

FINAL REPORT

ENVIRONMENTAL EVALUATION OF POLLUTANTS IN SITKA, ALASKA

**BASED ON A RECONNAISSANCE SURVEY CONDUCTED AUGUST 26-31, 1990,
BY**

**U.S. ENVIRONMENTAL PROTECTION AGENCY, REGION 10, IN COOPERATION WITH
U.S. FISH AND WILDLIFE SERVICE AND
ALASKA DEPARTMENT OF ENVIRONMENTAL CONSERVATION**

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| | | |
|------|--|----|
| I. | EXECUTIVE SUMMARY | 3 |
| II. | RECONNAISSANCE SURVEY | 5 |
| | A. METHODS | |
| | 1. Sampling | |
| | 2. Analysis | |
| | B. RESULTS AND DISCUSSION | |
| | 1. Dioxins and furans | |
| | 2. Organics | |
| | 3. Metals | |
| | 4. Conventional - pH, DO, T, TOC, grain size | |
| | 5. Other contemporaneous studies | |
| III. | SCREENING HUMAN HEALTH RISK ASSESSMENT FOR EXPOSURE TO DIOXINS AND FURANS | 22 |
| | A. Introduction | |
| | B. Toxicity of Dioxins and Furans | |
| | C. Dioxin and Furan Sampling and Analysis | |
| | D. Exposure Assessment | |
| | E. Risk Estimates | |
| IV. | BIBLIOGRAPHY | |
| V. | APPENDICES | |
| | A. Summary tables of dioxins and furans | |
| | B. Risk calculations | |
| | C. Raw data | |
| | D. Quality assurance reports | |

EXECUTIVE SUMMARY

Sitka, Alaska is in the southeastern end of the state. Primary activities in the area are tourism, fishing and pulp and paper production. Local citizens are concerned about their exposure to toxic chemicals, specifically dioxin, which may be released as a result of pulp mill operations. In response to this concern, the U.S. Environmental Protection Agency (EPA), in cooperation with Alaska Department of Environmental Conservation (ADEC) and the U.S. Fish and Wildlife Service (USFWS), conducted a reconnaissance survey for chemical contaminants in Sitka and the surrounding areas during August 26-31, 1990.

During June, July and August 1990 the State of Alaska Department of Environmental Conservation (ADEC) completed an analysis of dioxins and furans in ash and sludge samples near the pulp mill. Their data are also included in this evaluation.

The results of the reconnaissance survey for chemical pollutants in the Sitka area are described in two parts. The first part (Section II) includes the results of the reconnaissance survey for approximately 250 inorganic and organic chemicals as well as a detailed discussion of sampling and analytical methods for dioxins and furans. Measurements of dioxins and furans in environmental media (ash, sludges and sediments) for which there is no direct human exposure are also described in the first part. A preliminary risk assessment for exposure to dioxins and furans by people living in Sitka is presented in the second part (Section III).

Very low concentrations of dioxins and furans were found in the Sitka area. At these low concentrations, using worst case exposure assumptions, the likelihood of health effects occurring is also very low. The analysis presented in this report is not intended to be considered a formal risk assessment. The results may be used to determine if additional sampling and analysis are warranted.

The conclusions reached in this report are based on limited sampling of the following environmental media for dioxins and furans:

- 1) ash from the APC mill,
- 2) floating residue in Silver Bay,
- 3) leachate and sludge from the city landfill,
- 4) sediment from Silver Bay and Blue Lake,
- 5) soil from Sitka and surrounding areas,
- 6) water from Blue Lake, and
- 7) seafood from Silver Bay, Thimbleberry Lake and Blue Lake.

Trace levels (0 to 22 parts per trillion) of dioxins and furans were found in floating residue, soils and sediments. These levels appear to be similar to background concentrations reported for the U.S. and other countries throughout the world. No dioxins or furans were found in leachate from the landfill and water from Blue Lake. The highest levels were found in ash from the pulp mill power boilers. High concentrations of dioxins and furans in ash

from incinerators have been previously documented in other municipal and industrial settings.

Seafood from Silver Bay also contained trace levels of dioxins and furans. The range in concentration from (0 to 3 parts per trillion) is similar to other background levels measured in the U.S.

The results of the reconnaissance survey for other potential pollutants in the Sitka area are summarized below:

The degradation of water quality, demonstrated in previous studies, continues. Dissolved oxygen at depth was below levels considered adequate to support marine life. pH was below normal levels in surface waters. Organic material on the seafloor was built up considerably over pre-discharge levels. This blanket of solids depletes oxygen in sediments and bottom water, greatly affecting benthic organisms. Suspended solids and color in the upper water column can reduce photosynthesis adding further stress to the marine ecosystem.

Sediments, water and soils were sampled for the presence of more than 250 inorganic and organic chemicals. Several contaminants found in the sediments and water of Silver Bay may be a threat to the health of the aquatic organisms. Based on a comparison with values used to assess marine sediments in Puget Sound, Washington, 4-methylphenol, benzoic acid, and phenol may affect the aquatic organisms which live in the sediments. Copper and zinc in Silver Bay exceeded EPA's ambient water quality criteria for marine life.

II. RECONNAISSANCE SURVEY

A. Methods

1. Sampling

The sampling objectives were to: (1) evaluate the potential accumulation of dioxins and furans and metals in air emissions from the Alaska Pulp Corporation pulp mill (APC) into surface soils, Thimbleberry Lake fish, and Blue Lake water, fish, and sediment; (2) evaluate the presence of dioxins and furans, metals, and organic contaminants in the marine environment near the APC effluent discharge (for this study, marine sediments and biota were examined for dioxins and furans; but water column samples were not); and (3) evaluate the presence of dioxins and furans and metals in the City landfill and potential release of these contaminants from the landfill by sampling the ash at APC and sampling leachate and sludge at the City landfill. Table 1 summarizes details about the samples collected and lists the general types of contaminants that were evaluated.

Field Sampling

Marine sampling and freshwater sampling were conducted with the assistance of the USFWS. The vessel Curlew was used to occupy sampling stations and conduct the trawl as described below. A skiff was used to deploy and retrieve gill nets, set lines, and crab pots. An inflatable boat was used to collect samples at Blue Lake.

Marine Sampling

Water: Locations were chosen to (1) reflect increased distance from APC, (2) be within close proximity (less than 2.5 miles) to APC, (3) sample in Silver Bay as well as Eastern Channel, and (4) sample near or in Sawmill Cove. Based on vessel operation considerations, stations were occupied where water depth was approximately 60 meters. Four stations were occupied, one in Thimbleberry Bay (Station 1, off of Eastern Channel) and three in Silver Bay (Station 2, in the western arm of Silver Bay, between Sawmill Cove and Eastern Channel; Station 3, at the entrance to Sawmill Cove; and Station 4, near the southern point of the entrance to Herring Cove). The four stations are named for nearby water bodies; their locations are shown in Figure 1. Approximate station locations were plotted using distances and relative compass bearings indicated on the Curlew's radar. Sampling dates and times are in Appendix C-4, Table 1.

A Hydrolab unit was used to measure depth (m), temperature (°C), pH, and dissolved oxygen (mg/L-uncorrected). The unit was calibrated in the morning and evening each day it was used. Because this particular unit was not configured to measure conductivity in seawater, salinity was estimated based on surface, mid-depth, and bottom grab samples analyzed by use of a refractometer at the EPA Region 10 lab.

Table 1. Summary of information on sample collection and analysis. Matrix sampled, location of sample, collection method, contaminants measured, and laboratory where analyzed.

| <u>Matrix</u> | <u>Location</u> | <u>Sampling Method</u> | <u>Measurement</u> | <u>Analysis</u> |
|-----------------------------------|--|---|---|---|
| <u>SEDIMENT AND SLUDGE</u> | | | | |
| Marine Sediment | Silver Bay: 3 stations Eastern Channel (#2) Sawmill Cove (#3) Herring Cove (#4) | Van Veen grab composite of 4 grabs | Dioxins & furans Sediment particle size Total organic carbon Metals & organics | EPA Duluth Contract lab Contract lab EPA Region 10 |
| Lake Sediment | Blue Lake: 1 station | Petite Ponar grab | Dioxins & furans Sediment particle size Total organic carbon Metals & organics | EPA Duluth Contract lab Contract lab EPA Region 10 |
| Landfill Sludge | City Landfill: leachate collection sump | Grab, using stainless steel spoon | Dioxins & furans Sediment particle size Total organic carbon Metals & organics | EPA Duluth Contract lab Contract lab EPA Region 10 |
| <u>WATER AND LEACHATE</u> | | | | |
| Marine Water | Silver Bay: 3 stations Eastern Channel (#2) Sawmill Cove (#3) Herring Cove (#4) Thimbleberry Bay: 1 Sta. | Composite of top, mid, & bottom of water col. Brass Kemmerer for organics, plastic for metals In situ measurements every 10 m water depth | Metals & organics Salinity (separate anal. for top, mid, & bot.) T, DO, pH | EPA Region 10 EPA Region 10 Hydrolab |
| Lake Water | Water Treatment Plant | Grab at pressure relief valve | Dioxins & furans Metals & organics | EPA Region 7 EPA Region 10 |
| Leachate | City Landfill: leachate collection sump Herring Cove: hogfuel storage area | Grab, using large collection bottle Grab | Dioxins & furans Metals & organics Dioxins & furans Metals & organics | EPA Duluth EPA Region 10 EPA Duluth EPA Region 10 |
| <u>SOIL AND ASH</u> | | | | |
| Soil | 10 stations: Sitka City: N, central, S, Japonski Galankin I, Mill (APC) Jamestown Bay Thimbleberry Bay Blue Lake-launch ramp Deep Inlet (background) | Composite of 4 grabs (5 at Deep Inlet) Collected using stainless steel spoons | Metals Dioxins & furans | EPA Region 10 Contract lab (7) EPA Region 7 (5) (both labs analyzed samples from the Mill & Galankin I.) |
| Power Boiler | 2 stations: Bottom ash Multiclone ash | Grabs, collected with spatulas (bottom ash) or jar (multiclone) | Dioxins & furans Metals | EPA Duluth EPA Region 10 (bottom ash only) |
| <u>TISSUE</u> | | | | |
| Marine | Herring Cove | 4 set lines Quillback rockfish (2) | Dioxins & furans | EPA Duluth |
| | Thimbleberry Bay - E.end | 2 set lines Pacific sanddab (10) Quillback rockfish (1) | Dioxins & furans Dioxins & furans | EPA Duluth EPA Duluth |
| | - Entrance to Silver Bay | Grab-hand collection Mussels (jar) | Dioxins & furans | EPA Duluth |
| | Sawmill Cove - SW side | Gill net & 3 set lines Quillback rockfish (3) Pac. stag. sculpin (5) Crab pots Dungeness crab (2) | Dioxins & furans Dioxins & furans Dioxins & furans | EPA Duluth EPA Duluth EPA Duluth |
| | Silver Bay - main channel | Trawl Shrimp (150) English sole (5) | Dioxins & furans Dioxins & furans | EPA Duluth EPA Duluth |
| Lake | Blue Lake - near boat launch | Minnow traps Rainbow trout (3) | Dioxins & furans | EPA Duluth |
| | Thimbleberry Lake - near trail end | Minnow traps E. brook trout (4) | Dioxins & furans | EPA Duluth |

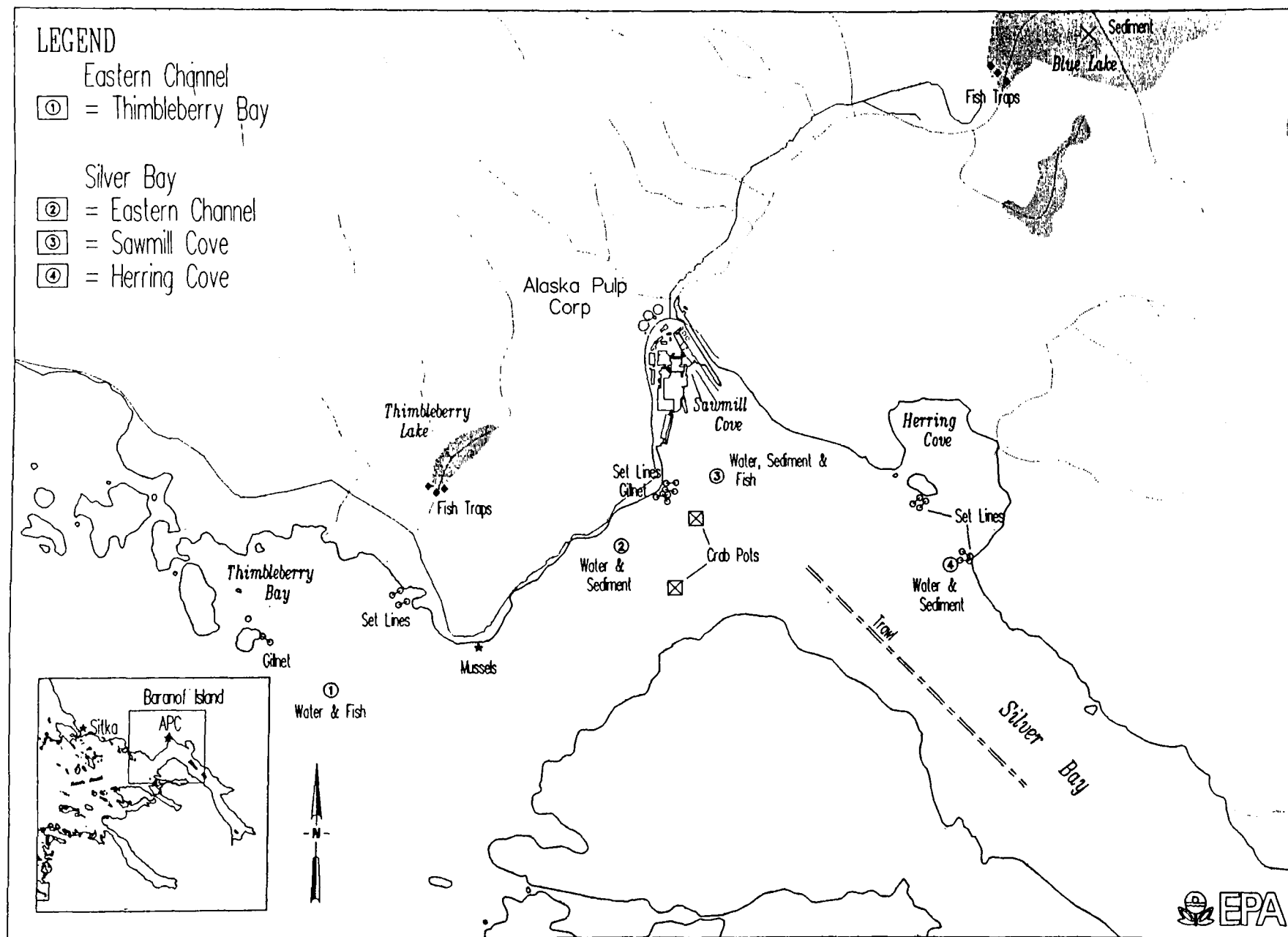


Figure 1. Location of marine sampling stations (for water, sediment, and biota) and lake sampling stations (for sediment and fish). Marine stations 2-4 are in Silver Bay and are named after the water body nearest the station.

