	6	7535	6155
A81-09-007-495	MOHAWK 2 XAD	4	7536	6156

B. Handling and Preparation

The test materials were received at LBI on February 8, 1982. The samples were shipped in small, clear-glass vials sealed with crimp-top aluminum caps with rubber liners. The samples were received intact and were stored at +4°C in the dark until processed.



BIONETICS

Pretest sample preparation consisted of solvent exchanging the samples into dimethylsulfoxide (DMSO). The samples were transferred with methylene chloride rinses into graduated conical tubes. The methyl chloride was gradually evaporated (50°C under a stream of nitrogen) and DMSO was sequentially added. The samples were brought to volume in 2.0 ml of DMSO. The samples were transferred to glass vials and sealed with teflon-coated rubber rounds. The solvent exchanged samples were stored at +4°C in the dark.



BIONETICS

5-138

IV. MATERIALS

A. Indicator Microorganisms

The Salmonella typhimurium strains used in this assay were obtained from Dr. Bruce Ames, University of California at Berkeley⁷⁻¹². The following four strains were used.

Strain Designation	Gene Affected	Additional Mutations			Mutation Type Detected
		Repair	LPS	R Factor	
TA-1535	<u>his</u> G	Δ <u>uvr</u> B	<u>rfa</u>	-	Base-pair substitution
TA-1537	<u>his</u> C	Δ <u>uvr</u> B	<u>rfa</u>	-	Frameshift
TA-98	<u>his</u> D	Δ <u>uvr</u> B	<u>rfa</u>	pKM101	Frameshift
TA-100	<u>his</u> G	Δ <u>uvr</u> B	<u>rfa</u>	pKM101	Base-pair substitution

All the above strains have, in addition to the mutation in the histidine operon, mutation (rfa-) that leads to defective lipopolysaccharide coat, a deletion that covers genes involved in the synthesis of vitamin biotin (bio-) and in the repair of ultraviolet (uv) - induced DNA damage (uvrB-). The rfa- mutation makes the strains more permeable to many large molecules. The uvrB- mutation decreases repair of some types of chemically or physically damaged DNA and thereby enhances the strain's sensitivity to some mutagenic agents. The resistant transfer factor plasmid (R factor) pKM101 in TA-98 and TA-100 is believed to cause an increase in error-prone DNA repair that leads to many more mutations for a given dose of most mutagens¹¹. In addition, plasmid pKM101 confers resistance to the antibiotic ampicillin, which is a convenient marker to detect the presence of plasmid in the cells.

All indicator strains were kept at 4°C on minimal medium plates supplemented with a trace of biotin and an excess of histidine. In addition, the plates with plasmid-carrying strains contained ampicillin (25 µg/ml) to



BIONETICS

ensure stable maintenance of plasmid pKM101. New stock culture plates were made as often as necessary from the frozen master cultures or from single colony reisolates that were checked for their genotypic characteristics (his, rfa, uvrB, and bio) and for the presence of plasmid. For each experiment, an inoculum from the stock culture plates was grown overnight at 37°C in nutrient broth (Oxoid CM67) and used.

B. Media

The bacterial strains were cultured in Oxoid Media #2 (Nutrient Broth). The selective medium was Vogen Bonner Medium E with 2 percent glucose¹³. The overlay agar consisted of 0.6 percent purified agar with 0.05 mM histidine, 0.05 mM biotin and 0.1M NaCl according to the methods of Ames et al¹².

C. Activation System

1. S9 Homogenate

A 9,000 x g supernatant prepared from Sprague-Dawley adult male rat liver induced by Aroclor 1254 (Ames et al.¹²) was purchased commercially and used in these assays.

2. S9 Mix

S9 mix used in these assays consisted of the following components:

Components	Concentration per Milliliter S9 Mix
NADP (sodium salt)	4 μmoles
D-glucose-6-phosphate	5 μmoles
MgCl ₂	8 μmoles
KC1	33 μmoles
Sodium phosphate buffer pH 7.4	100 μmoles
Organ homogenate from rat liver (S9 fraction)	100 μliters



V. EXPERIMENTAL DESIGN

A. Dosage Selection

Test strategy and dose selection depend upon sample type and sample availability. The Level 1 manual¹ recommends solids to be initially tested at the maximum applicable dose (MAD) of 5 mg per plate and at lower concentrations of 2.5, 1, 0.5, 0.1 and 0.05 mg per plate. Liquids are tested initially at the MAD of 200 µl per plate, and at lower concentrations of 100, 50 and 10 µl per plate. Samples are retested over a narrower range of concentrations with strains showing positive results initially. Alternate doses are employed if sample size is limiting or at the direction of the sponsor.

Because sample size was limiting, a preliminary test was performed with strain TA-98, both with and without S9 metabolic activation, at 10 µl per plate for each sample. Doses selected for the second trial with each sample were either 1, 5, 10 and 50 µl per plate for TOSCO 2 XAD + OMC or 1, 10, 25 and 50 µl per plate for the other three samples. The third trial used 50 µl per plate and, in some cases, also included 25 and 75 µl per plate.

B. Mutagenicity Testing

The procedure used was based on the paper published by Ames *et. al.*¹² and was performed as follows:

1. Nonactivation Assay

To a sterile 13 x 100 mm test tube placed in a 43°C water bath the following was added in order:

- 2.00 ml of 0.6 percent agar containing 0.05 mM histidine and 0.05 mM biotin.
- Aliquot of test sample to give the appropriate dose.



BIONETICS

- 0.1 ml to 0.2 ml of indicator organism(s).
- 0.50 ml of 0.2M phosphate buffer, pH 7.4.

This mixture was swirled gently and then poured onto minimal agar plates (see IV B, Media). After the top agar had set, the plates were incubated at 37°C for approximately 2 days. The number of his⁺ revertant colonies growing on the plates were counted with an automatic colony counter and recorded.

2. Activation Assay

The activation assay was run concurrently with the nonactivation assay. The only difference was the addition of 0.5 ml of S9 mix (see IV C, Activation System) to the tubes in place of 0.5 ml of phosphate buffer which was added in nonactivation assays. All other details were similar to the procedure for nonactivation assays.

A detailed flow diagram for the plate incorporation assay is provided in Figure 1.

C. Control Compounds

A negative control consisting of the solvent used for the test material was also assayed concurrently with the test material. For negative controls, step 'b' of Nonactivation Assays was replaced by 0.05 ml of the solvent. The negative controls were employed for each indicator strain and were performed in the absence and presence of S9 mix. The solvent used to prepare the stock solution of the test material is given in the Results section of this report. All dilutions of the test material were made using this solvent. The amount of solvent used was equal to the maximum volume used to give the appropriate test dose.

AMES ASSAY [PLATE INCORPORATION METHOD]

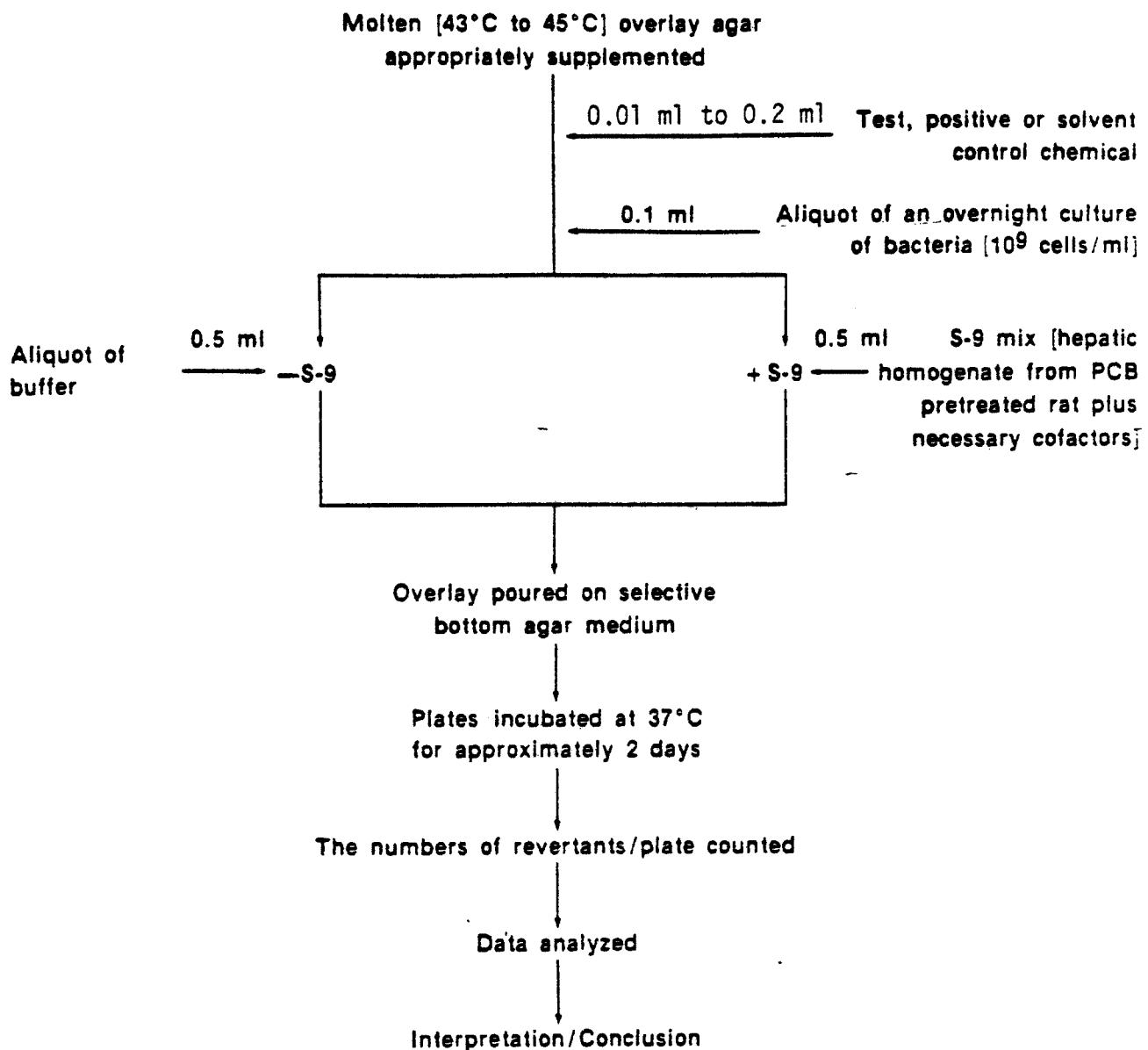


Figure 1 AMES SALMONELLA/MICROSOME MUTAGENESIS ASSAY

Specific positive control compounds known to revert each strain were also used and assayed concurrently with the test material. The concentrations and specificities of these compounds to specific strains are given in the following table:

Assay	Chemical	Solvent	Concentration per plate (μg)	Salmonella Strains
Nonactivation	Sodium azide	Water	10.0	TA-1535, TA-100
	2-Nitrofluorene (NF)	Dimethyl-sulfoxide	10.0	TA-98
	9-aminoacridine - (9AA)	Ethanol	50.0	TA-1537
Activation	2-anthramine (ANTH)	Dimethyl-sulfoxide	2.5	For all strains

D. Recording and Presenting Data

The number of colonies on each plate were counted and recorded on printed forms. These raw data were analyzed in a computer program and reported on a printout. The results are presented as revertants per plate for each indicator strain employed in the assay. The positive and solvent controls are provided as reference points.



BIONETICS

VI. RESULTS

A. Interpretation

With the limited amount of each sample received, the size of each Ames test was reduced to two strains, TA-98 and TA-100. A preliminary range-finding trial was performed with each sample (Trial 1 for each sample) using the single dose of 10 μ l/plate in strain TA-98. The results from this single data point were used to select an appropriate dose range for testing. This modified test strategy helped somewhat to conserve limited test material. Even with this step, the amount of sample was not sufficient to produce reproducible MEC value. The MEC values calculated for this study used the best data available from each series of tests.