At each station, surface, mid-depth, and bottom grab samples were composited. A brass Kemmerer sampler and a PVC Kemmerer sampler were used to collect samples for organics and metals analyses (respectively). Collection containers were filled one third from each grab, except for samples destined for volatile organics analysis (VOA). For VOAs, separate containers were filled from each grab. These were later combined at the EPA Region 10 lab to produce a composite sample prior to analysis. A few drops of acid were added to the VOA samples. Water samples were provided to USFWS for their use in conducting bioassays. Each Kemmerer sampler was rinsed with methylene chloride and then distilled water between uses.

Sediment: A stainless-steel, 0.1-m² Van Veen sampler was used to collect marine sediments. A maximum of 10 attempts was made at each of the stations to obtain 4 acceptable grabs. Despite problems such as failure of the sampler to trip, washout due to wood, rocks, or trash in the jaws, canted sample, shallow penetration, etc., grabs were collected successfully at 3 of the stations (at Thimbleberry Bay, no successful samples were obtained within 10 attempts due to the rocky substrate). At these three stations a single sediment sample was obtained by compositing subsamples from separate grabs. The sediment VOA samples were collected from the first successful grab except for the Eastern Channel Sample in the western arm of Silver Bay, when a composite was used. All traces of sediment were rinsed from the Van Veen sampler using ambient seawater prior to each successive grab attempt. At each site, a new, cleaned (and previously foil-wrapped) stainless steel spoon was used to transfer and mix sediments.

Tissue: Set lines, gill nets, crab pots, pole and line, a bottom trawl, and collection by hand in the intertidal were used to obtain organisms for tissue analysis. The sampling sites (Figure 1) were located within the vicinity of APC and the water and sediment sampling stations. Organisms were identified, measured, weighed, and evaluated for external evidence of problems such as lesions or tumors. Next, in accordance with the protocols established for the National Bioaccumulation Study (USEPA 1989), they were wrapped in foil and frozen. Mussels were collected from the intertidal into a glass jar and then frozen.

A variety of marine organisms was caught. Shrimp (mixed), mussels and crabs were the shellfish selected for analysis. Crab hepatopancreas and the remainder of the crab body were analyzed separately. In consultation with NOAA personnel familiar with the effects of sediment contamination on fish as well as fish life histories (Long 1990), Pacific sanddab, English sole, and Pacific staghorn sculpin were selected for analysis. Quillback rockfish were collected from 3 of the 4 marine sampling stations; these were analyzed to investigate the variability in tissue contamination across stations.

Freshwater Sampling

Water: A water sample was obtained August 30, 1990, at the Water Treatment Plant northwest of APC. Water from Blue Lake is piped down to the plant. Sample jars were filled directly from the pressure relief valve.

Sediment: A sample of sediment at Blue Lake was collected on August 30, 1990, by compositing grab samples from two locations (approximately 150 ft water depth on the west side of the lake and 50 ft water depth on the east side), using a Petite Ponar grab sampler, deployed 100 ft from shore (Figure 1).

Tissue: Minnow traps were used to collect fish from Thimbleberry Lake and Blue Lake (Figure 1). Fish were identified, measured, weighed, and evaluated for external evidence of problems such as lesions or tumors. In accordance with the protocols established for the National Bioaccumulation Study (USEPA 1989), fish were wrapped in foil and frozen. Rainbow trout from Blue Lake and Eastern brook trout from Thimbleberry Lake were submitted for analysis.

Soil Sampling

Preliminary sampling locations that were identified included day care facilities, schools, playgrounds, residences, and areas potentially in the path of air emissions from APC, recognizing the possible confounding influence of the Sitka City incinerator. Final locations were chosen based on a survey, August 29, 1990, of Sitka, Japonski Island, and areas in vicinity of the road leading from town to APC. A reference location, assumed to be away from the influence of APC or City emissions was selected at the head of Deep Inlet. As shown in Appendix C-6 and Figures 2 and 3, at each of 10 locations, composite samples of four aliquots (5 at Deep Inlet) were collected for analysis. A field replicate was collected at APC. At each location, a new, cleaned (and previously wrapped) stainless steel spoon was used to transfer soils from the top 0.5 to 1 inch into collection jars. The field replicate was collected by spooning soil into two sets of jars.

Landfill Sampling

At the landfill (Figure 2), on August 31, 1990, samples of leachate and sludge were collected from the catch basin. A field transfer blank sample (transferring distilled water to containers while on site) was also collected.

The leachate water was collected by dipping a glass 1 gal jar into the catch basin then filling the sample jars from this grab. The "sludge" that had collected in the bottom of the basin was scooped out with a stainless steel spoon. Each scoop was apportioned among the sample jars. No acid was added to the sediment VOAs.

APC Ash Sampling

Ash samples were collected in the power house (Figure 3). Bottom ash was collected from the power boilers. The power boilers produce ash which remains at the bottom of the boiler on the first floor of the power house. This ash is raked into dumpsters. A composite, representative sample of power boiler bottom ash was scooped out of the hoppers using a wooden spatula. On the third floor of the power house, exhaust from the boilers passes through multiclones which connect the power boilers with the electrostatic

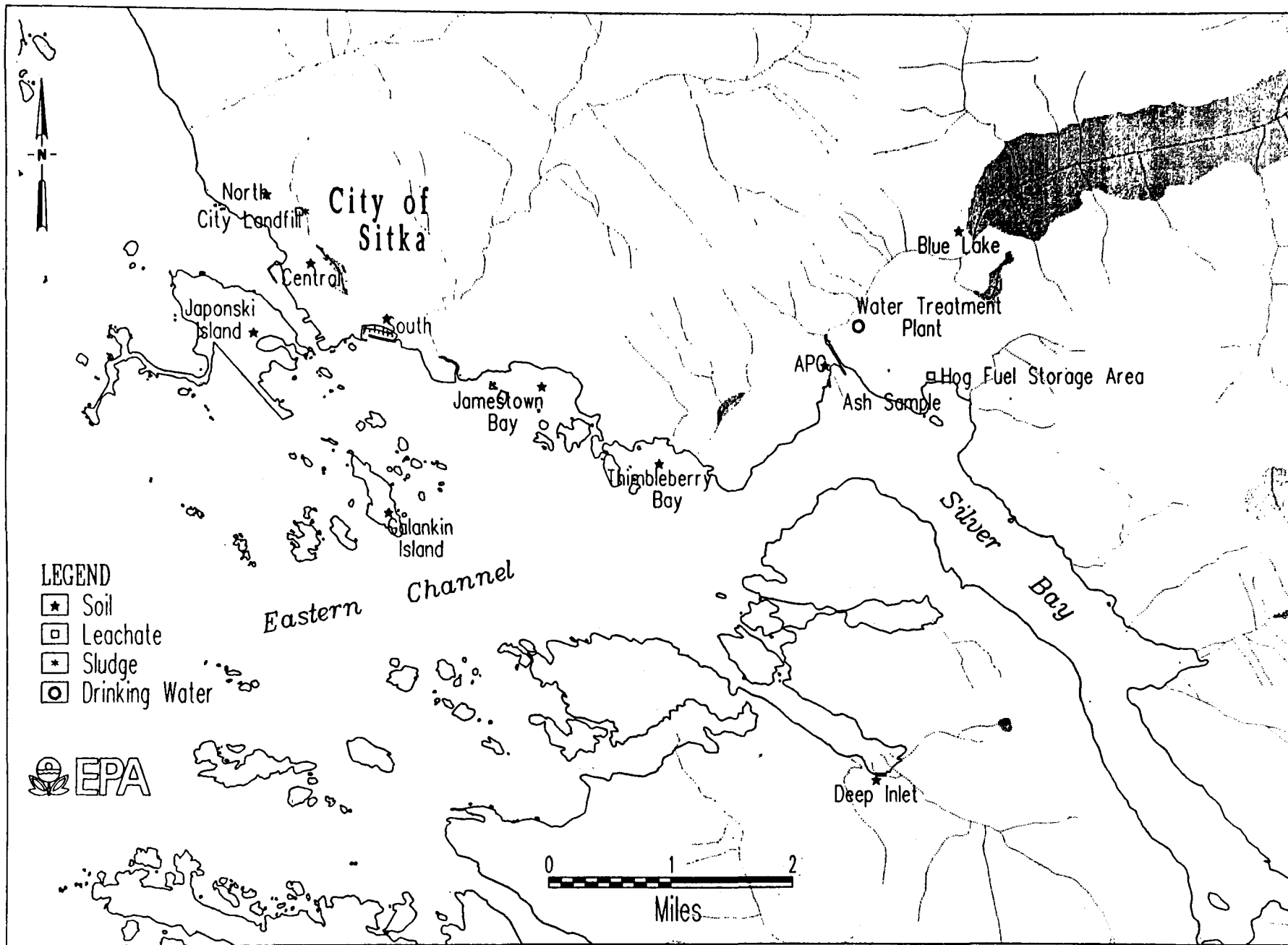


Figure 2. Location of soil, leachate, leachate-sludge, ash (from APC), and drinking water (from Blue Lake; at the water treatment plant) samples in and around Sitka, AK (August 26-31, 1990).

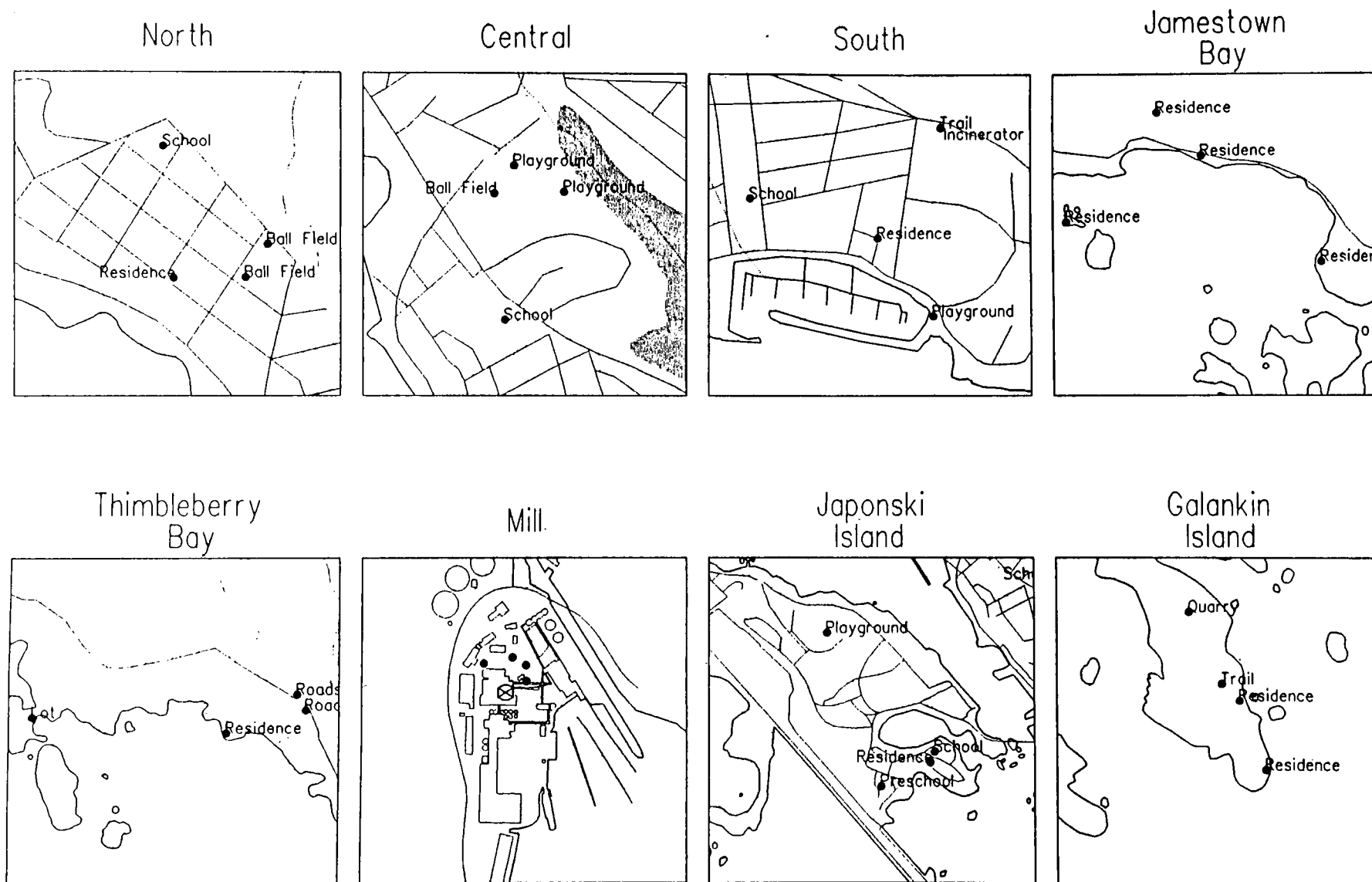


Figure 3. Location of individual subsamples used to produce the composite soil samples, for 8 of the 10 stations (Blue Lake and Deep Inlet samples are not shown). The X at the Mill indicates the location of the ash samples.

precipitator (ESP). The multiclones remove large particles of ash which drop down into a wet whirler. The remaining boiler exhaust continues to the ESP where fly ash is removed (fly ash was not sampled). A composite sample of the multiclone ash was obtained by APC personnel who held a jar through the observation ports and collected the ash as it fell down from the multiclone, before it reached the wet whirler.