The results of the assays showed that three of the four samples were mutagenic in either one strain or both. The most mutagenic sample on a per-weight basis was TOSCO 2 XAD (MEC of 30 μ g/plate) followed by Mohawk 1 XAD (MEC of 75 μ g/plate) and TOSCO 1 XAD (MEC of 85 μ g/plate) Mohawk 2 XAD was not mutagenic under the test conditions employed up to a level of 100 μ g organics per plate.

1. TOSCO 2 XAD + OMC

The primary stock of the sample was prepared at 6 μ g organics per μ l DMSO. The test material was evaluated in Salmonella strains TA-98 and TA-100 both with and without S9 metabolic activation mix. The dose range used in Trial 2 was 1, 5, 10 and 50 μ l per plate. The test sample was positive in TA-98 and marginally positive in TA-100 and appeared to be slightly more active without S9 mix. The sample was retested (Trial 3) at 50 μ l and 75 μ l (TA-100 only) per plate to verify the positive effect and because the TA-98 spontaneous background in Trial 2 was higher than assay acceptance criteria permit (see Section VII.B.).



BIONETICS

The minimum effective concentration (MEC) based on this data was 5 μ l (30 μ g organics) per plate with strain TA-98 without S9 metabolic activation. The sample showed a dose response over the range of concentrations tested in Trial 2. Based upon the EPA Level 1 evaluation criteria, the sample was ranked as having high (H) mutagenic activity.

2. TOSCO 1 XAD

The primary stock of the sample was prepared at 8.5 μ g organics per ml of DMSO. The initial trial at 10 μ l per plate with TA-98 was positive, both with and without S9 metabolic activation. The test material was then evaluated in Salmonella strains TA-98 and TA-100 both with and without S9 metabolic activation mix. The dose range used in Trial 2 was 5, 10, 25 and 50 μ l per plate. The test sample was not mutagenic for TA-98 in Trial 2, but showed a positive effect in Trial 3 at 50 μ l per plate. The reason for this was attributed to the high spontaneous mutant background with TA-98 in Trial 2.

The test sample was weakly mutagenic for TA-100 in Trial 2 showing a dose-related increase in the number of revertants, but never quite reaching the threshold for a positive response. The Trial 3 TA-100 results were positive with activation 50 and 75 μ l per plate. The minimum effective concentration was selected from Trial 1 for TA-98 (MEC of 10 μ l per plate; 85 μ g organics) TOSCO 1 XAD was found to have moderate mutagenic activity, but at a level closely approaching the high mutagenicity category. The sample was ranked as having moderate (M) mutagenicity based upon the EPA Level 1 evaluation criteria. Had sufficient sample been available for adequate testing, a clearer picture of the potency could have been obtained.

3. MOHAWK 1 XAD

The test material was prepared at a stock concentration of 3 μ g organics per ml DMSO. The test material was evaluated in Salmonella strains TA-98 and TA-100 both with and without S9 mix. The initial range-



finding trial with TA-98 at 10 μ l per plate was negative. The dose range used in Trial 2 was 5, 10, 25 and 50 μ l per plate. The material was positive for both TA-98 and TA-100 although the solvent control values for TA-98 were higher than normal. Activity in the presence of S9 mix was greater than without S9 mix. This suggests the presence of promutagens in the mixture. Strain TA-98 exhibited a dose-related increase in the number of revertants both with and without metabolic activation. Trial 3 was conducted to confirm Trial 2 and because the TA-98 spontaneous values from Trial 2 were higher than assay acceptance criteria permit.

The minimum effective concentration (MEC) based on this data was 25 μ l (75 μ g organics) per plate with both TA-98 and TA-100 in the presence of S9 mix (Trial 2 and Trial 3). Therefore, the Mohawk 1 XAD sample was ranked as having moderate (M) mutagenic activity based upon the EPA Level 1 evaluation criteria. The MEC of 75 μ g per plate closely approaches the moderate/high borderline of 50 μ g per plate. Had sufficient test material been supplied to adequately test the sample, the MEC could have been determined with greater precision.

4. MOHAWK 2 XAD

Only 4 mg of test material was supplied for both Ames and CHO testing. The sample was prepared in 2 ml DMSO giving a primary stock concentration of 2 μ g organics per μ l. The initial range-finding trial was conducted with TA-98 with and without S9 mix at 10 μ l per plate. The nonactivation trial was negative, while the activation trial just reached the threshold for a positive response. The test material was then evaluated in Salmonella strains TA-98 and TA-100 both with and without S9 mix. The dose range used in Trial 2 was 5, 10, 25 and 50 μ l per plate. Both strains appeared to be negative and no dose-related increase in mutant levels was noted. The TA-98 solvent controls in Trial 2 were higher than assay acceptance criteria permit. Trial 3 was conducted at 50 μ l per plate to confirm Trial 2 responses and because of the elevated TA-98 spontaneous mutant background level.



This test material was not mutagenic in either strains TA-98 or TA-100 under test conditions with and without S9 mix. No MEC could be detected under the test conditions employed. The very weakly positive result with strain TA-98 with activation at 10 μ l per plate (Trial 1) was considered to be an anomaly because it was not reproducible and because no dose response was observed. Additional testing at higher concentrations was impossible because of the lack of sufficient test material. Since the sample could not be tested up to the maximum applicable dose (MAD) of 5000 μ g per plate and none of the doses tested were mutagenic, the test material could not be evaluated. The MEC, if the sample is mutagenic, was found to be greater than 100 μ g per plate. The mutagenicity of the sample was interpreted as being undetermined, but moderate or less.

B. Tables

This report is based on the data provided in Tables 1 through 12.



BIONETICS

RESULTS

TABLE I

A. NAME OR CODE DESIGNATION OF THE TEST COMPOUND: TOSCO 2 XAD-OMC (A81-07-011-562,559)

E. SOLVENT: NONE

C. TEST INITIATION DATE: 03/05/82

D. TEST COMPLETION DATE: 03/08/82

F. S-9 LYTH: REKCB

NOTE: CONCENTRATIONS ARE GIVEN IN MICROLITERS PER PLATE

TEST	SPECIES	TISSUE	REVERTS PER PLATE				
			TA-98	1	2		
<hr/>							
NONACTIVATION							
NEGATIVE CONTROL		RAT		67			
NEGATIVE CONTROL		RAT		69			
POSITIVE CONTROL**		RAT		948			
POSITIVE CONTROL**		RAT		971			
TEST COMPOUND	10.00	UL	---	31			
<hr/>							
ACTIVATION							
NEGATIVE CONTROL	RAT	LIVER	72				
NEGATIVE CONTROL	RAT	LIVER	61				
POSITIVE CONTROL***	RAT	LIVER	1921				
POSITIVE CONTROL***	RAT	LIVER	856				
TEST COMPOUND	10.00	UL	RAT	LIVER	0		
<hr/>							

** TA-98 2-NITROFLUORENE

10 μG/PLATE

*** TA-98 2-ANTHRAPINE 2.5 μG/PLATE

RESULTS

TABLE 2

A. NAME OR CODE DESIGNATION OF THE TEST COMPOUND: TOSCO 2 XAD+OMC (A81-07-011-562,559)

B. SOLVENT: NONE

C. TEST INITIATION DATES: 03/15/82

D. TEST COMPLETION DATE: 03/16/82

F. S-9 LOTN: RFK 085

NOTE: CONCENTRATIONS ARE GIVEN IN MICROLITERS PER PLATE

TEST	SPECIES	TISSUE	REVERTANTS PER PLATE					
			TA-98			TA-100		
			1	2	3	1	2	3
NONACTIVATION								
SOLVENT CONTROL	---	---	104		175			
SOLVENT CONTROL	---	---	105		174			
POSITIVE CONTROL**	---	---	508		1121			
POSITIVE CONTROL**	---	---	565		1217			
TEST COMPOUND								
1.00	UL	---	152		203			
5.00	UL	---	226		210			
10.00	UL	---	300		235			
50.00	UL	---	553		307			
ACTIVATION								
SOLVENT CONTROL	RAT	LIVER	97		138			
SOLVENT CONTROL	RAT	LIVER	101		146			
POSITIVE CONTROL***	RAT	LIVER	2245		552			
POSITIVE CONTROL***	RAT	LIVER	2286		1051			
TEST COMPOUND								
1.00	UL	RAT	132		152			
5.00	UL	RAT	144		150			
10.00	UL	RAT	199		147			
50.00	UL	RAT	401		261			

** TA-98 2-NITROFLUORENE
 TA-100 SODIUM AZIDE
 SOLVENT 50 UL/PLATE

*** TA-98 2-ANTHRAMINE 2.5 UG/PLATE
 TA-100 2-ANTHRAMINE 2.5 UG/PLATE

RESULTS

TABLE 3

A. NAME OR CODE DESIGNATION OF THE TEST COMPOUND: TOSCO 2 XAD+OMC(A81-07-011-562,559)
 B. SOLVENT: NONE
 C. TEST INITIATION DATE: 03/19/82
 D. TEST COMPLETION DATE: 03/22/82
 E. S-9 LOTS: REK085

NOTE: CONCENTRATIONS ARE GIVEN IN MICROLITERS PER PLATE

TEST	SPECIES TISSUE	REVERTANTS PER PLATE					
		TA-98			TA-100		
		1	2	3	1	2	3
NONACTIVATION							
SOLVENT CONTROL	RAT LIVER	---	---	27	101		
SOLVENT CONTROL	RAT LIVER	---	---	20	97		
POSITIVE CONTROL**	RAT LIVER	---	---	853	1328		
POSITIVE CONTROL**	RAT LIVER	---	---	848	1367		
TEST COMPOUND							
50.00	UL RAT LIVER	---	---	282	150		
75.00	UL RAT LIVER	---	---	-	427		
ACTIVATION							
SOLVENT CONTROL	RAT LIVER	---	---	27	96		
SOLVENT CONTROL	RAT LIVER	---	---	29	113		
POSITIVE CONTROL***	RAT LIVER	---	---	1551	2327		
POSITIVE CONTROL***	RAT LIVER	---	---	1661	2393		
TEST COMPOUND							
50.00	UL RAT LIVER	---	---	251	206		
75.00	UL RAT LIVER	---	---	-	384		

** TA-98 2-NITROFLUORENE
 TA-100 SODIUM AZIDE
 SOLVENT 75 UL/PLATE
 - INDICATES TEST WAS NOT DONE

*** 10 µG/PLATE TA-98 2-ANTHRAMINE 2.5 µG/PLATE
 10 µG/PLATE TA-100 2-ANTHRAMINE 2.5 µG/PLATE

RESULTS

TABLE 4

A. NAME OR CODE DESIGNATION OF THE TEST COMPOUND: TOSCO 1 XAD (A81-07-011-569)
 B. SOLVENT: NONE
 C. TEST INITIATION DATE: 03/05/82
 D. TEST COMPLETION DATE: 03/08/82
 E. S-9 LOTH: RFK005

NOTE: CONCENTRATIONS ARE GIVEN IN MICROLITERS PER PLATE

TEST	SPECIES	TISSUE	REVERTANTS PER PLATE		
			TA-98	1	2
NONACTIVATION					
NEGATIVE CONTROL	---	---	---	67	
NEGATIVE CONTROL	---	---	---	69	
POSITIVE CONTROL**	---	---	---	948	
POSITIVE CONTROL**	---	---	---	971	
TEST COMPOUND	10.00	UL	---	141	
ACTIVATION					
NEGATIVE CONTROL	RAT	LIVER	72		
NEGATIVE CONTROL	RAT	LIVER	61		
POSITIVE CONTROL***	RAT	LIVER	1921		
POSITIVE CONTROL***	RAT	LIVER	856		
TEST COMPOUND	10.00	UL	RAT	LIVER	165

** TA-98 2-NITROFLUORENE

10 UG/PLATE

*** TA-98 2-ANTHRAMINE 2.5 UG/PLATE

RESULTS

TABLE 5

A. NAME OR CODE DESIGNATION OF THE TEST COMPOUND: TOSCO 1 XAD (A81-07-011-569)

B. SOLVENT: NONE

C. TEST INITIATION DATES: 03/15/82

D. TEST COMPLETION DATE: 03/18/82

E. S-9 LOTN: REK085

NOTE: CONCENTRATIONS ARE GIVEN IN MICROLITERS PER PLATE

TEST	SPECIES TISSUE	REVERTANTS PER PLATE						
		TA-98			TA-100			
		1	2	3	1	2	3	
NONACTIVATION								
SOLVENT CONTROL								
SOLVENT CONTROL	---	---	104		175			
SOLVENT CONTROL	---	---	105		174			
POSITIVE CONTROL**	---	---	508		1121			
POSITIVE CONTROL**	---	---	565		1217			
TEST COMPOUND								
5.00	UL	---	65		190			
10.00	UL	---	106		204			
25.00	UL	---	112		232			
50.00	UL	---	94		290			
ACTIVATION								
SOLVENT CONTROL								
SOLVENT CONTROL	RAT	LIVER	97		138			
SOLVENT CONTROL	RAT	LIVER	101		146			
POSITIVE CONTROL***	RAT	LIVER	2245		932			
POSITIVE CONTROL***	RAT	LIVER	2286		1051			
TEST COMPOUND								
5.00	UL	RAT	LIVER	107	194			
10.00	UL	RAT	LIVER	97	196			
25.00	UL	RAT	LIVER	114	211			
50.00	UL	RAT	LIVER	102	192			

** TA-98 2-NITROFLUORENE
 TA-100 SODIUM AZIDE
 SOLVENT 50 UL/PLATE

10 UG/PLATE
 10 UG/PLATE

*** TA-98 2-ANTHRAMINE 2.5 UG/PLATE
 TA-100 2-ANTHRAMINE 2.5 UG/PLATE

5-153

19

RESULTS

TABLE 6

A. NAME OR CODE DESIGNATION OF THE TEST COMPOUND: TOSCO 1 XAD (A81-07-011-569)

B. SOLVENT: NONE

C. TEST INITIATION DATE: 03/19/82

D. TEST COMPLETION DATE: 03/22/82

E. S-9 LOTH: REK085

NOTE: CONCENTRATIONS ARE GIVEN IN MICROLITERS PER PLATE

TEST	SPECIES	TISSUE	REVERTANTS PER PLATE					
			TA-98			TA-100		
			1	2	3	1	2	3
NONACTIVATION								
SOLVENT CONTROL	---	---	27			101		
SOLVENT CONTROL	---	---	20			97		
POSITIVE CONTROL**	---	---	853			1367		
POSITIVE CONTROL**	---	---	848			1328		
TEST COMPOUND								
50.00	UL	---	---	136		154		
ACTIVATION								
SOLVENT CONTROL	RAT	LIVER	27			56		
SOLVENT CONTROL	RAT	LIVER	29			113		
POSITIVE CONTROL***	RAT	LIVER	1551			2337		
POSITIVE CONTROL***	RAT	LIVER	1661			2353		
TEST COMPOUND								
50.00	UL	RAT	LIVER	199		232		
75.00	UL	RAT	LIVER	-		323		

** TA-98 2-NITROFLUORENE
 TA-100 SODIUM AZIDE
 SOLVENT 75 UL/PLATE
 - INDICATES TEST WAS NOT DONE

*** TA-98 2-ANTHRAMINE 2.5 UG/PLATE
 TA-100 2-ANTHRAMINE 2.5 UG/PLATE

RESULTS

TABLE 7

- A. NAME OR CODE DESIGNATION OF THE TEST COMPOUND: MOHAWKIXAD (A81-09-007-488)
 B. SOLVENT: NONE
 C. TEST INITIATION DATE: 03/05/82
 D. TEST COMPLETION DATE: 03/08/82
 E. S-9 LOT#: RFK085

NOTE: CONCENTRATIONS ARE GIVEN IN MICROLITERS PER PLATE

TEST	SPECIES TISSUE	REVERTANTS PER PLATE		
		1	2	3
NONACTIVATION				
NEGATIVE CONTROL	RAT LIVER	---	---	67
NEGATIVE CONTROL	RAT LIVER	---	---	69
POSITIVE CONTROL**	RAT LIVER	---	---	948
POSITIVE CONTROL**	RAT LIVER	---	---	971
TEST COMPOUND 10.00	UL	---	---	110
ACTIVATION				
NEGATIVE CONTROL	RAT LIVER	---	72	
NEGATIVE CONTROL	RAT LIVER	---	61	
POSITIVE CONTROL***	RAT LIVER	---	1921	
POSITIVE CONTROL***	RAT LIVER	---	1856	
TEST COMPOUND 10.00	UL RAT LIVER	88		

** TA-98 2-NITROFLUORENE

10 ug/plate

*** TA-98 2-ANTHRAMINE 2.5 ug/plate

RESULTS

TABLE 8

- A. NAME OR CODE DESIGNATION OF THE TEST COMPOUND: MOHAWK XAD (A81-09-007-488)
 B. SOLVENT: NONE
 C. TEST INITIATION DATE: 03/15/82
 D. TEST COMPLETION DATE: 03/18/82
 E. S-9 LOT #: RFK085
 NOTE: CONCENTRATIONS ARE GIVEN IN MICROLITERS PER PLATE

TEST	SPECIES TISSUE	P E V E R T A N T S P E R P L A T E					
		TA-98			TA-100		
		1	2	3	1	2	3
NONACTIVATION							
SOLVENT CONTROL	---	---	104		175		
SOLVENT CONTROL	---	---	105		174		
POSITIVE CONTROL**	---	---	508		1121		
POSITIVE CONTROL**	---	---	565		1217		
TEST COMPOUND							
5.00	UL	---	120		230		
10.00	UL	---	150		244		
25.00	UL	---	204		219		
50.00	UL	---	255		250		
ACTIVATION							
SOLVENT CONTROL	RAT	LIVER	97		138		
SOLVENT CONTROL	RAT	LIVER	101		144		
POSITIVE CONTROL**	RAT	LIVER	2245		992		
POSITIVE CONTROL**	RAT	LIVER	2296		1051		
TEST COMPOUND							
5.00	UL	RAT	113		229		
10.00	UL	RAT	174		197		
25.00	UL	RAT	265		284		
50.00	UL	RAT	438		343		

** TA-98 2-NITROFLUORENE
 TA-100 SODIUM AZIDE
 SOLVENT 50 UL/PLATE

10 µG/PLATE TA-98 2-ANTHRAMINE 2.5 µG/PLATE
 10 µG/PLATE TA-100 2-ANTHRAMINE 2.5 µG/PLATE

RESULTS

TABLE 9

A. NAME OR CODE DESIGNATION OF THE TEST COMPOUND: MOHAWK 1 XAD (A81-09-007-488)
 B. SOLVENT: NONE
 C. TEST INITIATION DATES: 03/19/82
 D. TEST COMPLETION DATE: 03/22/82
 E. S-9 LOTS: REKORS

NOTE: CONCENTRATIONS ARE GIVEN IN MICROLITERS PER PLATE

TEST	SPECIES	TISSUE	REVERTANTS PER PLATE					
			TA-98			TA-100		
			1	2	3	1	2	3
NONACTIVATION								
SOLVENT CONTROL	---	---	27			101		
SOLVENT CONTROL	---	---	20			97		
POSITIVE CONTROL**	---	---	853			1328		
POSITIVE CONTROL**	---	---	849			1367		
TEST COMPOUND								
50.00	UL	RAT	---	118		180		
75.00	UL	RAT	---	-		125		
ACTIVATION								
SOLVENT CONTROL	RAT	LIVER	27			96		
SOLVENT CONTROL	RAT	LIVER	29			113		
POSITIVE CONTROL***	RAT	LIVER	1541			2337		
POSITIVE CONTROL***	RAT	LIVER	1601			2393		
TEST COMPOUND								
25.00	UL	RAT	LIVER	113		440		
50.00	UL	RAT	LIVER	235		-		

** TA-98 2-NITROFLUORENE
 TA-100 SODIUM AZIDE
 SOLVENT 75 UL/PLATE
 - INDICATES TEST WAS NOT DONE

*** 10 ug/plate TA-98 2-ANTHRAMINE 2.5 ug/plate
 10 ug/plate TA-100 2-ANTHRAMINE 2.5 ug/plate

RESULTS

TABLE 10

A. NAME OR CODE DESIGNATION OF THE TEST COMPOUND: MOHAWK2 XAD (AB1-09-007-495)
 B. SOLVENT: NONE
 C. TEST INITIATION DATES: 03/05/82
 D. TEST COMPLETION DATE: 03/08/82
 E. S-9 LOTS: REK085

NOTE: CONCENTRATIONS ARE GIVEN IN MICROLITERS PER PLATE

TEST	SPECIES TISSUE	REVERTANTS PER PLATE			
		TA-98	1	2	
NONACTIVATION					
ACTIVATION					
NEGATIVE CONTROL	RAT LIVER	72			
NEGATIVE CONTROL	RAT LIVER	61			
POSITIVE CONTROL**	RAT LIVER	1921			
POSITIVE CONTROL***	RAT LIVER	856			
TEST COMPOUND 10.00	UL	30			
NEGATIVE CONTROL	RAT LIVER	72			
NEGATIVE CONTROL	RAT LIVER	61			
POSITIVE CONTROL***	RAT LIVER	1921			
POSITIVE CONTROL***	RAT LIVER	856			
TEST COMPOUND 10.00	UL RAT LIVER	134			

** TA-98 2-NITROFLUORENE

10 ug/plate

*** TA-98 2-ANTHRAMINE 2.5 ug/plate

RESULTS

TABLE II

- A. NAME OR CODE DESIGNATION OF THE TEST COMPOUND: MOHAWK 2 XAD (A81-09-007-495)
 B. SOLVENT: NONE
 C. TEST INITIATION DATES: 03/15/82
 D. TEST COMPLETION DATE: 03/18/82
 E. S-9 LOTS: RFK045

NOTE: CONCENTRATIONS ARE GIVEN IN MICROLITERS PER PLATE

TEST	SPECIES	TISSUE	REVERTANTS PER PLATE					
			TA-98			TA-100		
			1	2	3	1	2	3
NONACTIVATION								
SOLVENT CONTROL	---	---	104		175			
SOLVENT CONTROL	---	---	105		174			
POSITIVE CONTROL**	---	---	508		1121			
POSITIVE CONTROL**	---	---	565		1217			
TEST COMPOUND								
5.00	UL	---	60		89			
10.00	UL	---	96		121			
25.00	UL	---	98		168			
50.00	UL	---	98		161			
ACTIVATION								
SOLVENT CONTROL	RAT	LIVER	97		138			
SOLVENT CONTROL	RAT	LIVER	101		146			
POSITIVE CONTROL***	RAT	LIVER	2245		992			
POSITIVE CONTROL***	RAT	LIVER	2286		1051			
TEST COMPOUND								
5.00	UL	RAT	LIVER	107	172			
10.00	UL	RAT	LIVER	109	184			
25.00	UL	RAT	LIVER	106	140			
50.00	UL	RAT	LIVER	105	139			

** TA-98 2-NITROFLUORENE
 TA-100 SODIUM A2TDF
 SOLVENT 50 UL/PLATE

10 ug/plate
 10 ug/plate

*** TA-98 2-ANTHRAMINE 2.5 ug/plate
 TA-100 2-ANTHRAMINE 2.5 ug/plate

RESULTS

TABLE 12

- A. NAME OR COD. DESIGNATION OF THE TEST COMPOUND: MOHAWK2 XAD (A81-09-007-495)
 B. SOLVENT: NONE
 C. TEST INITIATION DATE: 03/19/82
 D. TEST COMPLETION DATE: 03/22/82
 E. S-9 LOT#: REFK095
 NOTE: CONCENTRATIONS ARE GIVEN IN MICROLITERS PER PLATE

REVERTANTS PER PLATE

TEST	SPECIES TISSUE	TA-98		
		1	2	3
NONACTIVATION				
SOLVENT CONTROL	---	---	27	
SOLVENT CONTROL	---	---	20	
POSITIVE CONTROL**	---	---	853	
POSITIVE CONTROL***	---	---	849	
TEST COMPOUND	50.00	UL	---	24

ACTIVATION

SOLVENT CONTROL	RAT	LIVER	27		
SOLVENT CONTROL	RAT	LIVER	29		
POSITIVE CONTROL**	RAT	LIVER	1551		
POSITIVE CONTROL***	RAT	LIVER	1681		
TEST COMPOUND	50.00	UL	RAT	LIVER	47

** TA-98 2-NITROFLUORENE
 SOLVENT 75 UL/PLATE

10 UG/PLATE

*** TA-98 2-ANTHRAMINE 2.5 UG/PLATE

VII. ASSAY ACCEPTANCE AND EVALUATION CRITERIA

Statistical methods are not currently used, and evaluation is based on the criteria included in this protocol.