Quality Assurance/Quality Control

A QA plan was developed for this sampling effort (Appendix D). In the field, two QA samples were collected: A field duplicate sample for the soil sampling conducted at APC; and a field transfer sample obtained when sampling at the City landfill catch basin.

Laboratory QA procedures involved matrix spikes, duplicates, and blanks. These were analyzed to provide information on analytical reliability and variability and to qualify data when necessary.

Other Contemporaneous Sampling

The Alaska Department of Environmental Conservation (ADEC) collected samples of floating residue from Silver Bay and Eastern Channel and fly ash samples from the mill (ENSECO 1990, 1991). The power boiler fly ash samples were collected June 21, 1990, from the location where the electrostatic precipitator conveyor screw drops the fly ash into collection bags. A 1 L grab sample was collected at the end of each of three 8 hr periods for a total of 3 samples.

Five grabs of floating residue were collected July 20, 1990 (3 samples within 100 ft of the log booms and 2 samples near the log pullout area in Sawmill Cove) and composited for a single analysis of residue in Sawmill Cove. Two additional residue samples were collected August 8, 1990, from Eastern Channel (one near the east end of Thimbleberry Bay and one from the midpoint of Eastern Channel, south of the west end of Jamestown Bay).

During the present survey, the US Fish and Wildlife Service (USFWS) obtained water samples from the four marine stations occupied in the present survey as well as floating residue from Sawmill Cove.

2. Analysis

Analyses were completed by US EPA Region 10, Region 7, and Duluth labs, as well as contract labs (Table 1). As indicated in Table 1, in addition to dioxins and furans, analyses for selected samples included metals, organics (volatiles, semivolatiles, base, neutral, acid compounds, etc.), organic carbon content, sediment particle size, and salinity. The methods used by each laboratory are listed or referenced in Appendix D.

B. Results and Discussion

Measured concentrations were compared with a variety of screening values. These screening values consisted of the EPA ambient water quality criteria (for freshwater, leachate, and marine water samples), apparent effects thresholds (for Puget Sound, WA, sediments), and New York Department of Environmental Conservation screening level (for dioxins in fish tissue).

The results of concern include: (1) Dioxins and furans were present in sediments, fish, and shellfish in Silver Bay. These contaminants appear to be at low levels, but their presence in the benthic food web is unknown; (2) Dioxins and furans were found in Blue Lake sediment in concentrations as high as in Sawmill Cove. Again, it is not known whether these contaminants are in the limnetic food web; (3) Four-methylphenol, benzoic acid, and phenol were present at high levels of concern in the sediments near the mill; (4) Copper and zinc were present at high levels of concern in the water column near the mill; (5) Copper was present at high levels in the Herring Cove hogfuel storage leachate; (6) Dissolved oxygen was depressed below levels considered to be adequate to support marine life; (7) pH was depressed in surface waters; and (8) Organic material has built up, possibly beyond levels measured in 1970.

In addition to those stresses listed above, suspended solids and color reduce photosynthesis in the upper water column and thereby impact the marine ecosystem.

1. Dioxins and Furans (Appendices A, C-1)

Marine Sediment: In general, the highest values were found in the Sawmill Cove sediment where more congeners were detected than at the other two stations. Without some screening value against which to compare the measured concentration, it can only be stressed here that these compounds are present in most of the organisms sampled in Silver Bay which is consistent with the known ability of dioxins and furans to bioaccumulate and enter the food web (see, for example, Rabert 1990).

Marine Tissue: Dioxins and furans were measured and detected in the tissues of a variety of fish and shellfish. The following discussion is based on the review of the effects of tetrachlorodibenzodioxin (TCDD) and tetrachlorodibenzofuran (TCDF) releases from pulp and paper mills on aquatic organisms by Rabert (1990). Lethality has not been shown to be related to TCDD body burdens in fish. In fact, fish mortality can occur even after significant loss of TCDD from tissues. However, fish are very sensitive to TCDD and TCDF and delayed lethality can result from exposures as short as 6 hours. In addition, fish may develop lesions following exposure. In the present study, the only evidence of visible external abnormalities found was fin damage in 4 of 6 total copper rockfish collected from Herring Cove and a growth in the lower eye of one arrowtooth flounder collected from Thimbleberry Bay. Further investigation is necessary to determine the cause of these abnormalities.

Because chlorinated dioxins and furans can bioconcentrate, piscivorous birds and mammals may be at risk. Rabert (1990) cites Newell, 1987, concerning the New York Department of Environmental Conservation's screening value. They (NYDEC) determined that wildlife are at risk when feeding on fish with body burdens greater than 3 ppb (ng/kg) TCDD. In the present study, the highest TCDD value in tissue was the analysis of English sole at 1.17 ng/kg which is below the NY screening value. The preliminary data collected during this survey suggest that wildlife may not be at risk from TCDD when feeding on fish from the Silver Bay and Eastern Channel areas, at least for those fish species which were evaluated.

Freshwater: Dioxins and furans were not detected in leachate or freshwater.

Lake Sediment: Dioxins and furans were found in the Blue Lake sediment sample. The levels of these contaminants were similar to those found in Sawmill Cove and, apart from the heptadioxins and tetrafurans, were higher than Sawmill Cove values. There is concern that dioxins and furans may be in the food web, although they were not in the rainbow trout in concentrations sufficient to be detected by the laboratory analysis.

Lake Tissue: Based on the laboratory detection limits, dioxin and furan congeners (containing chlorine at positions 2,3,7,8) were not detected in Blue Lake rainbow trout or in Thimbleberry Lake brook trout (whole body analyses). Detection limits, however, were higher than the NYDEC screening value so that risk to piscivorous wildlife could not be evaluated.

Soil, Ash, and Landfill Sludge Sampling: Because of the emphasis on human exposure (locations were playgrounds, ball fields, residences, catch basins, etc.), these sample results are not very useful in evaluating ecological effects. Dioxins and furans were detected in the ash samples, the landfill sludge sample, and many of the soil samples (see Section III). The sludge and soil levels were very low compared with the fly ash. At Deep Inlet, the background site, dioxins and furans were not detected. However, interference due to high levels of organic matter resulted in fairly high detection limits.

2. Organics (Appendix C-2)

Marine water: Analyses for organic compounds were made on composite water samples from the four marine stations. In general, only a few compounds were detected. For most of the organic compounds, especially those volatiles and semivolatiles detected, there are no water quality criteria to use as a screening tool to evaluate the potential risk to the ecosystem.

Marine Sediment: Because no contaminant-specific marine sediment standards are available for the state of Alaska, apparent effects threshold values (AETs), as described below, were used. Bioassays are often used to evaluate sediment toxicity relative to control or reference sediments. By measuring contaminant concentrations for the same sediments, it is possible to derive AETs. These values represent contaminant concentrations above which biological effects are always observed. An AET is developed for each effect; increased oyster larvae mortality, or decreased bacterial luminescence, for

example. The range of AETs can be used then to determine a High AET (HAET) and a Low AET (LAET). AETs have been established for Puget Sound, WA, and these are used to evaluate the Silver Bay sediments. Although these two waterbodies cannot be considered to have exactly the same biota, they do share many genera and species (compare information in Barnes, et al. 1956 and Eldridge, et al. 1957 with Gotshall & Laurent 1980 and Kozloff 1983). Table 2 summarizes the exceedances based upon Puget Sound, WA, AETs.

At the Sawmill Cove station, six contaminants exceeded the AETs. In particular, 4-Methylphenol was 50 times the concentration set for the HAET and 270 that set for the LAET. Benzoic acid was 16 times the HAET. Phenol was 11 times the HAET and 31 times the LAET. For three other contaminants, the LAETs but not the HAETs were exceeded (fluoranthene, fluorene, and phenanthrene). The only other station to show exceedances was Eastern Channel, for 4-Methylphenol and benzoic acid.

Table 2. Exceedances of EPA ambient water quality criteria applied to water samples, and exceedances of Puget Sound, WA, apparent effects threshold values applied to marine sediment samples (Eastern Channel refers to station 2 within Silver Bay, see Figure 1).

| Contaminant | Sample | Concentration | Ambient Water Quality Criterion | | | |
|-------------------------|---------------------------------------|---------------|--|----|-------------------|--|
| | | | Acute Criterion | | Chronic Criterion | |
| MARINE WATER | | | | | | |
| Copper | Sawmill Cove | 181 ug/L | 2.9 ug/L | | 2.9 ug/L | |
| | Herring Cove | 16.2 ug/L | | | | |
| | Herring Cove-hogfuel storage leachate | 13.5 ug/L | | | | |
| Zinc | Sawmill Cove | 267 ug/L | 95 ug/L | | 86 ug/L | |
| Mercury | Herring Cove | 0.19 ug/L | 2.1 ug/L | | 0.025 ug/L | |
| | Eastern Channel | 0.16 ug/L | | | | |
| Lead | Herring Cove | 18 ug/L | 140 ug/L | | 5.6 ug/L | |
| | Thimbleberry Bay | 12 ug/L | | | | |
| FRESHWATER | | | | | | |
| Zinc | City Landfill leachate | 81.3 ug/L | 65.04 ug/L | | 58.91 ug/L | |
| Iron | City Landfill leachate | 76,500 ug/L | -- | -- | 1000 ug/L | |
| Lead | City Landfill | 2 ug/L | 33.78 ug/L | | 1.32 ug/L | |
| MARINE SEDIMENT SAMPLES | | | Puget Sound Apparent Effects Threshold | | | |
| 4-Methylphenol | Sawmill Cove Eastern Channel | 180,000 ug/kg | High AET | | Low AET | |
| | | 4,400 ug/kg | 1,900 ug/kg | | 670 ug/kg | |
| Benzoic acid | Sawmill Cove Eastern Channel | 12,000 ug/kg | 760 ug/kg | | 650 ug/kg | |
| | | 880 ug/kg | | | | |
| Fluoranthene | Sawmill Cove | 2,300 ug/kg | 30,000 ug/kg | | 1,700 ug/kg | |
| Flourene | Sawmill Cove | 570 ug/kg | 3,600 ug/kg | | 540 ug/kg | |
| Phenanthrene | Sawmill Cove | 2,600 ug/kg | 6,900 ug/kg | | 1,500 ug/kg | |
| Phenol | Sawmill Cove | 13,000 ug/kg | 1,200 ug/kg | | 420 ug/kg | |

Freshwater: Leachate from the City landfill and water from Blue Lake were evaluated against the EPA freshwater water quality criteria. Leachate from the hogfuel storage at Herring Cove was evaluated using EPA marine water quality criteria since it discharges into Silver Bay. In general, few organics were detected. For the volatiles and semivolatiles, few EPA freshwater criteria are available.

Lake Sediment: No standards (AETs or criteria) are available at present for screening analysis of freshwater sediments.

3. Metals (Appendix C-3)

Marine Water: Analyses for metals were made on composite water samples from the four marine stations. In general, only a few metals were detected. Some metals exceeded EPA water quality criteria for aquatic organisms, with the largest exceedances at the Sawmill Cove station (Table 2).

Copper. Copper was about 60 times the EPA acute (which is the same as the chronic or continuous exposure) criterion (181 ug/L compared to the criterion of 2.9 ug/L) at the Sawmill Cove station and about 6 times the EPA acute criterion at the Herring Cove station. Copper was 2.3 ug/L at the Eastern Channel station and not detected at the Thimbleberry Bay station.