Plate test data consists of direct revertant colony counts obtained from a set of selective agar plates seeded with populations of mutant cells suspended in a semisolid overlay. Because the test material and the cells are incubated in the overlay for approximately 2 days and a few cell divisions occur during the incubation period, the test is semiquantitative in nature. Although these features of the assay reduce the quantitation of results, they provide certain advantages not contained in a quantitative suspension test:

- The small number of cell divisions permits potential mutagens to act on replication DNA, which is often more sensitive than nonreplicating DNA.
- The combined incubation of the test article and the cells in the overlay permits constant exposure of the indicator cells for approximately 2 days.

A. Surviving Populations

Plate test procedures do not permit exact quantitation of the number of cells surviving chemical treatment. At low concentrations of the test material, the surviving population on the treatment plates is essentially the same as that on the negative control plate. At high concentrations, the surviving population is usually reduced by some fraction. Our protocol will normally employ several doses ranging over two or three log concentrations, the highest of these doses being selected to show slight toxicity as determined by subjective criteria.



BIONETICS

B. Dose-Response Phenomena

The demonstration of dose-related increased in mutant counts is an important criterion in establishing mutagenicity. A factor that might modify dose-response results for a mutagen would be the selection of doses that are too low (usually mutagenicity and toxicity are related). If the highest dose is far lower than a toxic concentration, no increases may be observed over the dose range selected. Conversely, if the lowest dose employed is highly cytotoxic, the test material may kill any mutants that are induced, and the test material will not appear to be mutagenic.

C. Control Tests

Positive and negative control assays were conducted with each experiment and consisted of direct-acting mutagens for nonactivation assays and mutagens that require metabolic biotransformation in activation assays. Negative controls consisted of the test material solvent in the overlay agar together with the other essential components. The negative control plate for each strain gave a reference point to which the test data was compared. The positive control assay was conducted to demonstrate that the test systems were functional with known mutagens.

The following normal range of revertants for solvent controls are generally considered acceptable.

TA-1535: 8-30
TA-1537: 4-30
TA-98: 20-75
TA-100: 80-250

D. Evaluation Criteria for Ames Assay

Because the procedures to be used to evaluate the mutagenicity of the test material are semiquantitative, the criteria to be used to determine

positive effects are inherently subjective and are based primarily on a historical data base. Most data sets will be evaluated using the following criteria.

1. Strains TA-1535 and TA-1537

If the solvent control value is within the normal range, a test material that produces a positive dose response over three concentrations with the highest increase equal to three times the solvent control value will be considered to be mutagenic.

2. Strains TA-98 and TA-100

If the solvent control value is within the normal range, a test material that produces a positive dose response over three concentrations with the highest increase equal to twice the solvent control value for TA-98 and TA-100 will be considered to be mutagenic.

3. Pattern

Because TA-1535 and TA-100 are both derived from the same parental strain (G-46), to some extent there is a built-in redundancy in the microbial assay. In general, the two strains of a set respond to the same mutagen and such a pattern is sought. Generally, if a strain responds to a mutagen in nonactivation tests, it will do so in activation tests.

4. Reproducibility .

If a test material produces a response in a single test that cannot be reproduced in additional runs, the initial positive test data lose significance.



BIONETICS

The preceding criteria are not absolute, and other extenuating factors may enter into a final evaluation decision. However, these criteria will be applied to the majority of situations and are presented to aid those individuals not familiar with this procedure. As the data base is increased, the criteria for evaluation can be more firmly established.

E. Relation Between Mutagenicity and Carcinogenicity

It must be emphasized that the Ames Salmonella/Microsome Plate Assay is not a definitive test for chemical carcinogens. It is recognized, however, that correlative and functional relations have been demonstrated between these two endpoints. The results of comparative tests on 300 chemicals by McCann *et al.*⁷ show an extremely good correlation between results of microbial mutagenesis tests and in vivo rodent carcinogenesis assays.

All evaluations and interpretation of the data to be presented in the final report will be based only on the demonstration, or lack, of mutagenic activity.

F. Criteria for Ranking Samples in the Ames Assay

The goal of EPA Level 1 Ames testing is to rank source streams by relative degree of genetic toxicity (mutagenicity). Samples are first identified as mutagenic or nonmutagenic by the criteria in Section D above and then ranked using the mutagenicity categories presented in the table below. The lowest concentration giving a positive response in any strain, with or without metabolic activation, is identified as the minimum effective concentration (MEC) for that sample. The mutagenicity of the sample is evaluated as high (H), moderate (M), low (L), or nondetectable (ND) according to the evaluation criteria developed in the Level 1 manual¹ and summarized below. Samples with no detectable activity at the maximum applicable dose (MAD) are ranked nondetectable (ND).



Another evaluation scheme is proposed for extracts obtained from SASS train gas volumes. The proportion of the total gas volume corresponding to the volume of extract used in the bioassay is calculated and expressed as liters per plate. A criterion of 5000 L/plate is set as the limit for nondetectable toxicity. The subsequent toxicity ranges are defined by 10-fold dilution steps to conform to standard procedure. Evaluation criteria based on equivalent gas volumes are tentative and under evaluation.

Ames Assay Mutagenicity Ranking Criteria¹

Mutagenic Activity	Solids (MEC in µg/plate)	Liquids ^a (MEC in µl/plate)	Equivalent Gas Volumes (MEC in liters/plate)
High (H)	<50	<2	<50
Moderate (M)	50-500	2-20	50-500
Low (L)	500-5000	20-200	500-5000
Not Detectable (ND)	>5000	>200	>5000

^aConcentration of organic extracts is based upon organic content (µg organics per plate) and not volume (µl extract per plate) of sample tested.



BIONETICS

VIII. REFERENCES

1. Brusick, D.J. and Young, R.R.: IERL-RTP Procedures Manual: Level 1 Environmental Assessment Biological Tests. EPA-600/8-81-024, Litton Bionetics, Inc., Kensington, MD, October, 1981, 150 pp.
2. Brusick, D.J.: Level 1 Bioassay Assessment and Data Formatting. EPA-600/7-80-079, Litton Bionetics Inc., Kensington, MD, April 1980; 100 pp.
3. Brusick, D.J. and Young, R.R.: Level 1 Bioassay Sensitivity. EPA-600/7-81-135, Litton Bionetics, Inc., Kensington, MD, August 1981, 52 pp.
4. DHEW Food and Drug Administration "Nonclinical Laboratory Studies; Good Laboratory Practice Regulations." Federal Register, volume 43, No. 247, pp. 59986-60020, Part II, December 22, 1978.
5. Proposed Health Effects Test Standard for Toxic Substances Control Act Test Rules; Good Laboratory Practice Standards for Health Effects. Federal Register, Part II, volume 44, No. 91, May 1979 and Part IV, volume 44, No. 145, July, 1979.
6. Guidelines for Registering Pesticides in the United States: Proposed Good Laboratory Practice Guidelines for Toxicology Testing. Federal Register, volume 45, No. 77, April, 1980.
7. McCann, J., Choi, E., Yamasaki, E. and Ames, B.N.: Detection of carcinogens as mutagens in the Salmonella/microsome test: Assay of 300 chemicals. Proc. Nat. Acad. Sci., USA 72:5135-5139, 1975.
8. Ames, B.N., Gurney, E.G., Miller, J.A. and Bartsch, H.: Carcinogens as frameshift mutagens: Metabolites and derivatives of 2-acetylaminofluorene and other aromatic amine carcinogens. Proc. Nat. Acad. Sci., USA 69:3128-3132, 1972.



9. Ames, B.N., Lee, F.D., and Durston, W.E.: An improved bacterial test system for the detection and classification of mutagens and carcinogens. Proc. Nat. Acad. Sci., USA 70:782-786, 1973.
10. Ames, B.N., Durston, W.E., Yamasaki, E. and Lee, F.D.: Carcinogens are mutagens: A simple test system combining liver homogenates for activation and bacteria for detection. Proc. Nat. Acad. Sci., USA 70:2281-2285, 1973.
11. McCann, J., Springarn, N.E., Kobori, J. and Ames, B.N.: Detection of carcinogens as mutagens: Bacterial tester strains with R factor plasmids. Proc. Nat. Acad. Sci. USA 72:979-983, 1975.
12. Ames, B.N., McCann, J. and Yamasaki, E.: Methods for detecting carcinogens and mutagens with the Salmonella/mammalian-microsome mutagenicity test. Mutation Res., 31:347-364, 1975.
13. Vogel, H.J. and Bonner, D.M.: Acetylornithinase of E. coli partial purification and some properties. J. Biol. Chem., 218:97-106, 1966.



GENETICS ASSAY NOS. 6153-6156
LBI SAFETY NOS. 7535-7538

CYTOTOXIC EVALUATION OF
XAD RESIN EXTRACTS
IN THE
EPA LEVEL 1
RODENT CELL (CHO)
CLONAL TOXICITY ASSAY

FINAL REPORT

SUBMITTED TO:

ACUREX CORPORATION
485 CLYDE AVENUE
MOUNTAIN VIEW, CALIFORNIA 94042

SUBMITTED BY:

LITTON BIONETICS, INC.
5516 NICHOLSON LANE
KENSINGTON, MARYLAND 20895

LBI PROJECT NO. 22064

REPORT DATE: MARCH 1982

5-168

PREFACE

These assays conform to the standard EPA Level 1 procedure for the Chinese hamster ovary cell (CHO) clonal toxicity assay as described in "IERL-RTP Procedures Manual: Level 1 Environmental Assessment Biological Tests"¹. The data were evaluated and formatted as recommended in "Level 1 Biological Testing Assessment and Data Formatting"².

The CHO clonal toxicity assay has been shown to be a sensitive method for detecting cytotoxic activity for a variety of chemicals representing various chemical classes ³. This assay is one of several recommended by EPA to identify, categorize and rank the pollutant potential of influent and effluent streams from industrial and energy-producing processes. This assay has been well validated with a wide range of positive and negative control chemicals and complex environmental samples.

All procedures and documents pertaining to the receipt, storage, preparation, testing and evaluation of the test material shall conform to Litton Bionetics, Inc. standard operating procedures, the U.S. Food and Drug Administration's Good Laboratory Practices Regulations of 1979⁴ and the proposed U.S. Environmental Protection Agency's Good Laboratory Practice Guidelines.^{5,6} Deviations from standard procedure shall be fully documented and noted in the report.

All test and control results in this report are supported by fully documented raw data which are permanently maintained in the files of the Department of Molecular Toxicology or in the archives of Litton Bionetics, Inc., 5516 Nicholson Lane, Kensington, Maryland 20895. Copies of raw data will be supplied to the sponsor upon request.

TABLE OF CONTENTS

Page No.

PREFACE	i
I. ASSAY SUMMARY	1
II. OBJECTIVE	2
III. TEST MATERIAL	3
A. Description	3
B. Handling and Preparation	3
IV. MATERIALS	5
A. Indicator Cells	5
B. Media	5
C. Controls	6
V. EXPERIMENTAL DESIGN	7
A. Dose Selection	7
B. Clonal Toxicity Assay	7
VI. RESULTS	9
A. Interpretation	9
1. TOSCO 2 XAD + OMC	9
2. TOSCO 1 XAD	9
3. MOHAWK 1 XAD	10
4. MOHAWK 2 XAD	11
B. Tables and Figures	12
VII. ASSAY ACCEPTANCE CRITERIA	21
VIII. ASSAY EVALUATION CRITERIA	22
IX. REFERENCES	24



BIONETICS

I. SUMMARY

Four resin extract sample supplied by Acurex Corporation were tested and evaluated for their cytotoxicities in the EPA Level 1 Chinese hamster ovary (CHO) cell clonal toxicity assay. The samples, supplied in methylene chloride, were solvent exchanged to dimethylsulfoxide (DMSO) prior to testing. The samples were identified as TOSCO 2 XAD + OMC (A81-07-011-562, 559), TOSCO 1 XAD (A81-07-011-569), MOHAWK 1 XAD (A81-09-007-488) and MOHAWK 2 XAD (A81-09-007-495).

TOSCO 2 XAD + OMC and TOSCO 1 XAD were both ranked as having moderate toxicity with EC₅₀ values of 70.8 µg organics/ml and 37.4 µg organics/ml, respectively. CHO toxicity testing was limited by the small amount of test material supplied for MOHAWK 1 XAD (6 mg organics) and MOHAWK 2 XAD (4 mg organics). EC₅₀ values could not be located for these two samples. No toxicity was observed up to the maximum applicable dose (MAD) of 20 µl/ml (60 µg organics/ml) with MOHAWK 1 XAD and only slight toxicity at the MAD of 20 µl/ml (40 µg organics/ml) with MOHAWK 2 XAD. Therefore, the toxicities of these two samples were evaluated as being undetermined but moderate or less.

The results indicate that quantities less than 10 mg organics of this type of sample were not sufficient for adequate Level 1 testing. While minimum testable sample size is often a function of the biological activity of the sample, efforts should be made to supply at least 20 mg of SASS train organics for combined Ames and CHO testing.

Submitted by:

Study Director

Robert Young

Robert Young, M.S.
Study Director,
Environmental Assessment
Section
Department of Molecular
Toxicology

3-25-82
Date

Reviewed by:

David J. Brusick
David J. Brusick, Ph.D.
Director,
Department of Molecular
Toxicology

4/7/82
Date



BIONETICS

II. OBJECTIVE

The objective of this study was to determine and rank the cytotoxicities of four resin extract samples to cultured Chinese hamster cells (CHO-K1 cell line). The samples were solvent exchanged into dimethylsulfoxide (DMSO) before CHO testing was initiated. The measure of cytotoxicity was the reduction in colony-forming ability of CHO-K1 cells after a 24-hour exposure to the test material. After a period of recovery and growth, the number of colonies that developed in the treated cultures was compared to the colony number in unexposed vehicle control cultures. The concentration of test material that reduced the colony number by 50 percent was estimated graphically and referred to as the EC₅₀ value. Standard EPA Level 1 toxicity evaluation criteria for the CHO clonal toxicity assay were used to rank the toxicity potential of each test material.

III. TEST MATERIAL

A. Description

Four samples were supplied by Acurex Corporation, Mountain View, California. The samples were assigned LBI safety numbers and LBI assay numbers upon receipt. The Acurex code numbers, sample identification, LBI safety numbers and LBI assay numbers are identified below. All laboratory documentation used the LBI assay number to identify samples.

The four test materials were received as clear, yellow solutions of organic material in methylene chloride except for the TOSCO 1 XAD sample. That sample was an amber-colored, clear solution with a few suspended particles. The quantity of organic material in each sample, as determined by Acurex Corporation, is identified below. No information on sampling parameters (such as the equivalent volume of stack gas represented by the sample) was provided.

Acurex Corp. Code	Sample Identification	Quantity (mg organic)	LBI Safety No.	LBI Assay No.
A81-07-011-562, 559	TOSCO 2 XAD + OMC	12	7537	6153
A81-07-011-569	TOSCO 1 XAD	17.	7538	6154
A81-09-007-488	MOHAWK 1 XAD	6	7535	6155
A81-09-007-495	MOHAWK 2 XAD	4	7536	6156

B. Handling and Preparation

The test materials were received at LBI on February 8, 1982. The samples were shipped in small, clear-glass vials sealed with crimp-top aluminum caps with rubber liners. The samples were received intact and were stored at +4°C in the dark until processed.

Pretest sample preparation consisted of solvent exchanging the samples into dimethylsulfoxide (DMSO). The samples were transferred with methylene chloride rinses into graduated conical tubes. The methyl chloride was gradually evaporated (50°C under a stream of nitrogen) and DMSO was sequentially added. The samples were brought to volume in 2.0 ml of DMSO. The samples were transferred to glass vials and sealed with teflon-coated rubber rounds. The solvent exchanged samples were stored at +4°C in the dark.

A total volume of 0.42 ml of each test sample was used in the CHO assays. The maximum concentration of 20 µl/ml was obtained by adding 0.12 ml of sample to 5.88 ml of F12 medium; this resulted in 2 percent (v/v) DMSO in the medium and effectively limited the concentration of test material that could be assayed. Only two plates were dosed at the top dose in order to conserve sample. Another 0.12 ml aliquot of sample was added to 11.88 ml of F12 medium to prepare the 10 µl/ml test concentration. An additional 0.18 ml of test sample was used to prepare a series of dilutions in DMSO from which 1:100 dilutions into growth medium were performed to obtain the lower assayed concentrations. Thus, except for the 20 µl/ml test concentrations, the final DMSO concentration was constant at 1 percent (v/v).



BIONETICS

IV. MATERIALS

A. Indicator Cells

The indicator cells for these assays were Chinese hamster CHO-K1 cells (ATCC No. CCL 61) obtained from Flow Laboratories, Inc., Rockville, MD. This cell type was derived from ovarian tissue and has spontaneously transformed to a stable, hypodiploid line of rounded, fibroblastic cells with unlimited growth potential. Monolayer cultures have a fast doubling time of 11 to 14 hours, and untreated cells can normally be cloned with an efficiency of 80 percent or greater. Laboratory stocks were maintained by routine serial subpassage. Cells were cultivated in Ham's F-12 nutrient medium at 37°C in an atmosphere of 5 percent CO₂ and saturated humidity. Stocks were continually observed macroscopically and microscopically for possible microbial contamination. Laboratory cultures were periodically checked by culturing and staining methods for the absence of mycoplasma contamination. Laboratory cultures were discarded every three months and new cultures started from mycoplasma-free, long-term frozen cultures.

B. Media

The CHO-K1 cell line has an absolute requirement for proline and therefore must be maintained in culture medium containing sufficient amounts of this amino acid. Ham's F12 medium, which contains 3×10^{-4} M L-proline was used, supplemented with 10 percent fetal bovine serum, 2mM L-glutamine, 100 units/ml of penicillin, 100 µg/ml of streptomycin, and 0.9 µl/ml of amphotericin B. .

C. Controls

The negative control for all four assays consisted of three untreated cultures in F12 medium carried through the same experimental time period as the treated cells. Since the test materials were tested as solutions in an organic vehicle (DMSO) and were diluted into the medium to provide each test concentration, two sets of vehicle control cultures containing



BIONETICS

DMSO at one percent and two percent by volume were prepared in triplicate for each assay. The average number of colonies in the negative control established the cloning efficiency of the CHO cells used in the assays and the appropriate vehicle controls provided the reference points for determining the effects of different concentrations of the solid test materials on cell survival.



BIONETICS

5-176

V. EXPERIMENTAL DESIGN

A. Dose Selection

Unless the approximate toxicity is already known, or the sample size is limiting, the following minimum dose ranges are recommended by the Level 1 manual¹ for testing different sample forms. Aqueous samples, suspensions, or slurries are tested from 600 µl/ml to 6 µl/ml, usually in five dose steps. Dry, particulate material is dissolved or suspended in culture medium and tested at five dose levels from 1000 µg/ml to 10 µg/ml. Samples that are solvent-exchanged into DMSO are tested from 20 µl/ml to 0.5 µl/ml, in five dose steps. Eight doses are often used when the amount of test sample is limited to provide a more precise description of toxicity in the event of sharp dose-response curves. A second dose study is performed with an adjusted dose range if the EC₅₀ was not located properly in the initial test. However, EC₅₀ values greater than 1000 µg/ml for particulate material, 600 µl/ml for aqueous samples, or 20 µl/ml for organic solutions will not be determined.

In this work the test materials were tested as resin extracts in an organic vehicle. The concentrations used for the samples started with the maximum applicable dose (MAD) of 20 µl/ml and included seven other doses that were 10, 6, 3, 1, 0.6, 0.3 and 0.1 µl/ml.

B. Clonal Toxicity Assay

Cells from monolayer stock cultures in logarithmic growth phase were trypsinized with 0.1 percent trypsin plus 0.01 percent versene for 4 minutes, and the density of the resulting cell suspension was determined by hemocytometer. A number of 60-mm culture dishes were then seeded with 200 cells and 4 ml of culture medium per dish. The cultures were incubated for approximately 6 hours at 37°C in a humidified atmosphere containing 5 percent CO₂ to allow attachment of the cells. The 6-hour attachment period was used in order to avoid cell division and the subsequent formation of two-cell colonies prior to treatment.



BIONETICS

The medium was aspirated from the cultures and 4 ml of control medium or medium containing the test material was applied. Three cultures were exposed to each test concentration. After an exposure time of 24 hours at 37°C, the medium was removed by aspiration and each culture washed two times with approximately 4 ml aliquots of Dulbecco's phosphate buffered saline (pre-warmed to 37°C). Fresh culture medium (5 ml) was placed in each dish, and incubation at 37°C was continued for an additional 6 days to allow colony development.

If the test material caused a color change in the culture medium, the pH of the medium containing the high dose would be determined at the time of treatment. The pH at the lowest dose that results in a slight color change would also be recorded. At the end of the treatment period, the pH values of the discarded media from the two described treatments would be recorded again. No sample-related pH changes were noted in any of the treatments.

After the incubation period, the medium was drained from the cultures, and the surviving colonies were fixed with 100 percent ethanol and stained with Giemsa. Colonies were counted by eye; tiny colonies of approximately 50 cells or less were arbitrarily excluded from the counts.



BIONETICS

VI. RESULTS

A. Interpretation

The results of the Chinese hamster ovary (CHO) clonal toxicity assays are presented in Tables 1 through 4. The calculated relative survival values were obtained by comparing the average number of colonies per dish for each of the assayed concentrations to the appropriate control value. The relative survival values were then plotted as functions of the applied concentration of test material per ml of culture medium in Figures 1 through 4. Curves were fitted to the data points by eye in order to determine the EC₅₀ value for each sample and to rank each sample according to EPA Level 1 evaluation criteria presented in Section VIII.

1. TOSCO 2 XAD + OMC (A81-07-011-562, 559)

The application of the DMSO solution of the TOSCO 2 XAD + OMC sample (A81-07-011-562, 559) to the CHO cell cultures caused a rapid lowering of the number of cells able to form colonies as the concentration was increased above 6.0 µl/ml. As shown in Figure 1, the relative survival remained above 90 percent in the 0.1 to 6.0 µl/ml range but dropped to zero at the 20.0 µl/ml dose level. The full range of survival was expressed between 6 µl/ml (>90 percent relative survival) and 20 µl/ml (0 percent relative survival).

The concentration expected to kill 50 percent of the cells (EC₅₀) was found to be 11.8 µl of test material per ml of culture medium. The concentration was equivalent to 70.8 µg of organic material per ml of culture medium. This value placed the test material in the moderate (M) toxicity range defined for the IERL-EPA CHO clonal toxicity bioassay¹.

2. TOSCO 1 XAD (A81-07-011-569)

The TOSCO 1 XAD sample (A81-07-011-569) in DMSO was found to be toxic to CHO cells in culture at concentrations above 1.0 µl/ml. The



BIONETICS

ability of CHO cells to form colonies was unaffected by a 24-hour exposure to concentrations of test material between 0.1 and 1.0 μ l/ml. The relative survival values covered the full range of toxicity between 1 μ l/ml (104.3 percent relative survival) and 10 μ l/ml (0 percent relative survival).

The EC₅₀ was found to be 4.4 μ l of test material per ml of culture medium. This concentration was equivalent to 37.4 μ g of organic material per ml of culture medium. The test material was ranked as having moderate (M) toxicity based upon the evaluation criteria developed for the IERL-EPA CHO clonal toxicity bioassay¹.