Zinc. The concentration at the Sawmill Cove station (267 ug/L) was about 3 times the EPA acute criterion of 95 ug/L. Zinc was 25 ug/L at Herring Cove and not detected at the other two stations.

Other Metals. Mercury concentrations at the Herring Cove and East Channel stations exceeded the EPA chronic (continuous) criterion but not the acute criterion. Similarly, lead concentrations at Herring Cove and Thimbleberry Bay exceeded the EPA chronic criterion (Table 2).

Marine Sediment: No metals were detected at concentrations exceeding the Puget Sound AETs.

Freshwater: Blue Lake water had no exceedances of water quality criteria. The Sitka City landfill leachate, however, exceeded the freshwater acute criterion for zinc (by 1.25 x) and the chronic criterion for iron (77 x) and lead (just exceeded; Table 2). Because the leachate commingles with other waste streams prior to treatment and eventual discharge these results indicate the probable low toxicity of the leachate rather than a potential effect on biota. The leachate from the Herring Cove hogfuel storage area exceeded the marine chronic criterion for copper (4.7 x; Table 2). The volume of discharge and dilution of the leachate within Herring Cove is unknown, but the marine copper criterion was exceeded in Silver Bay at the Herring Cove station. This suggests that effects of copper in Silver Bay should be investigated further.

Lake Sediment: No freshwater sediment criteria are available at present for screening analysis.

4. Conventional: pH, DO, T, TOC, and Grain Size (Appendix C-4)

Marine Water: Figures 4-6 show the profiles of dissolved oxygen (DO), temperature (T, in °C), and pH. Salinities were lower in the surface grabs (approximately 20 to 26 ‰, with mid and bottom grabs having similar values (approximately 32 ‰; Appendix C4-Table 1). Salinity and temperature were used to evaluate percent oxygen saturation (Figure 4). The DO, T, and pH profiles show: (1) A reduction in DO from 5 - 9.5 mg/L in the surface to 1.5 - 3.5 mg/L at 60 m with most of the reduction occurring below 40 m water depth (changes in % saturation of DO follow a similar pattern); (2) A gradual decline of T with depth; and (3) A depression of pH in the surface 20 m (about 1.8 units below the fairly constant pH of 7.4 measured from 20 to 60 m water depth). The pH depression was most noticeable at the Herring Cove station.

pH. -- Near surface pH depressions in an August 26, 1965, survey by the US Dept of Interior (USDOI 1966) was attributed to "the combined result of biochemical waste decomposition, acid nature of the pulp mill wastes, and reduced photosynthetic production in the waste layer." However, during the 1965 survey, pH depressions did not fall below 7 units. In our study, pH below 7 was measured at the surface at 3 of the four stations, and did reach a low of 5.5 units (Figure 6). One anomalously high reading was noted at 50 m water depth for the Thimbleberry Bay station.

Dissolved Oxygen. Several previous studies in the Silver Bay area measured DO. Two studies that preceded the operation of the mill generally found DO to be above 6 mg/L from the surface to 60 m water depth. In May and July of 1956, a University of Washington study measured DO values from 6.37 to 11.20 mg/L at water depths of 50 to 75 m regardless of location (Barnes, et al. 1956). This same study did measure DO as low as 0.5 mg/L, but these values were in water deeper than 1,300 m. The Alaska Water Pollution Control Board measured DO in July, August, September, and October, 1956 (Eldridge 1957). At 30 m, the maximum depth sampled, the lowest DO measured at any location was 6.7 mg/L. In the surface 0.5 to 1.5 m, DO was always greater than 8.5 mg/L, apart from a single measurement of 2.6 mg/L measured at 5 ft water depth (in this instance, DO was 9.4 mg/L at the 2 ft water depth and 9.6 mg/L at 15 ft).

After the mill began operating, DO was again measured on August 26, 1965 (USDOI 1966). Low DO concentrations (4.4 to 7.1 mg/L) were measured in surface water. DO then increased rapidly to a maximum value at a depth between 2 and 10 m (maximum values ranges from 7.2 to 8.0 mg/L). Below this, DO gradually decreased with depth. At 60 m water depth, DO ranged from 3.9 to 5.8 mg/L. According to the U.S. Dept of Interior (USDOI 1966), up-welled oxygen-deficient ocean water is present in bays and inlets along the north Pacific coast in late summer. This water can have DO concentrations below 5 mg/L in the absence of any detectable mill discharges. Within Silver Bay, the extreme oxygen deficit at depth is attributable to oxygen depletion due to mill discharges exacerbating the low DO found in the up-welled oceanic water (USDOI 1966). This previous study also suggested that "Any reduction of dissolved oxygen beyond that observed ... in the surface waters of Silver Bay will definitely place the values below recommended minimum levels." The

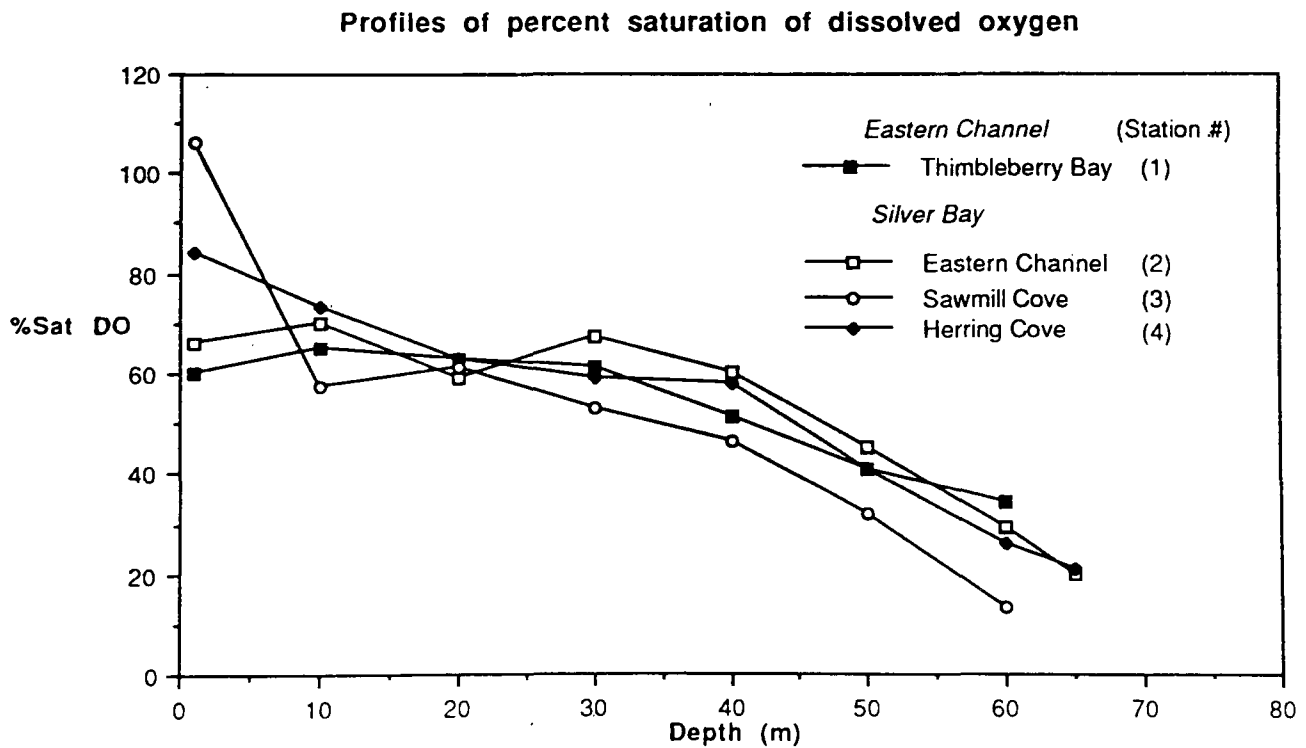
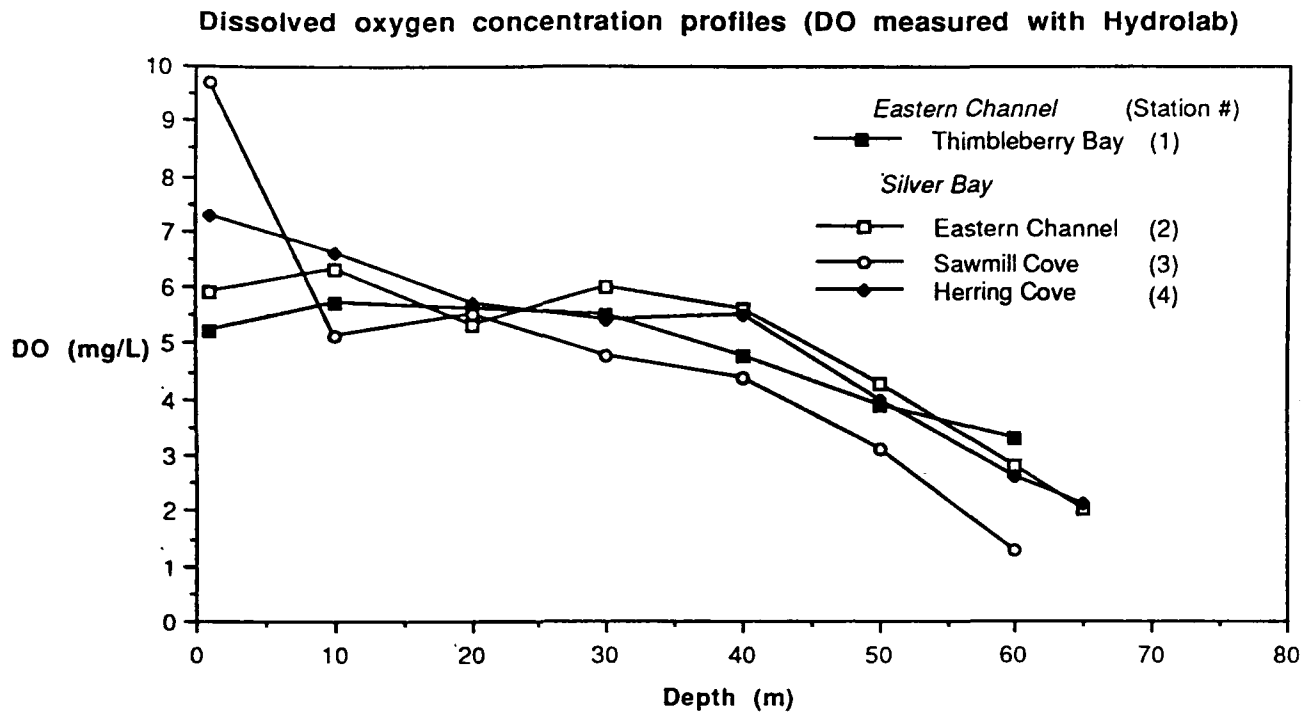


Figure 4. Dissolved oxygen concentration (mg/L) as a function of water depth (m) at the four marine sampling stations shown in Figure 1. Note the variability in the surface measurement, the relatively constant concentration between 10 m and 30 m, and the decline at all four stations below 40 m.

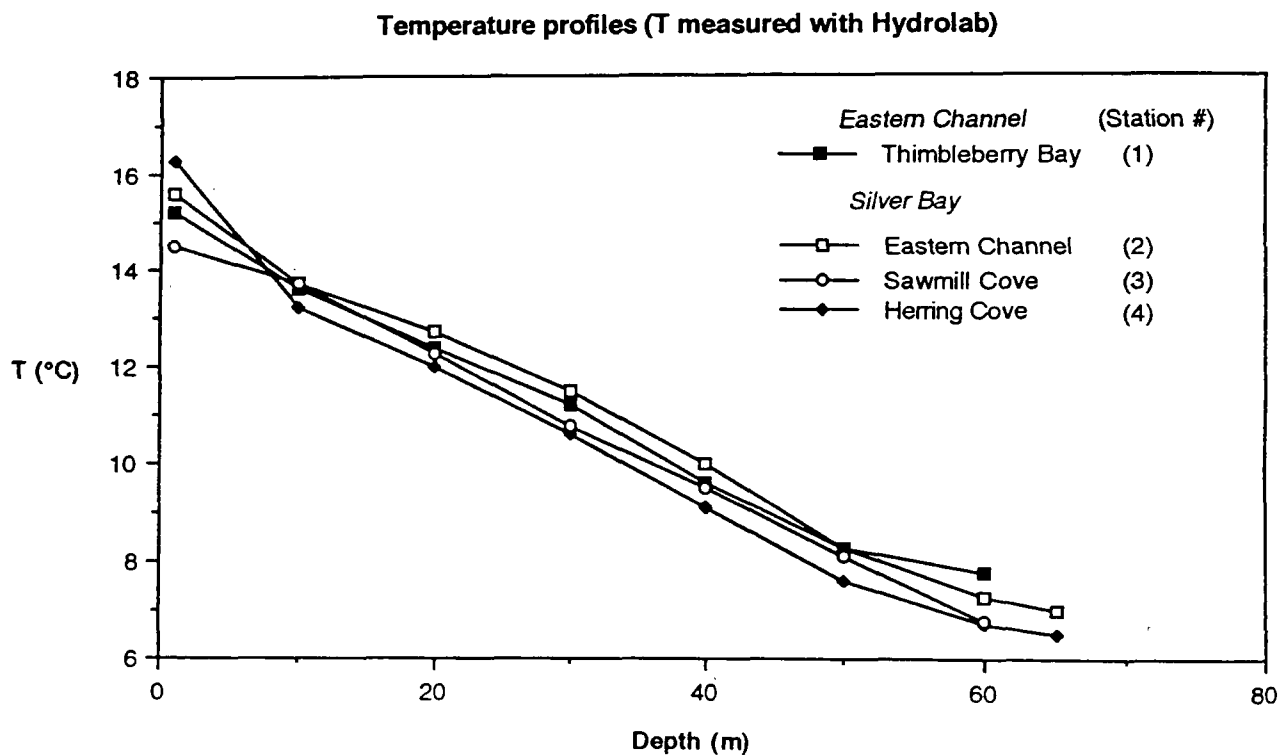


Figure 5. Water temperature ($^{\circ}\text{C}$) as a function of water depth (m) at the four marine water sampling stations shown in Figure 1. Note the relatively constant decline with depth and the similarity in profiles among the stations.