3. MOHAWK 1 XAD (A81-09-007-488)

The exposure of CHO cells in culture to the MOHAWK 1 XAD sample (A81-09-007-488) in DMSO essentially caused no decrease in the number of cells able to form colonies as the concentration was increased to the maximum applicable dose (MAD) of 20 μ l/ml. As shown in Figure 3 the relative survival to all of the treatments remained above 90 percent.

Since none of the tested doses caused killing that even approached 50 percent of the cells, an EC₅₀ could not be estimated. However, the lack of toxicity at the MAD of 20 μ l/ml (60 μ g organics/ml) excluded the sample from the high toxicity category and from at least 60 percent of the moderate toxicity range, based on the IERL-RTP evaluation criteria¹. Because very sharp and well-defined toxicity curves have been observed with similar samples, it is possible that if sufficient test material had been supplied, toxicity could have been observed in the moderate or low toxicity ranges. The toxicity of the sample was therefore evaluated as undetermined but moderate (M) or less.

Testing and evaluating materials such as this sample indicate the need to supply sufficient quantities of test material to ensure adequate testing. While minimum testable sample size is often a function of the biological activity of the sample, efforts should be made to supply at least 20 mg of SASS train organics for combined Ames and CHO testing.

4. MOHAWK 2 XAD (A81-09-007-495)

The application of DMSO solution of the MOHAWK 2 XAD sample (A81-09-007-495) to the CHO cells in culture caused only a small decrease in the number of cells able to form colonies as the concentration was increased to the maximum applicable dose (MAD) of 20 μ l/ml. Relative survival remained above 90 percent between 0.1 and 3 μ l/ml. Between 3 and 10 μ l/ml there was a noticeable increase in sample toxicity. As shown in Figure 4, the relative survival decreased to about 72 percent for the 20 μ l/ml treatment.

Since none of the tested doses caused killing approaching 50 percent of the cells, an EC₅₀ could not be calculated. However, an EC₅₀ value greater than 20 μ l/ml (40 μ g organics/ml) excluded the sample from the high toxicity category and nearly half of the moderate toxicity categories based on the IERL-RTP evaluation criteria¹. The plotted results suggested the possibility of an EC₅₀ in the 30 to 90 μ l/ml range (60 to 180 μ g organics/ml), so the sample could have been evaluated as having moderate or low toxicity had sufficient sample been available. The toxicity of the sample was therefore evaluated as undetermined but moderate (M) or less.

Testing and evaluating materials such as this sample and MOHAWK 1 XAD indicate the need to supply sufficient quantities of test material to ensure adequate testing. Only 4 mg of organic material was supplied for both Ames and CHO testing - an insufficient quantity for adequate testing for all but the most mutagenic and toxic samples.

The cells used for the four assays were in logarithmic growth phase and the cells in suspension prior to cell plating were 99.6 percent viable as determined by trypan blue dye exclusion. Greater than 77 percent of the seeded cells formed colonies in the negative controls for each of the four assays (80.5 ± 2.9 percent). Colony growth was normal and well distributed on the culture dishes in all trials. The combined results achieved the assay acceptance criteria discussed in Section VII, which



BIONETICS

provided confidence in the assumption that the recorded data represented typical responses to the test materials.

B. Tables and Figures

This report is based on the data provided in Tables 1 through 4 and Figures 1 through 4.



BIONETICS

5-182

12

TABLE I
CLONAL CYTOTOXICITY ASSAY

SAMPLE IDENTITY: TOSCO 2 XAD + OMC
(AB1-07-011-562, 559)

DESCRIPTION OF SAMPLE: CLEAR, YELLOW LIQUID

LBI ASSAY NO. 6153

DATE RECEIVED: FEBRUARY 8, 1982

TEST DATE: MARCH 4, 1982

VEHICLE: DIMETHYLSULFOXIDE (DMSO)

CELL TYPE: CHO-K1

CELLS SEEDED PER DISH: 200

EC50 VALUE: 70.0 μ G/ML (11.8 μ L/ML)

TOXICITY
CLASSIFICATION: MODERATE (M)

PH ALTERATIONS: NONE

COMMENTS ON TREATMENT: 12 MG OF ORGANIC MATERIAL WERE SUPPLIED FOR TESTING. SAMPLE WAS SOLVENT EXCHANGED INTO 2.0 ML DMSO GIVING A PRIMARY STOCK CONCENTRATION OF 6 μ G ORGANIC PER μ L OF DMSO.

5-183

13

COLONY COUNTS

	APPLIED CONCENTRATION μ L/ML	DISH #1	DISH #2	DISH #3	AVERAGE COUNT	RELATIVE SURVIVAL* (PERCENT)	CLONING EFFICIENCY (PERCENT)
NC	---	165	180	162	169.0	100.0	84.5
VC, 1%	10	144	167	157	156.0	100.0	78.0
VC, 2%	20	156	155	149	153.3	100.0	76.7
TEST	0.1	160	167	161	162.7	104.3	
TEST	0.3	169	157	149	158.3	101.5	
TEST	0.6	165	159	156	160.0	102.6	
TEST	1.0	146	159	152	152.3	97.6	
TEST	3.0	153	158	143	151.3	97.0	
TEST	6.0	149	142	152	147.7	94.7	
TEST	10.0	104	114	105	107.7	69.0	
TEST	20.0	0	0	5	0.0	0.0	

NC = NEGATIVE CONTROL, F12 MEDIUM

VC = VEHICLE CONTROL, PERCENT GIVEN FOR DIMETHYLSULFOXIDE (DMSO)

S = PLATE NOT SET UP TO CONSERVE LIMITED SAMPLE

*RELATIVE TO 2% VC FOR 20 μ L/ML TREATMENT AND TO 1% VC FOR OTHER TREATMENTS

TABLE 2
CLONAL CYTOTOXICITY ASSAY

SAMPLE IDENTITY: TOSCO 1 XAD
(ARI-07-011-569)

DESCRIPTION OF SAMPLE: CLEAR, YELLOW LIQUID

LBT ASSAY NO. 6154

DATE RECEIVED: FEBRUARY 8, 1982

TEST DATE: MARCH 4, 1982

VEHICLE: DIMETHYL SULFOXIDE (DMSO)

CELL TYPE: CHO-K1

CELLS SEeded PER DISH: 200

EC50 VALUE: 37.4 UG/ML (4.4 UL/ML)

TOXICITY
CLASSIFICATION: MODERATE (M)

PH ALTERATIONS: NONE

COMMENTS ON TREATMENT: 17 MG OF ORGANIC MATERIAL WERE SUPPLIED FOR TESTING. SAMPLE WAS SOLVENT EXCHANGED INTO 2.0 ML DMSO GIVING A PRIMARY STOCK CONCENTRATION OF 8.5 UG ORGANIC PER UL DMSO.

5-184

14

COLONY COUNTS

SAMPLE	APPLIED CONCENTRATION UL/ML	DISH #1	DISH #2	DISH #3	AVERAGE COUNT	RELATIVE SURVIVAL* (PERCENT)	CLONING EFFICIENCY (PERCENT)
NC	---	144	169	165	159.3	100.0	79.7
VC, 1%	10	154	142	145	147.0	100.0	73.5
VC, 2%	20	154	144	140	146.0	100.0	73.0
TEST	0.1	154	148	140	147.3	100.2	
TEST	0.3	138	153	149	146.7	99.8	
TEST	0.6	137	146	156	146.3	99.5	
TEST	1.0	148	150	149	149.0	101.4	
TEST	3.0	115	127	120	120.7	82.1	
TEST	6.0	38	38	27	34.3	23.3	
TEST	10.0	0	0	0	0.0	0.0	
TEST	20.0	0	0	S	0.0	0.0	

NC = NEGATIVE CONTROL, F12 MEDIUM

VC = VEHICLE CONTROL. PERCENT GIVEN FOR DIMETHYLSULFOXIDE (DMSO)

S = PLATE NOT SET UP TO CONSERVE LIMITED SAMPLE

*RELATIVE TO 2% VC FOR 20 UL/ML TREATMENT AND TO 1% VC FOR OTHER TREATMENTS

TABLE 3
CLONAL CYTOTOXICITY ASSAY

SAMPLE IDENTITY: MOHAWK 1 XAD
(ARL-09-007-48B)

EC50 VALUE: > 60 μ G/ML (> 20 μ L/ML)

TOXICITY
CLASSIFICATION: UNDETERMINED

pH ALTERATIONS: NONE

LBI ASSAY NO. 6155

COMMENTS ON TREATMENT: 6 MG OF ORGANIC MATERIAL WERE
SUPPLIED FOR TESTING. SAMPLE WAS SOLVENT EXCHANGED
INTO 2.0 ML DMSO GIVING A PRIMARY STOCK CONCENTRATION
OF 3 μ G ORGANIC PER μ L DMSO.

DATE RECEIVED: FEBRUARY 8 1982

TEST DATE: MARCH 4, 1982

VEHICLE: DIMETHYLSULFOXIDE (DMSO)

CELL TYPE: CHO-K1

CELLS SEeded PER DISH: 200

COLONY COUNTS

SAMPLE	APPLIED CONCENTRATION μ L/ML	COLONY COUNTS			AVERAGE COUNT	RELATIVE SURVIVAL* (PERCENT)	CLONING EFFICIENCY (PERCENT)
		DISH #1	DISH #2	DISH #3			
NC	---	160	171	150	160.3	100.0	80.2
VC, 1%	1.0	135	148	149	144.0	100.0	72.0
VC, 2%	2.0	137	151	146	144.7	100.0	72.4
TEST	0.1	142	150	157	149.7	104.0	
TEST	0.3	153	135	140	142.7	99.1	
TEST	0.6	142	150	140	144.0	100.0	
TEST	1.0	130	143	132	135.0	93.8	
TEST	3.0	129	124	129	127.3	88.4	
TEST	6.0	131	129	135	131.7	91.5	
TEST	10.0	132	129	130	130.3	90.5	
TEST	20.0	135	137	S	136.0	94.0	

NC = NEGATIVE CONTROL, F12 MEDIUM

VC = VEHICLE CONTROL, PERCENT GIVEN FOR DIMETHYLSULFOXIDE (DMSO)

S = PLATE NOT SET UP TO CONSERVE LIMITED SAMPLE

*RELATIVE TO 2% VC FOR 20 μ L/ML
TREATMENT AND TO 1% VC FOR OTHER
TREATMENTS

TABLE 4
CLONAL CYTOTOXICITY ASSAY

SAMPLE IDENTITY: MOHAWK 2 XAD
(AB1-09-007-495)

DESCRIPTION OF SAMPLE: CLEAR, YELLOW LIQUID

LBI ASSAY NO. 6156

DATE RECEIVED: FEBRUARY 8, 1982

TEST DATE: MARCH 4, 1982

VEHICLE: DIMETHYLSULFOXIDE (DMSO)

CELL TYPE: CHO-K1

CELLS SEeded PER DISH: 200

EC50 VALUE: 60 TO 180 UG/ML (30 TO 90UL/ML)

TOXICITY
CLASSIFICATION: UNDETERMINED

PH ALTERATIONS: NONE

COMMENTS ON TREATMENT: 4 MG OF ORGANIC MATERIAL WERE SUPPLIED FOR TESTING. SAMPLE WAS SOLVENT EXCHANGED INTO 2.0 ML DMSO GIVING A PRIMARY STOCK CONCENTRATION OF 2 UG ORGANIC PER UL DMSO.