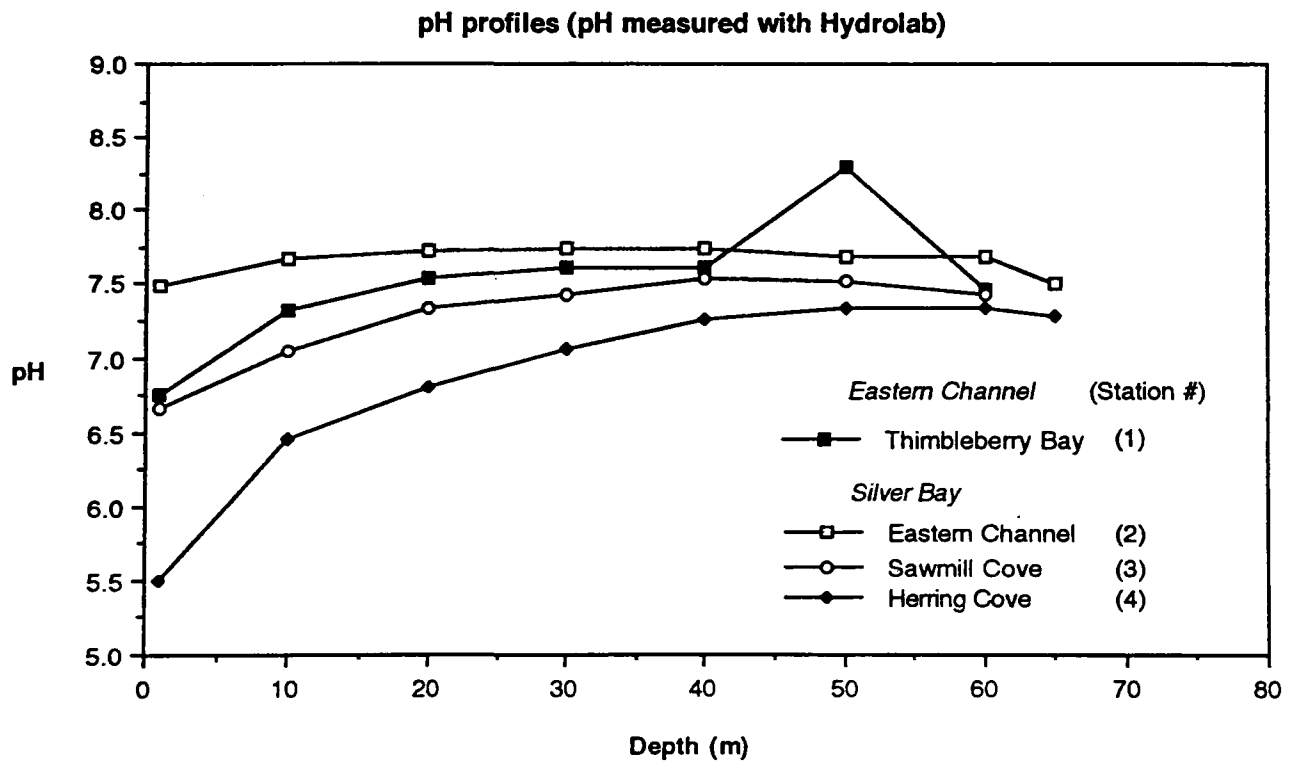


Figure 6. pH as a function of water depth (m) at the four marine water sampling stations shown in Figure 1. Note the general depression in surface waters, especially at the Herring Cove station.

present study did measure values below the 5 mg/L originally recommended in the 1956 studies as adequate to protect marine life.

Another study in 1968-70 (USEPA 1971) reported considerable reductions in DO in the surface layers of Silver Bay and Eastern Channel compared with the level measured prior to operation of the mill. The 1971 EPA report generally found DO to be above 5 mg/L (apart from some measurements in August, 1970, when DO varied between 4.6 and 6.7 mg/L at a depth of 1 m). The present study found DO in surface waters was reduced by approximately 2.5 to 11 mg/L below pre-mill measurements reported by Eldridge et al. (1957) and Barnes et al. (1956).

Marine Sediment: Total Organic Carbon (TOC) and Particle Size Analysis (Appendix C-4 - Table 2). TOC was 30% at the Sawmill Cove station, decreasing both in Herring Cove (17.5%) and Eastern Channel (8.2%). The 1956 studies measured organic carbon in Sawmill Cove at 3%, near Herring Cove at approximately 6.6%, and near our Eastern Channel station at 8.4%. In August, 1970, at three stations in Sawmill Cove, organic carbon was measured at 40%, 30%, and 27%, with increasing distance from the mill (USEPA 1971). Such a change in organic carbon in Sawmill Cove as noted in 1970 and 1990 indicates a major deposition of organic matter since the mill began discharging. The present Sawmill Cove station had an average of 28% TOC. Because it was located farther from the mill than the 1970 stations, this may indicate an increase in organic content of bottom sediments or an increase in the area affected. Comparison of the particle sizes further demonstrates the large difference in bottom type among the stations. Approximately 53% of the total weight of the sediment at Sawmill Cove was in the silt-clay fraction, while 69% was silt-clay at Herring Cove with just 11% at Eastern Channel.

Lake Sediment: The particle size and organic carbon analyses (95% silt-clay and 2.7% TOC; Appendix C-4 - Table 2) were consistent with a fine sediment lake bed.

5. Other Contemporaneous Studies

The results of dioxin and furan analysis of the ADEC ash and floating residue samples were reviewed by EPA Region 10, for quality assurance. These data are discussed in Section III (see Table 4 and Appendix A for summary data). In general, the fly ash samples had the greatest concentrations of dioxins and furans of all samples analyzed by EPA or ADEC. The floating residue samples had lower concentrations, but did contain TCDD, which was not detected in any of the EPA sediment samples. No consistent pattern was seen in relative proportions of the different dioxin and furan congeners when comparing sediment and sludge samples. How fish and wildlife come in contact with floating residue is unknown.

Effects of the residue on biota in Silver Bay and Eastern Channel should be assessed further. In addition to the dioxins and furans, the ADEC found metals in the residue. Furthermore, the results of the bioassay conducted by USFWS using marine bacteria indicated that filtrate from one of two floating residue samples was highly toxic (USDOT 1990).

III. SCREENING HUMAN HEALTH RISK ASSESSMENT FOR EXPOSURE TO DIOXINS AND FURANS

Introduction

The screening risk assessment for exposure to dioxins and furans in surface soils, floating residue, fish, shellfish and crabs is presented in this section. Section II includes the results of the reconnaissance survey for all contaminants as well as a more detailed discussion of sampling and analytical methods for dioxins and furans. Measurements of dioxins and furans in environmental media (ash, sludges and sediments) for which there is no direct human exposure are also described in Section II.

The risk analysis presented in this report is not intended to be considered as a formal risk assessment. Extremely conservative exposure assumptions were chosen as screening tools. These do not reflect actual exposures which may occur in Sitka. The results of this analysis should be used to determine what further studies or actions could be taken to protect public health in Sitka. Before additional sampling or analysis is done, the assumptions (seafood consumption rate, etc) used in screening assessment should be revised to reflect actual exposures which may occur in Sitka.

Exposure to dioxins and furans may result in cancer, reproductive and developmental effects. The chance of developing disease is dependent upon the concentration of the chemical and the likelihood of coming into contact with material contaminated with these chemicals. Risks for individuals (children and adults) who live in Sitka for 30 years in contact with surface soils, water, floating residue and consuming fish, shellfish and crabs (for 75 years) is a concern because of potential health effects.

There are a number of uncertainties which may cause the risk estimates to be too high or too low. Some of these uncertainties are listed below.

- only dioxins and furans were measured in seafood, thus the overall risks are not addressed
- samples were collected during the summer; concentrations may vary over time.
- dioxin levels being discharged have been and continue to be reduced, thus the concentrations in the environment should be diminishing over time
- the toxicity of dioxins and furans is being re-evaluated
- a limited number of samples in presumably worst case areas were sampled; these may not be representative of the whole community
- the seafood (bottom fish, crabs and mussels) collected in Silver Bay may not be part of the regular sport fishing catch

- there are no data regarding what happens to the chemicals when seafood is cooked
- exposures are assumed to occur over a lifetime (30-75 years)
- the toxic equivalent concentrations reported here include measured values as well as a correction for chemical analytical error; 1/2 the detection limit for some of the 12 toxic dioxins and furans was assumed to be present

In order to be protective of public health, EPA uses conservative assumptions for risk assessment. Given these conservative assumptions, the risks may range from the upper bound estimates given in this report to zero.

Toxicity of Dioxins and Furans

Polychlorinated dibenzo-p-dioxins and -furans are a group of 210 synthetic compounds commonly referred to as dioxins and furans. This complex group of chemicals are subdivided into homologous groups which are identified by the degree of chlorination:

- (4 Chlorines) tetrachlorodibenzodioxin - TCDD, and
tetrachlorodibenzofuran - TCDF
- (5 Chlorines) pentachlorodibenzodioxin PCDD, and
pentachlorodibenzofuran - PCDF
- (6 Chlorines) hexachlorodibenzodioxin - HxCDD, and
hexachlorodibenzofuran - HxCDF
- (7 Chlorines) heptachlorodibenzodioxin - HpCDD, and
heptachlorodibenzofuran - HpCDF
- (8 Chlorines) octachlorodibenzodioxin - OCDD, and
octachlorodibenzofuran - OCDF

Within the groups of homologues there are individual isomers which are identified according to the location of the chlorine atom on the dioxin or furan molecule. e.g. 2,3,7,8 tetrachlorodibenzo dioxin (2,3,7,8 TCDD). The most toxic dioxin or furan compound is 2,3,7,8 tetrachlorodibenzo-p-dioxin (TCDD). In order to simplify the estimates of risks to human health, a relative ranking of 12 individual congeners (homologues with chlorine atoms at the 2,3,7,8 positions) which constitute the class dioxins and furans was completed. In this ranking system, (Table 3), TCDD is given a rank of one; the other 11 in these classes are ranked from 0.5 to 0.001 relative to TCDD.

Table 3. Dioxin and Furan toxic equivalency procedure. Dioxin and furan isomer concentrations are converted into a toxic equivalent concentration (TEC) using the appropriate toxic equivalent factor (TEF).

| | TEF | CONC ng/Kg | TEC ng/Kg | | TEF | CONC ng/Kg | TEC ng/Kg |
|--|-------|---------------|--------------|---------------------|-------|---------------|--------------|
| DIOXIN | | | | FURAN | | | |
| 2378 TCDD | 1 | 1 | 1 | 2378 TCDF | 0.100 | 1 | 0.100 |
| 2378 PeCDD | 0.5 | 1 | 0.5 | 12378 PeCDF | 0.050 | 1 | 0.050 |
| 2378 HxCDD | 0.1 | 1 | 0.1 | 23478 PeCDF | 0.500 | 1 | 0.500 |
| 2378 HpCDD | 0.01 | 1 | 0.01 | 2378 HxCDF | 0.100 | 1 | 0.100 |
| OCDD | 0.001 | 1 | 0.001 | 2378 HpCDF | 0.010 | 1 | 0.010 |
| | | | | OCDF | 0.001 | 1 | 0.001 |
| TOTAL DIOXINS | | 5 | 1.611 | TOTAL FURANS | | 6 | 0.761 |
| TOTAL TEF = 1.611 + 0.761 = 2.372 | | | | | | | |

Dioxin and furans can produce a variety of effects including cancer and reproductive effects in laboratory animals at very low doses (USEPA 1989a). Based on the results in animal studies EPA has classified 2,3,7,8 TCDD as a B2 carcinogen or probable human carcinogen. It is the most potent animal carcinogen (cancer slope factor of 1.56×10^5 per milligram of dioxins and furans per kilogram body weight per day) evaluated to date by EPA. Recent reports (Fingerhut et al. 1991) suggest that while cancer has been observed in humans exposed to relatively high doses of dioxin, dioxins and furans are less potent in humans than in animals.

In addition to causing cancer, exposures to dioxins and furans may also result in developmental and reproductive effects. Based on laboratory studies with animals a one day and lifetime health advisories have been established through the EPA Drinking Water Program (USEPA, 1988) for these non-cancer health effects. In order to be absolutely sure of protecting public health, these advisories are set well above the level which has resulted in adverse health effects in laboratory animals. Although these health advisories were primarily developed for the protection of public drinking water, the same advisories may be used for other environmental media (soils, seafood, etc). The one day health advisory is: 100 picograms per kilogram-body weight per day. For a 14 kg (31 pounds) child, a concentration of 7 picograms of dioxins or furans would be equivalent to the one day advisory. The lifetime health advisory is 1 picogram per kilogram-body weight per day.

EPA as well as other world-wide health agencies are reviewing the toxicity data for dioxins and furans. This may result in a change in the cancer potency estimates and the drinking water health advisories. However, until this evaluation is complete, risk estimates completed by EPA will continue to rely on the existing cancer potency factor and non-cancer health advisories.

Dioxin and Furan Sampling and Analysis

The sampling objectives were to evaluate the potential accumulation of dioxins and furans in surface soils, fish, shellfish and crabs to which people may be exposed. The absolute levels of dioxins and furans are presented in Appendix C-1 in this report. For the purpose of discussing risks to human health, the absolute dioxin and furan concentrations found in all media are converted to toxic equivalent concentrations.

Soil, Ash, and Landfill Sludge and Leachate. Dioxins and furans were detected in the ash samples, the landfill sludge sample, and many of the soil samples. The sludge and soil levels were very low compared with the fly ash (Tables 4 and 5). No dioxins or furans were detected in the leachate samples (Appendix A). Detailed discussion of sampling and analysis is provided in Section II. The high ash level is within the range found in other incinerator residues (Table 4). Since, people are not expected to encounter ash from Alaska Pulp Corporation incinerator operations, landfill sludge, landfill leachate or sediments during regular activities at work, at home or during recreation, risk estimates are therefore not calculated for direct contact with these media.

Table 4. Toxic equivalent concentration (TEC) of dioxins and furans measured in ash, sludge, sediment and floating residue from Sitka Alaska. The TEC is calculated with 1/2 the detection limit included with the measured values.

| APC Power Boiler | TEC (ng/Kg) | SITKA LANDFILL ¹² | TEC (ng/Kg) |
|-------------------------------------|-------------|--------------------------------------|-------------|
| Fly Ash ¹¹ | 1752 | Sludge | 6 |
| Hog Fuel Bottom Ash ¹² | 17 | Leachate | ND |
| Multi Clone Ash ¹² | 10 | | |
| NATIONAL DIOXIN STUDY ¹³ | | EPA Sludge Regulations ¹⁴ | 10 |
| Fly Ash | 5000-80 | | |

¹¹ADEC, 1990

¹²This Study - U.S. EPA 1991

¹³U.S. EPA, 1989

¹⁴U.S. EPA, 1991a; TEC for only 2,3,7,8 TCDD and 2,3,7,8 TCDF
TEC of 1/2 Detection Limit - 119 pg/L

Soils. Composites of 4 to 5 soil samples were collected on August 29, 1990 in Sitka and surrounding areas (Japonski Island, Deep Inlet, and Blue Lake, Figure 2). Sample locations were selected to represent the most likely areas where people may come into contact with soils: day care facilities, schools, playgrounds, public walkways, and private residences (Figure 3).

The toxic equivalent concentrations shown in Table 5 represent not only what was measured in the soil, but they also include an additional amount to correct for uncertainty in chemical analytical methods. The measured concentrations of toxic dioxins and furans ranged from 0.2 parts per trillion at to 2.6 parts per trillion. When the correction (1/2 detection limit) for analytical error is included trace concentrations (non-detect to 14 parts per trillion) of dioxins and furans were found in surface soils in Sitka (Table 5). At Deep Inlet, which was chosen as the local background soil site, dioxins and furans were not detected. However, interference due to high organic matter in the sample resulted in fairly high detection limits. There was no significant difference in concentrations at the various locations sampled. The levels found in Sitka and surrounding areas were similar to background levels.

Table 5. Toxic equivalent concentration (TEC) of dioxins and furans measured in soils from Sitka, Alaska. The TEC is calculated with 1/2 the detection limit included with the measured values.

| | TEC (ng/Kg) | | TEC (ng/Kg) |
|---------------|-------------|------------------|-------------|
| City North | 3 | Galankin Island* | ND |
| City Center | 3 | Blue Lake | 12 |
| City South | 5 | Jamestown | 13 |
| City Japonski | 6 | Thimbleberry | 14 |
| APC | 9 | Deep Inlet* | ND |

BACKGROUND

Urban soils¹¹ ND - 11

*ND = ALL DIOXIN ISOMERS WERE AT DETECTION LIMIT (at 1/2 DL = 12 - 22 ng/Kg)

¹¹U.S. EPA, 1987; ND = 0 - 8

Sediment. Marine and freshwater sediment levels were generally low (Table 6) ranging from 4 to 12 ng/Kg. While there were not enough samples taken to establish any significant difference between sites, Sawmill Cove sediment levels appeared to be higher than other sediments in Silver Bay or Blue Lake.

Floating residue. Floating residue (fibers, organic matter) was sampled by ADEC in July and October 1990. The levels of dioxins and furans were relatively high when compared to concentrations found in soils in Sitka or in sediments in Silver Bay (Table 6). The measured concentrations (10, 12 and 25 parts per trillion) were similar to the corrected (1/2 detection limit) values presented in Table 6. The levels of dioxins and furans were primarily due to the presence of Octachlorodibenzodioxins (Appendix A, Table A-1) and furans. Since these compounds are relatively non-toxic (0.001 x 2,3,7,8 TCDD), the toxic equivalent concentrations in the floating residue were similar to that found in the soils in Sitka.

Table 6. Dioxins and furans reported as toxic equivalent concentrations, measured in sediments, floating residue from Sitka, Alaska and other areas in the U.S. The TEC is calculated with 1/2 the detection limit included with the measured values.

| SEDIMENT ¹¹ | TEC(ng/Kg) | FLOATING RESIDUE ¹² | TEC (ng/Kg) |
|--------------------------|------------|--------------------------------|-------------|
| SITKA, ALASKA | | SITKA, ALASKA | |
| Sawmill Cove | 12 | Sawmill Cove | 26 |
| Eastern Channel | 4 | Eastern Channel | 12 |
| Herring Cove | 4 | | |
| Blue Lake | 9 | | |
| WASHINGTON ¹³ | | | |
| Industrialized area | | | |
| Grays Harbor, WA | 1 - 15 | | |
| Reference area | | | |
| Sequim Bay, WA | 4 | | |
| West Beach, WA | 2 | | |

¹¹This study - U.S. EPA, 1991

¹²ADEC - July, October 1990

¹³U.S. COE, 1990

Water. Dioxins and furans were not detected in the water from Blue Lake (Appendices A and C-1) but may be present at concentrations below the method detection limits which is 100 pg/l (parts per quadrillion) for both chemical groups.

Fish, shellfish and crabs. Fish and shellfish concentrate dioxins and furans through ingestion of contaminated microorganisms, sediments or water. Therefore, they are a good indicator of general contamination in the environment. Measurements of tissue concentrations are used to estimate risk to human health from consumption of these aquatic organisms.

The types of seafood (Table 7), such as bottom fish, were selected because they tend to concentrate chemicals which may be found in the sediments at the bottom of the bay. They should therefore be worst case examples of dioxin and furan contamination. These organisms are part of commercial catches nor are they the types of seafood which may be actually caught during sport fishing in Sitka.

Mussels, crabs, and a variety of shrimp (mixed), were caught and selected for analysis. Crab internal organs and the remainder of the crab body were analyzed separately. In consultation with NOAA personnel familiar with the effects of sediment contamination on fish as well as fish life histories (Long 1990), Pacific sanddab, English sole, and Pacific staghorn sculpin were selected for analysis. Quillback rockfish were analyzed to investigate the variability in tissue contamination across marine sampling stations. Eastern brook trout and rainbow trout were collected from Thimbleberry Lake and Blue Lake. The whole fish (muscle, skin and internal organs) from each species was analysed.

Trace levels (0.1 parts per trillion to 1.3 parts per trillion before correction -1/2 detection limit- for analytical error) of dioxins and furans were found in the tissue samples for Sitka. Since, dioxins and furans tend to concentrate in organs which are high in lipids (fat) such as the liver, the whole fish samples will be higher than what may be measured if only the meat (muscle) were sampled. There was little difference in the levels of dioxins and furans found in all the aquatic organisms sampled during this study. The levels were also very close to the limits of detection (1 part per trillion). The range from the detection limit to 3 parts per trillion is similar to the background levels in the U.S. and the lowest levels measured in urban (Puget Sound) environments (Table 7).

Table 7. Toxic equivalent concentration (TEC) of dioxins and furans measured in fish and shellfish from various bodies of water in the U.S. The TEC is calculated with 1/2 the detection limit included with the measured values.

| FISH | | CRABS, MUSSELS, SHRIMP | |
|----------------------------|-------------|--|-------------|
| | TEC (ng/Kg) | | TEC (ng/Kg) |
| 1991 Sitka | | 1991 Sitka | |
| Blue Lake Trout* | ND | Crab Muscle | 2 |
| Thimbleberry Lake Trout* | ND | Crab Hepatopancreas | 3 |
| Thimbleberry Bay Sanddab | 2 | Silver Bay Shrimp | 2 |
| Herring Cove Quillback | 2 | Thimbleberry Bay Mussels | 2 |
| Thimbleberry Bay Quillback | 2 | | |
| Sawmill Cove Quillback | 2 | 1991 Puget Sound ¹¹ | |
| Silver Bay Sculpin | 2 | Crab Muscle | 3 - 0.4 |
| Silver Bay English Sole | 2 | Crab Hepatopancreas | 62 2 |
| 1985/87 Sitka | | NATIONAL BIOACCUMULATION STUDY ¹² | |
| Flathead Sole | 2 | Bearce Lake, Maine | |
| Sitka Halibut | 1 | Chain Pickerel | 0.3 |
| Red Striped Rockfish | 2 | Androscoggin R., Maine | |
| | | Sucker | 68 |

*ND = ALL ISOMERS WERE AT THEIR DETECTION LIMIT

TEC for 1/2 Detection limit = 1 ng/Kg

¹¹U.S.EPA, 1991b

¹²U.S.EPA, 1989

Exposure Assessment

To assess the effects of chemicals on people, exposure must be demonstrated. In the case of the Sitka evaluation exposure to dioxins and furans could occur through ingestion, inhalation and skin absorption of soils and dust in the community, ingestion, inhalation and skin absorption of the floating residue in Silver Bay, and ingestion of fish, crabs and shellfish.

For this screening assessment conservative assumptions were used to estimate risk from exposure to soils and dust in Sitka and floating residues in Silver Bay. These assumptions were used:

Body weight:

Child - 10 Kg
Adult - 70 Kg

Exposure Duration for soils:

Child - 6 years
Adult - 24 years

Quantity of SOIL ingested:

Child - 200 mg per day
Adult - 100 mg per day
Absorption from SOIL - 100%

Area of SKIN contact:

Child - 3900 cm²
Adult - 5000 cm² summer
1900 cm² winter

SKIN Contact Rate:

Child and Adult - 1 mg/cm²
Amount Absorbed through SKIN:
Child and Adult - 20% 20%

Fish and shellfish consumption is based on the amount of fish, crab, mussels, and shrimp that a recreational harvester might ingest over a lifetime (75 years). A consumption rate of 54 grams per day (one quarter pound every other day) was used as a screening level to estimate fish and shellfish consumption in this report. This is an average value for recreational harvesters in the U.S. (Pao, et al., 1982). Since consumption rates may vary depending on individual preferences, risks are also presented for a range of consumption rates (Figures 7-9). In addition to the amount and frequency of fish consumed, it is also important to consider what portions of the fish may be eaten. Most people would only eat fish fillets and they would discard the internal organs. Another conservative assumption used in this assessment, is that someone catches all their fish from Silver Bay.

For crabs, consumption data are presented for two types of tissue: the hepatopancreas commonly referred to as "crab butter" and muscle tissue. For the hepatopancreas there are survey data from Puget Sound (DSHS 1986) which suggest that 14% of the individuals who harvest crabs also eat the hepatopancreas commonly referred to as "crab butter". In this study the crabs weighed about 1.5 lbs (1020.58 grams); the muscle represents 30% of the whole crab (0.45 lbs or 202 grams); and, the hepatopancreas represents 0.5 % (0.01 lbs or 5 grams). For this screening assessment risk estimates are based on an individual consuming one crab (muscle and/or hepatopancreas) per week over a

lifetime. Figures 7-9 illustrate how the risk estimates would change depending on individual consumption rates.

Risk Estimates

Risk assessment for carcinogens includes an estimate of carcinogenicity (carcinogenic slope factor) as well as an estimate of human exposure. The carcinogenic slope factor is generally based on laboratory studies of animals, although in some cases there are data from human exposures.

The risk of developing cancer is obtained from carcinogenic slope factor by multiplying the latter by the actual number of units of human exposure. For a given exposure, the higher the slope factor, the higher the risk.