5-186

91

COLONY COUNTS

SAMPLE	APPLIED CONCENTRATION UL/ML	DISH #1	DISH #2	DISH #3	AVERAGE COUNT	RELATIVE SURVIVAL* (PERCENT)	CLONING EFFICIENCY (PERCENT)
NC	---	164	153	149	155.3	100.0	77.7
VC, 1%	10	143	148	156	149.0	100.0	74.5
VC, 2%	20	150	147	148	148.3	100.0	74.2
TEST	0.1	148	153	156	152.3	102.2	
TEST	0.3	139	153	149	147.0	98.7	
TEST	0.6	143	139	145	142.3	95.5	
TEST	1.0	138	142	149	143.0	96.0	
TEST	3.0	133	143	144	140.0	94.0	
TEST	6.0	125	134	137	132.0	88.6	
TEST	10.0	113	128	123	121.3	81.4	
TEST	20.0	103	110	S	106.5	71.8	

NC = NEGATIVE CONTROL, F12 MEDIUM

VC = VFHTCLF CONTROL, PERCENT GIVEN FOR DIMETHYLSULFOXIDE (DMSO)

S = PLATE NOT SET UP TO CONSERVE LIMITED SAMPLE

*RELATIVE TO 2% VC FOR 20 UL/ML TREATMENT AND TO 1% VC FOR OTHER TREATMENTS

FIGURE 1

RODENT CELL (CHO) CLONAL TOXICITY ASSAY

EC₅₀ DETERMINATION

TOSCO 2 XAD + OMC

A81-07-011-562, 559

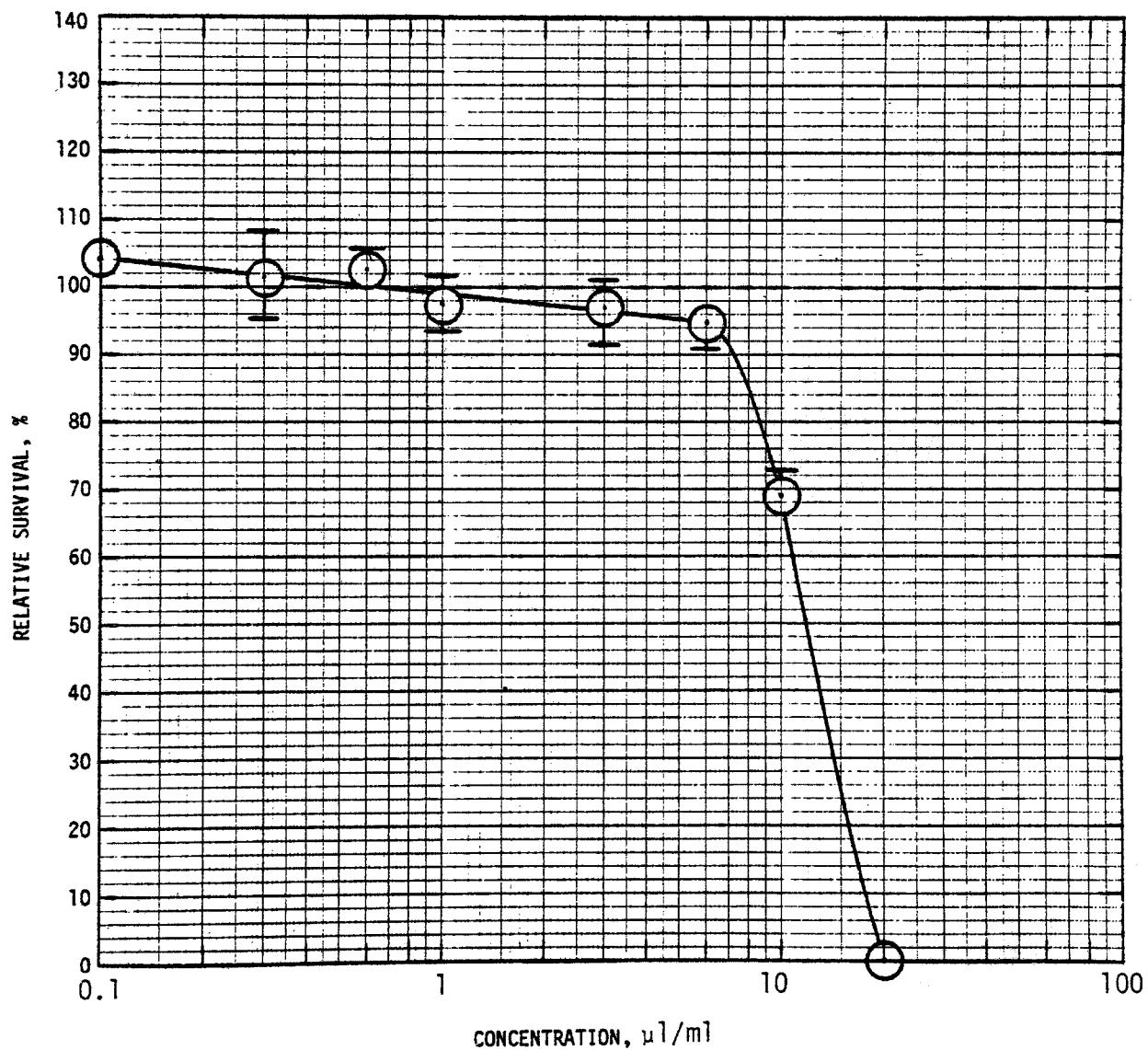


FIGURE 2

RODENT CELL (CHO) CLONAL TOXICITY ASSAY

EC₅₀ DETERMINATION

TOSCO 1 XAD

A81-07-011-569

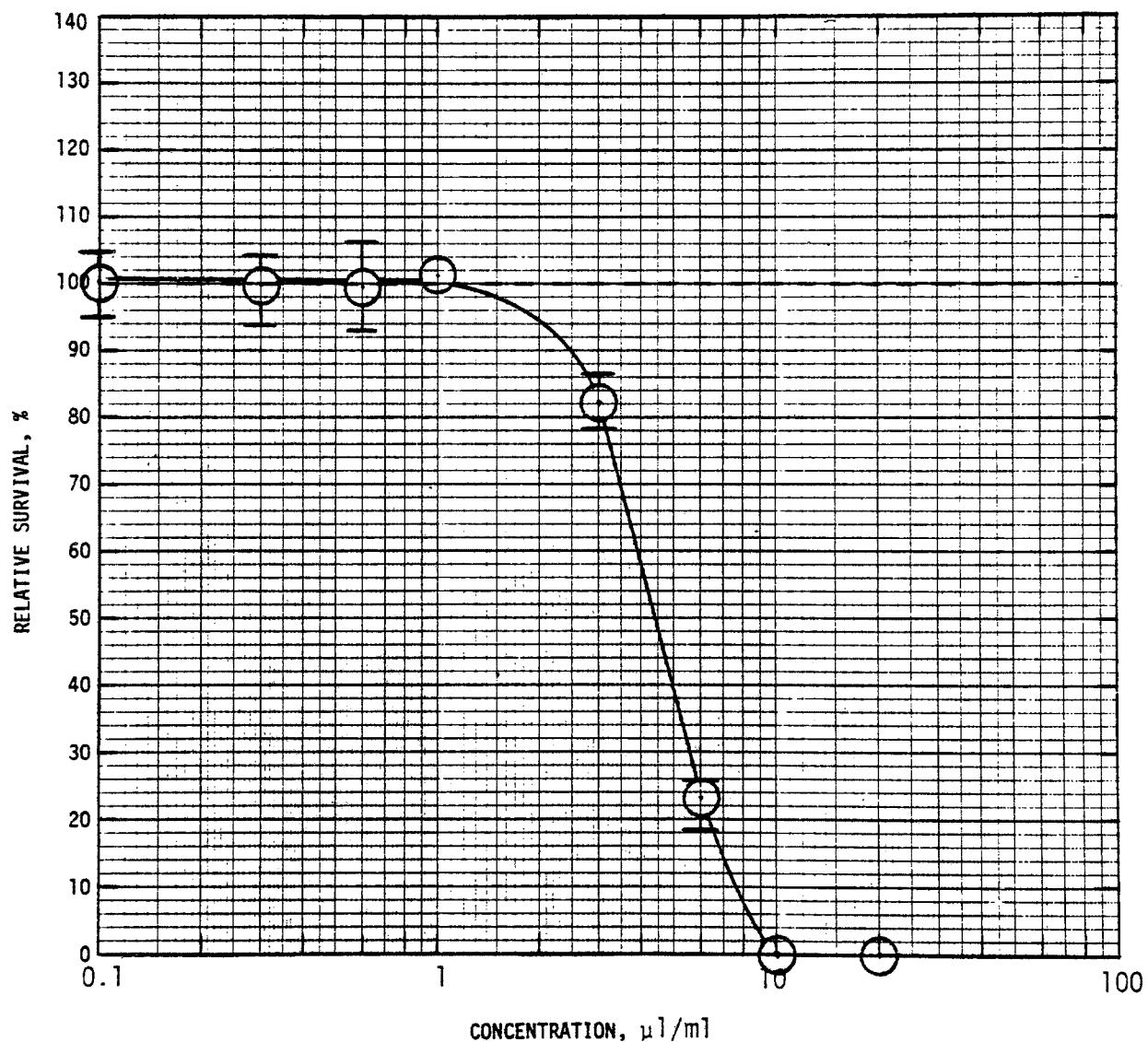


FIGURE 3

RODENT CELL (CHO) CLONAL TOXICITY ASSAY

EC₅₀ DETERMINATION

MOHAWK 1 XAD

A81-09-007-488

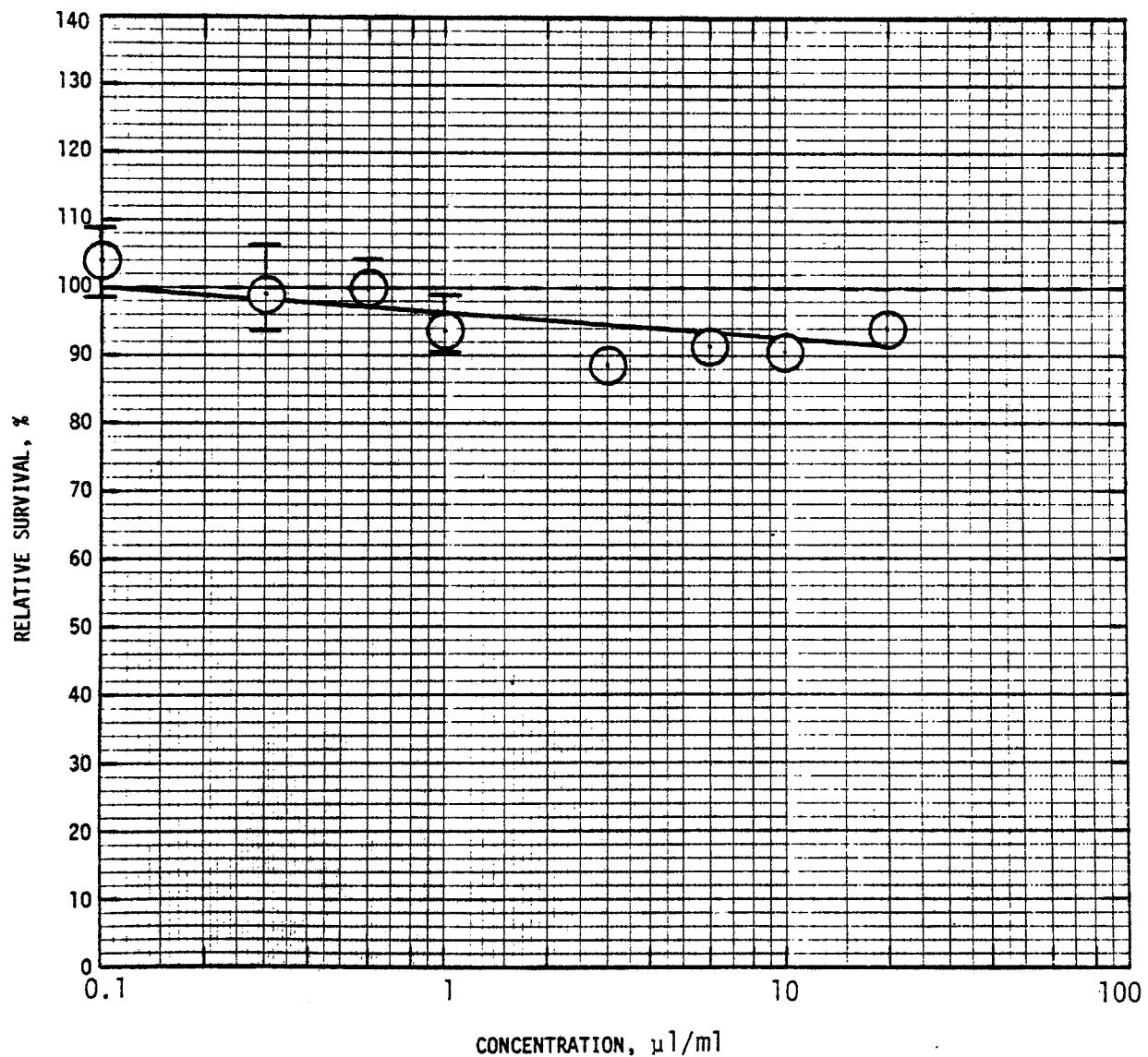
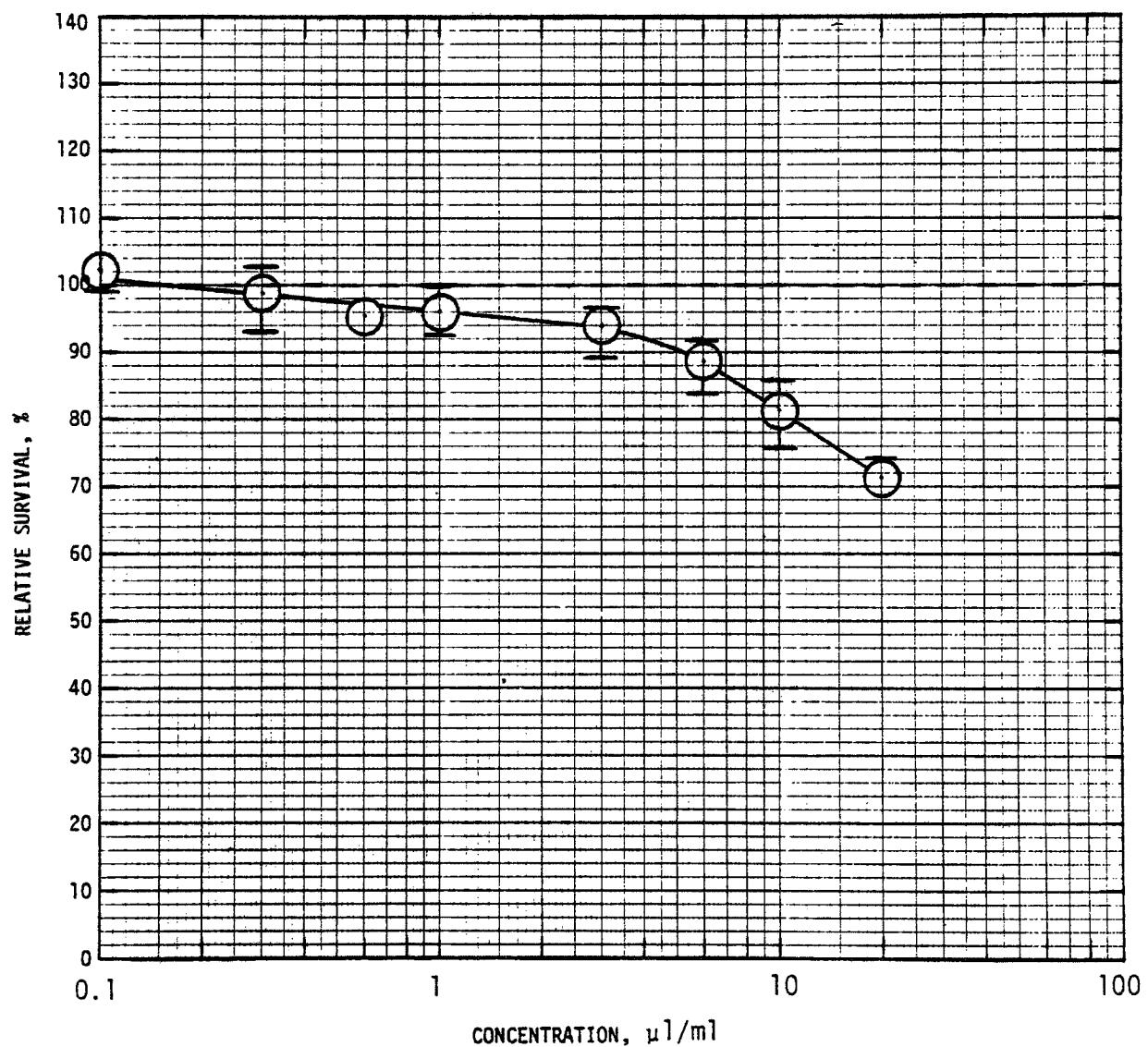


FIGURE 4

RODENT CELL (CHO) CLONAL TOXICITY ASSAY
 EC_{50} DETERMINATION
MOHAWK 2 XAD
A81-09-007-495



VII. ASSAY ACCEPTANCE CRITERIA

The assay is considered acceptable for evaluation of the test results if the following criteria are met:

- The average cloning efficiency of the CHO-K1 cells in the negative controls is 70 percent or greater, but not exceeding 115 percent.
- The distribution of colonies in the treated cultures is generally uniform over the surface of the culture dish.
- The data points for each test concentration critical to the location of the EC₅₀ are the averages of at least two treated cultures.
- A sufficient number of test concentrations are available to clearly locate the EC₅₀ within a toxicity region as defined under Assay Evaluation Criteria.
- If the EC₅₀ value is greater than 1000 µg/ml, 600 µliters of aqueous sample/ml, or 20 µliters of nonaqueous sample/ml, the plotted curve does not exceed 110 percent of the negative control.

VIII. ASSAY EVALUATION CRITERIA

The EC₅₀ value represents the concentration of test material that reduces the colony-forming ability of CHO cells to 50 percent of the vehicle or negative control value. EC₅₀ values are determined graphically by fitting a curve by eye through relative survival data plotted as a function of the logarithm of the applied concentration. Each data point normally represents the average of three culture dishes. In order to indicate the variability of the data, the high and low colony counts for each concentration are used to calculate the relative survivals, and the range is shown by a bar at the position of the plotted average. If no bar is shown, the variability was within the size of the symbol. Statistical analysis is unnecessary in most cases for evaluation.

The toxicity of the test material is evaluated as high, moderate, low, or nondetectable according to the range of EC₅₀ values defined in the following table:

CHO ASSAY EVALUATION CRITERIA

Toxicity ^a	Solids (EC ₅₀ in µg/ml)	Aqueous Liquids (EC ₅₀ in µl/ml)	Nonaqueous Liquids ^b (EC ₅₀ in µl/ml)
High	<10	<6	<0.2
Moderate	10 to 100	6 to 60	0.2-2
Low	100 to 1000	60 to 600	2-20
Not Detectable	>1000	>600	>20

^aEvaluation criteria formulated by Litton Bionetics, Inc. for IERL-RTP Procedures Manual: Level 1 Environmental Assessment Biological Tests^T.

^bCriteria for nonaqueous liquids are tentative and under evaluation. If the organic or solids content is known, the sample is evaluated under the solids criteria.



BIONETICS

Another evaluation scheme is proposed for extracts obtained from SASS train gas volumes. The proportion of the total gas volume corresponding to the volume of extract used in the bioassay is calculated and expressed as liters of gas per milliliter of culture medium (L/ml) or as dry standard cubic feet of gas per milliliter of culture medium (DSCF/ml). A criterion of 1000 L/ml is set as the limit for nondetectable toxicity. This gas volume corresponds to the average volume breathed by humans over a 2-hour period. The subsequent toxicity ranges are defined by 10-fold dilution steps to conform to standard procedure. The toxicity ranges are defined in the following table for liter and dry standard cubic feet units:

Toxicity	EC ₅₀ In Liters/ml (L/ml)	EC ₅₀ In Dry Standard Cubic Feet/ml (DSCF/ml)
High	<10	<0.35
Moderate	10-100	0.35-3.5
Low	100-1000	3.5-35
Nondetectable	>1000	>35



IX.

REFERENCES

1. Brusick, D.J. and Young, R.R.: IERL-RTP Procedures Manual: Level 1 Environmental Assessment Biological Tests. EPA-600/8-81-024, Litton Bionetics, Inc., Kensington, MD, October 1981, 150 pp.
2. Brusick, D.J.: Level 1 Bioassay Assessment and Data Formatting. EPA-600/7-80-079, Litton Bionetics, Inc., Kensington, MD, April 1980, 100 pp.
3. Brusick, D.J. and Young, R.R.: Level 1 Bioassay Sensitivity. EPA-600/7-81-135, Litton Bionetics, Inc., Kensington, MD, August 1981, 52 pp.
4. DHEW Food and Drug Administration "Nonclinical Laboratory Studies; Good Laboratory Practice Regulations" Federal Register, volume 43, No. 247, pp. 59986-60020, Part II, December 22, 1978.
5. Proposed Health Effects Test Standard for Toxic Substances Control Act Test Rules; Good Laboratory Practice Standards for Health Effects. Federal Register, Part II, volume 44, No. 91, May 1979 and Part IV, volume 44, No. 145, July, 1979.
6. Guidelines for Registering Pesticides in the United States: Proposed Good Laboratory Practice Guidelines for Toxicology Testing. Federal Register, volume 45, No. 77, April, 1980.



BIONETICS

TECHNICAL REPORT DATA
(Please read Instructions on the reverse before completing)

1. REPORT NO. EPA-600/7-84-074b	2.	3. RECIPIENT'S ACCESSION NO.
4. TITLE AND SUBTITLE Environmental Assessment of a Crude-Oil Heater Using Staged Air Lances for NO _x Reduction; Volume II. Data Supplement		5. REPORT DATE July 1984
7. AUTHOR(S) R. DeRosier and B. DaRos		6. PERFORMING ORGANIZATION CODE
9. PERFORMING ORGANIZATION NAME AND ADDRESS Acurex Corporation P.O. Box 7555 Mountain View, California 94039		8. PERFORMING ORGANIZATION REPORT NO. TR-82-94/EE
12. SPONSORING AGENCY NAME AND ADDRESS EPA, Office of Research and Development Industrial Environmental Research Laboratory Research Triangle Park, NC 27711		10. PROGRAM-ELEMENT NO.
		11. CONTRACT/GRANT NO. 68-02-3188
15. SUPPLEMENTARY NOTES IERL-RTP project officer is Robert E. Hall, Mail Drop 65; 919/541-2477. Volume I consists of Technical Results.		13. TYPE OF REPORT AND PERIOD COVERED Final; 6/81 - 11/83
16. ABSTRACT This volume of the report is a compendium of detailed emission and test data from field tests of a crude-oil process heater and laboratory analyses of collected samples. The process heater, burning a combination of oil and refinery gas, was tested in two operating modes: baseline (normal operation) and with staged combustion air for NO _x reduction. The test data include preliminary equipment calibration, detailed heater operation, and complete flue gas emission measurement results. Exhaust gas emission measurements included continuous monitoring for criteria gas pollutants; onsite gas chromatography (GC) for volatile hydrocarbons (C sub 1 to C sub 6); and source assessment sampling system (SASS) for total organics in two boiling point ranges (100-300 °C and >300 °C). Organic compound category information was obtained using infrared spectrometry (IR) with specific quantitation of the semi-volatile organic priority pollutants using gas chromatography with mass spectrometry (GC/MC). Fractions were determined using liquid chromatography separation of organic extracts with total organic and IR and low resolution mass spectrometry (LRMS). Trace elements were determined by spark source mass spectrometry (SSMS) and atomic absorption spectrometry (AAS). Biological assays of organic sample extracts were also performed.		14. SPONSORING AGENCY CODE EPA/600/13
17. KEY WORDS AND DOCUMENT ANALYSIS		
a. DESCRIPTORS Pollution Nitrogen Oxides Crude Oil Gases Assessments Flue Gases Lances	b. IDENTIFIERS/OPEN ENDED TERMS Pollution Control Stationary Sources Refinery Gas Environmental Assessment Staged Combustion Air Lances	c. COSATI Field/Group 13B 07B 11H, 08G 07D 14B 21B 13I
18. DISTRIBUTION STATEMENT Release to Public		19. SECURITY CLASS (<i>This Report</i>) Unclassified
		20. SECURITY CLASS (<i>This page</i>) Unclassified
		21. NO. OF PAGES 260
		22. PRICE