Risk = Carcinogenic Slope Factor multiplied times human exposure
Dioxin Carcinogenic Slope Factor = 15600 per mg per kg-body weight per day

Human exposure may be estimated from the following equation:

Human exposure from soil, water or food consumption = tissue or soil concentration multiplied times the quantity ingested multiplied times the duration of exposure divided by the body weight multiplied by average lifetime

Human exposure for skin contact = soil or floating residue concentration multiplied times skin surface area multiplied times contact rate multiplied times the duration of exposure divided by the body weight multiplied by average lifetime

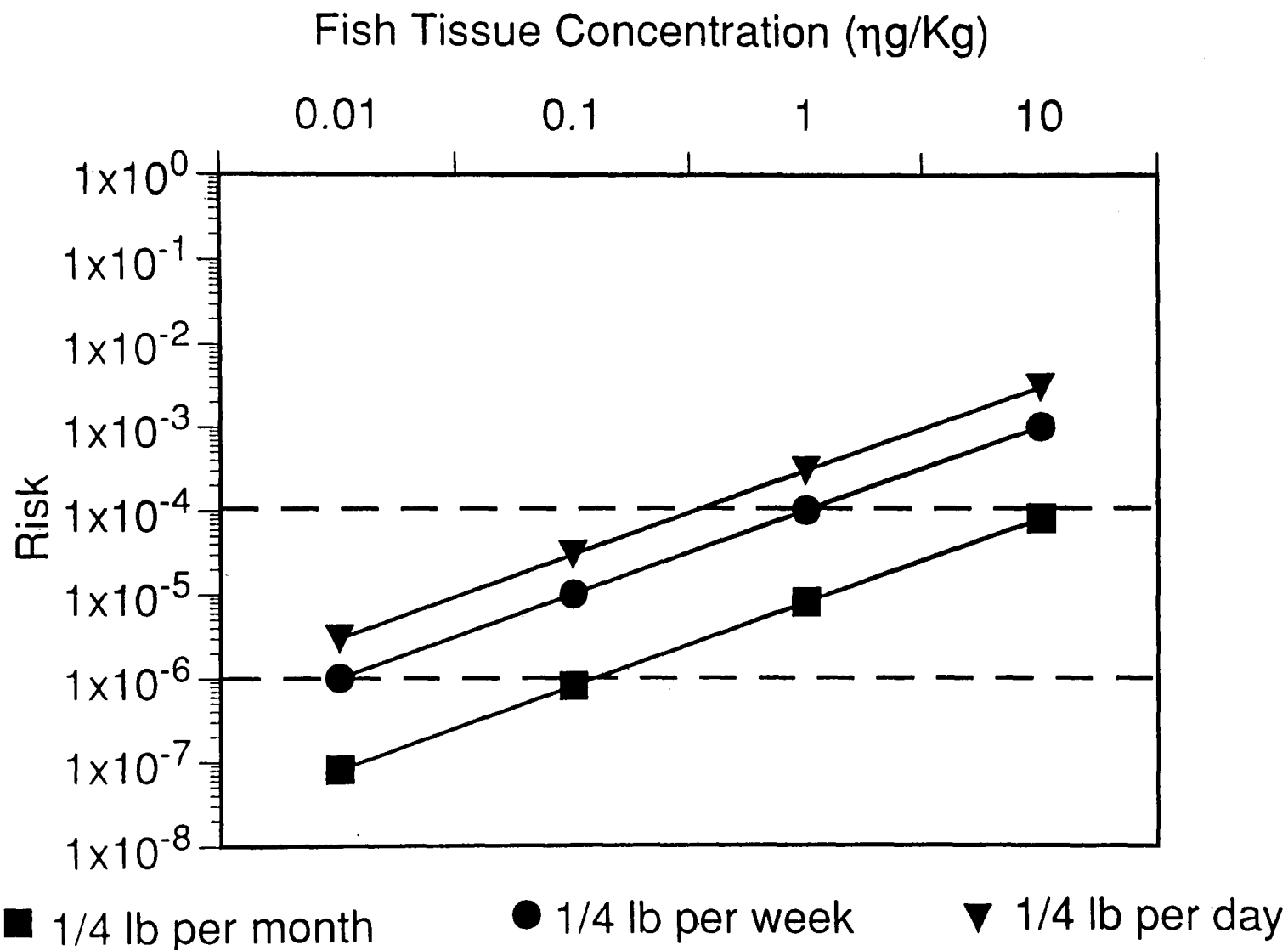


Figure 7. Relationship between dioxin and furan concentration in fish tissue and risk (probability of excess cancer) as a function of the amount of fish consumed.

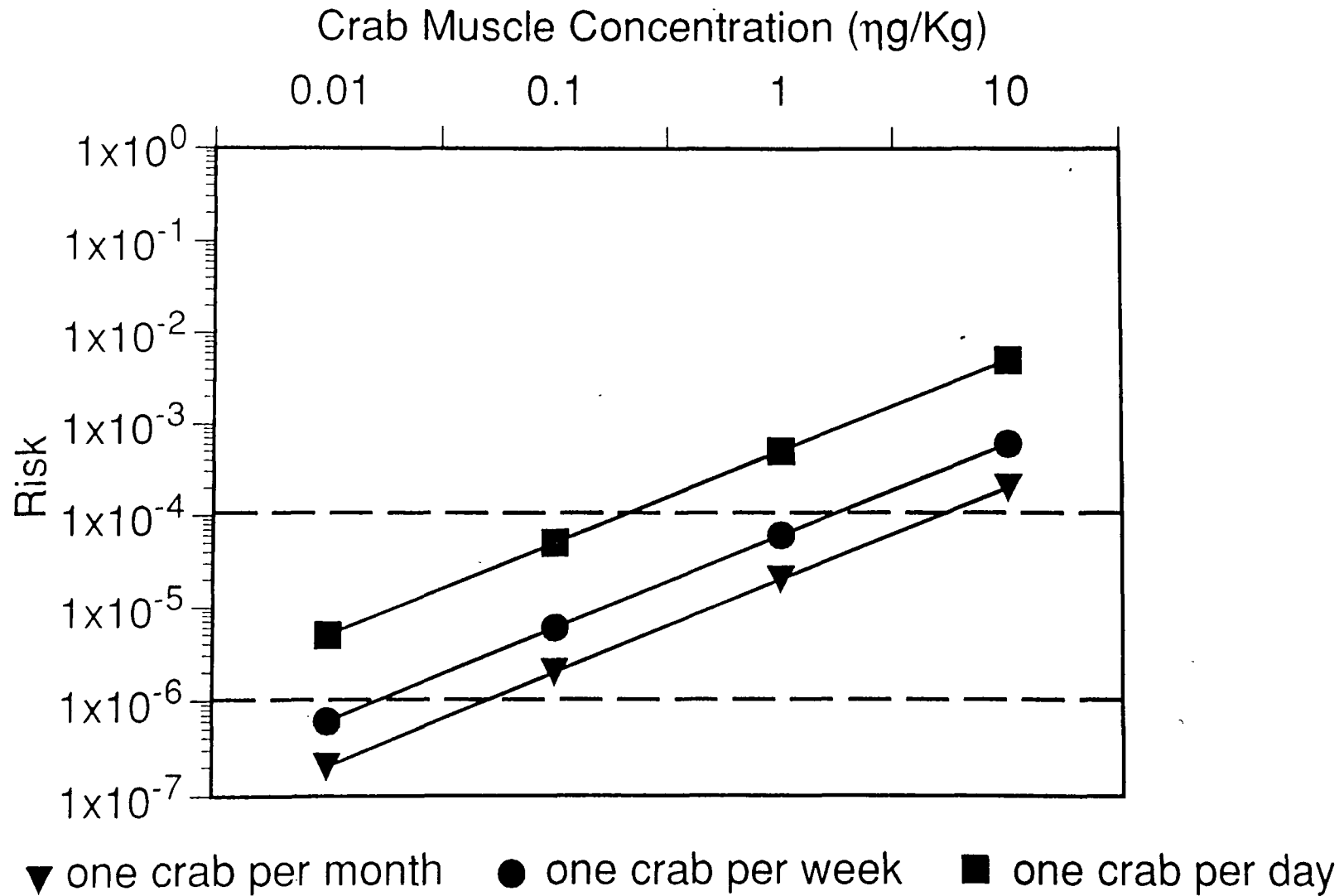


Figure 8. Relationship between dioxin and furan concentration in crab muscle and risk (probability of excess cancer) as a function of the amount of crab consumed.

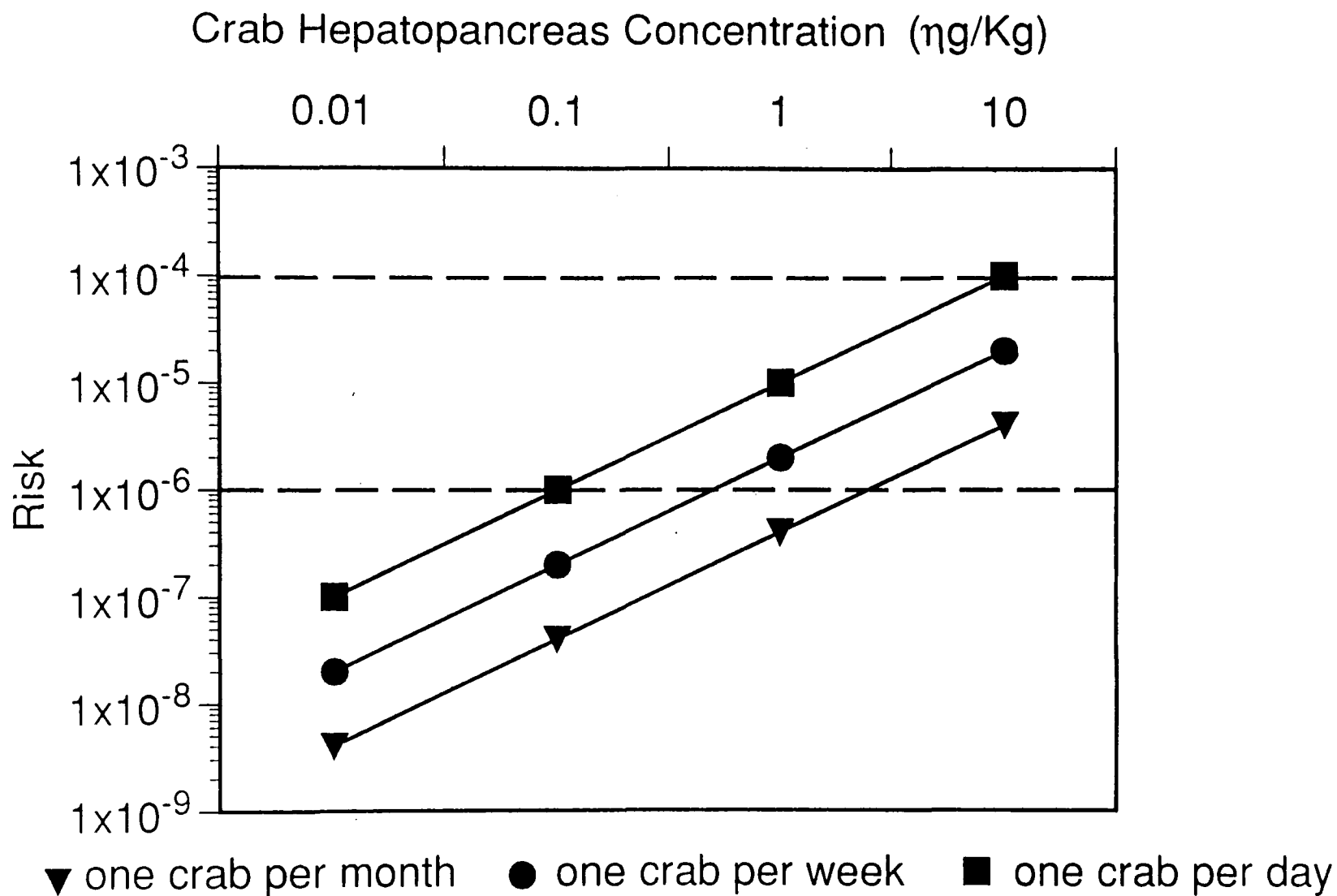


Figure 9. Relationship between dioxin and furan concentration in crab hepatopancreas and risk (probability of excess cancer) as a function of the amount of hepatopancreas consumed.

To estimate the potential for other health effects to occur the following equation is used:

$$\begin{aligned} \text{Ratio} &= \text{Human exposure divided by the one day health advisory or} \\ &\quad \text{lifetime health advisory} \\ \text{one day health advisory} &= 100 \text{ pg/kg-body weight/day} \\ \text{lifetime health advisory} &= 1 \text{ pg/kg-body weight/day} \end{aligned}$$

Soils. Risk estimates for exposure to soils assumes people ingest soil, inhale dust and absorb dioxins and furans through skin contact. The cumulative risks for developing cancer range from one in one million to twenty in one million (Table 8; Appendix B, Table B-1). There are a number of uncertainties in deriving this risk estimate for soils. It is assumed that an individual spends 30 years in direct contact with soils only from the locations presented in this study. It is also assumed that large amounts of chemical are absorbed through the skin. More information about the chemicals as well as individuals in the community would need to be understood in order to reduce some of the uncertainty. In addition to the exposure and toxicity questions, the concentrations found in the soil are also in question. Since the levels in the soil were very close to the detection limit, the risk at the detection limit (8 in one million to 20 in one million) is equivalent to the measured values. When exposures are that close to the detection limit, the accuracy of the risk estimates is not very good.

Exposures to levels of dioxins and furans measured in soils from Sitka and surrounding areas does not exceed the one day or lifetime health advisories for children and adults. Therefore it is not expected that the soils measured in this study present a health threat for non-cancer effects (Appendix B).

Floating residue. Contact with floating residues may occur through recreational activities such as swimming, playing on the beach or fishing. Exposure may be incidental ingestion of the material, inhalation or skin absorption. Since, these routes of exposure are similar to those for soil, the same assumptions as were used to derive the risk assessments for soil were used for the floating residue. The risks range from 2 in 100,000 to 4 in 100,000 (Table 8). It should be noted that these risks are based on an assumption that a child is exposed for 6 years every day to this material and an adult for 24 years, every day. The likelihood of encounters with floating residue occurring daily over a 30 year period are probably quite low. Therefore, the risks for individuals who live in or visit Sitka are more likely to be lower than this estimate.

Table 8. Carcinogenic risk from exposure (ingestion, dust inhalation and skin absorption) to dioxin and furans measured in surface soils and floating residue in Sitka, Alaska

| SOIL | | | |
|---------------|-----------------|----------------------|----------------|
| SITKA | RISK | SITKA | RISK |
| City North | 5 in 10,000,000 | Galankin Island | * |
| City Center | 4 in 10,000,000 | Galankin Island(dup) | 1 in 1,000,000 |
| City South | 1 in 1,000,000 | Jamestown | 2 in 1,000,000 |
| City Japanski | 1 in 1,000,000 | Thimbleberry | 2 in 1,000,000 |
| APC | 1 in 1,000,000 | Deep Inlet | * |

FLOATING RESIDUE
4 in 1,000,000 to 9 in 1,000,000

*NO DIOXINS OR FURANS DETECTED; 1/2 DETECTION LIMIT = 12 - 22 ng/Kg)
RISK AT 1/2 DETECTION LIMIT (1 in 1,000,000 to 2 in 1,000,000)

Fish, Shellfish and Crabs. While risk estimates may be calculated for the aquatic organisms collected during this study, there is a great deal of uncertainty in the levels of dioxins and furans which were reported for the fish tissue. The concentrations of dioxins and furans measured in seafood were at the limits of the laboratories chemical analytical capability. When tissue concentrations are close to the detection limit the accuracy of the risk estimate is questionable. Therefore, the risk estimates are clearly a maximum worst case estimate and may in fact be zero. Using the screening consumption of one 1/4 pound of seafood every other day, the data from this study indicate that the maximum risk of developing cancer could be two in ten thousand (Table 9; Appendix B). This estimate will vary depending on individual consumption rates (Figures 7-9). In estimating risks for consumption of fish it is also assumed that all seafood which an individual ingests are caught in Silver Bay. Since, the projected risks for consumption fish are at levels which may cause some health concerns, it should be understood that these risks are projected for only the worst case exposures. The assumptions used to derive these estimates need to be refined in order to better approximate the actual risk which may result from fish consumption in Sitka.

Consumption of whole fish, shellfish or crabs collected during this study does not result in a daily intake which exceeds the one day health advisory. Upper bound exposures may exceed the lifetime health advisory (Appendix B).

Table 9. Risk of developing cancer from consumption of dioxins and furans in fish, shellfish and crabs.

| FISH | | SHELLFISH | |
|----------------------------|--------------------|--------------------------|--------------------|
| | RISK ¹¹ | | RISK ¹¹ |
| Blue Lake Trout | * | Silver Bay Shrimp | 2 in 10,000 |
| Thimbleberry Lake Trout | * | Thimbleberry Bay Mussels | 2 in 10,000 |
| Thimbleberry Lake Sanddab | 2 in 10,000 | | |
| Herring Cove Quillback | 2 in 10,000 | | |
| Thimbleberry Bay Quillback | 2 in 10,000 | | |
| Sawmill Cove Quillback | 2 in 10,000 | | |
| Silver Bay Sculpin | 2 in 10,000 | | |
| Silver Bay English Sole | 2 in 10,000 | | |
| CRAB | | | |
| Sawmill Cove | RISK ¹² | | RISK |
| Hepatopancreas | 2 in 1,000,000 | | |
| Muscle | 8 in 100,000 | | |
| Whole Crab | 8 in 100,000 | | |

*DIOXINS AND FURANS WERE NOT DETECTED

RISK AT 1/2 DETECTION LIMIT = 2 in 10,000.

¹¹ Risk assumes one meal every other day for a lifetime (75 years)

¹² Risk estimate assumes one crab per week over a lifetime (75 years)

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APPENDIX A. SUMMARY TABLES OF DIOXINS AND FURANS

Table A . Dioxins and furans measured in Sitka Alaska.
DL=Detection limit. TEC = Toxic equivalent concentration.

| Descrip | SOIL (ng/Kg dry weight) | | | TEC | TEC | TEC |
|---------------------------|-------------------------|--------------|-----------|------|------|-------|
| | TOTAL DIOXINS | TOTAL FURANS | TOTAL D&F | | | |
| TEF-factor | | | | DL=0 | DL=1 | DL=.5 |
| APC | 625 | 30 | 655 | 2.4 | 10 | 6 |
| APC dup | 210 | ND | 210 | 0.2 | 24 | 12 |
| THIMBLEBERRY | 730 | 65 | 795 | 2.6 | 26 | 14 |
| JAMESTOWN | 580 | 43 | 623 | 0.9 | 26 | 13 |
| GALANKIN I | ND | ND | ND | ND | 12 | 6 |
| GALANKIN dup | ND | ND | ND | ND | 24 | 12 |
| CITY-SOUTH | 305 | 74 | 378 | 2.0 | 5 | 7 |
| CITY-JAPONSKI | 1066 | 125 | 1191 | 2.6 | 10 | 6 |
| CITY-CENTER | 80 | ND | 80 | 0.2 | 6 | 3 |
| CITY-NORTH | 166 | ND | 166 | 0.4 | 7 | 3 |
| BLUE LAKE | ND | ND | ND | ND | 24 | 12 |
| DEEP INLET | ND | ND | ND | ND | 44 | 22 |
| WATER (pg/L) | | | | | | |
| FIELD BLANK | ND | ND | ND | ND | 237 | 119 |
| BLUE LAKE | ND | ND | ND | ND | 237 | 119 |
| BLUE L. dup | ND | ND | ND | ND | 237 | 119 |
| TISSUE (ng/Kg wet weight) | | | | | | |
| BLUE L-R TROUT | ND | ND | ND | ND | 3 | 1 |
| THIMBLE L-Br TRT | ND | ND | ND | ND | 3 | 1 |
| HERRING C-QUIL | ND | 2 | 2 | 0.2 | 3 | 2 |
| SAWMILL C-QUIL | ND | 3 | 3 | 0.3 | 3 | 2 |
| THIMBLE B-QUIL | ND | 2 | 2 | 0.2 | 3 | 2 |
| SILVER B-SCULP | ND | 4 | 4 | 0.4 | 3 | 2 |
| SILVER B-E SOLE | 1 | 2 | 3 | 1.3 | 3 | 2 |
| THIMBLE B-SANDDAB | ND | 1 | 1 | 0.1 | 3 | 2 |
| SAW C-CRAB HEP | ND | 11 | 11 | 1.1 | 4 | 3 |
| SAW C-CRAB GHS | ND | 8 | 8 | 0.8 | 4 | 2 |
| SAW C-CRAB MUS | ND | 3 | 3 | 0.3 | 3 | 2 |
| THIMB B-MUSSELS | ND | 3 | 3 | 0.3 | 3 | 2 |
| SILVER B-SHRIMP | ND | 1 | 1 | 0.1 | 3 | 2 |
| FLOATING RESIDUE (ng/Kg) | | | | | | |
| SAWMILL COVE | 417 | 171 | 588 | 25 | 26 | 26 |
| EASTERN CHANNEL | 254 | 79 | 333 | 10 | 12 | 11 |
| EASTERN CHANNEL | 261 | 84 | 345 | 12 | 15 | 13 |
| SEDIMENT (ng/Kg) | | | | | | |
| HERRING COVE | 54 | 24 | 78 | 1 | 4 | 7 |
| EASTERN CHANNEL | 25 | 17 | 42 | 1 | 4 | 7 |
| SAWMILL COVE | 185 | 109 | 294 | 10 | 12 | 13 |
| BLUE LAKE | 48 | 78 | 126 | 8 | 9 | 11 |
| ASH (ng/Kg) | | | | | | |
| BOTTOM | 54 | 86 | 139 | | | |
| BOTTOM dup | ND | 38 | 38 | | | |
| MULTI CLONE | 6 | 47 | 52 | | | |
| FLY ASH - ESP | 21099 | 2930 | 24029 | | | |
| FLY ASH ESP | 47400 | 5392 | 52792 | | | |
| FLY ASH ESP | 25480 | 3859 | 29339 | | | |
| LEACHATE (pg/L) | | | | | | |
| HOGFUEL STORAGE | ND | ND | ND | | | |
| HOGF STOR-dup | ND | ND | ND | | | |
| SITKA LANDFILL | ND | ND | ND | | | |
| SLUDGE (ng/Kg) | | | | | | |
| SITKA LANDFILL | 103 | 107 | 210 | | | |

| | | | | | | | | | | | |
|------------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|
| | 2378 | 2378 | 2378 | 2378 | | 2378 | 12378 | 23478 | 2378 | 2378 | |
| Descrip | TCDD | PeCDD | HxCDD | HpCDD | OCDD | TCDF | PeCDF | PeCDF | HxCDF | HpCDF | OCDF |
| TEF-factor | 1.000 | 0.500 | 0.100 | 0.010 | 0.001 | 0.100 | 0.050 | 0.500 | 0.100 | 0.010 | 0.001 |

[illegible]

Table A . Dioxins and furans measured in samples collected in Sitka, Alaska
August 1990.

| Descrip | 2378 TCDD | 2378 PeCDD | 2378 HxCDD | 2378 HpCDD | OCDD | 2378 TCDF | 12378 PeCDF | 23478 PeCDF | 2378 HxCDF | 2378 HpCDF | OCDF |
|-------------------|--------------|---------------|---------------|---------------|-------|--------------|----------------|----------------|---------------|---------------|-------|
| TEF-factor | 1.000 | 0.500 | 0.100 | 0.010 | 0.001 | 0.100 | 0.050 | 0.500 | 0.100 | 0.010 | 0.001 |
| BLUE L-R TROUT | | | | | | | | | | | |
| THIMBLE L-Br TRT | | | | | | | | | | | |
| HERRING C-QUIL | | | | | | 2 | | | | | |
| SAWMILL C-QUIL | | | | | | 3 | | | | | |
| THIMBLE B-QUIL | | | | | | 2 | | | | | |
| SILVER B-SCULP | | | | | | 4 | | | | | |
| SILVER B-E SOLE | 1 | | | | | 2 | | | | | |
| THIMBLE B-SANDDAB | | | | | | 1 | | | | | |
| SAW C-CRAB HEP | | | | | | 11 | | | | | |
| SAW C-CRAB GHS | | | | | | 8 | | | | | |
| SAW C-CRAB MUS | | | | | | 3 | | | | | |
| THIMB B-MUSSELS | | | | | | 3 | | | | | |
| SILVER B-SHRIMP | | | | | | 1 | | | | | |

APPENDIX B. RISK CALCULATIONS

AVISO 1E-09
 ADVISO 1E-07 one day
 SLOPE 156000
 Duration Days Body weight Kg
 75 yrs 27375 Adult 70
 30 yrs 10950 Child 10
 6 year 2190

| CANCER RISK | | | | | | | | |
|-----------------|-----------|-------|----------------|-----------|-------|---------------|-----------|-------|
| TISSUE CONCENTR | | | 3 | | | | | |
| NG/KG | | | | | | | | |
| ADULT 75 years | | | ADULT 30 years | | | CHILD 6 years | | |
| ING | DOSE | RISK | conc | DOSE | RISK | conc | DOSE | RISK |
| G/Day | mg/kg/day | | ng/Kg | mg/kg/day | | ng/Kg | mg/kg/day | |
| 0.17 | 7E-12 | 1E-06 | 0.17 | 3E-12 | 5E-07 | 0.17 | 4E-12 | 6E-07 |
| 0.71 | 3E-11 | 5E-06 | 0.71 | 1E-11 | 2E-06 | 0.71 | 2E-11 | 3E-06 |
| 5 | 2E-10 | 3E-05 | 5 | 9E-11 | 1E-05 | 5 | 1E-10 | 2E-05 |
| 6.5 | 3E-10 | 4E-05 | 6.5 | 1E-10 | 2E-05 | 6.5 | 2E-10 | 2E-05 |
| 25 | 1E-09 | 2E-04 | 25 | 4E-10 | 7E-05 | 25 | 6E-10 | 9E-05 |
| 28.9 | 1E-09 | 2E-04 | 28.9 | 5E-10 | 8E-05 | 28.9 | 7E-10 | 1E-04 |
| 54 | 2E-09 | 4E-04 | 54 | 9E-10 | 1E-04 | 54 | 1E-09 | 2E-04 |
| 95.1 | 4E-09 | 6E-04 | 95.1 | 2E-09 | 3E-04 | 95.1 | 2E-09 | 4E-04 |
| 202 | 9E-09 | 1E-03 | 202 | 3E-09 | 5E-04 | 202 | 5E-09 | 8E-04 |

| NONCANCER RISK | | | | | |
|-----------------|-----------|-------|----------------|-----------|-------|
| TISSUE CONCENTR | | | 3 LIFETIME | | |
| NG/KG | | | | | |
| ADULT 75 years | | | ADULT 30 years | | |
| ING | DOSE | RATIO | conc | DOSE | RATIO |
| G/Day | mg/kg/day | | ng/Kg | mg/kg/day | |
| 0.17 | 7E-12 | 7E-03 | 0.17 | 3E-12 | 3E-03 |
| 0.71 | 3E-11 | 3E-02 | 0.71 | 1E-11 | 1E-02 |
| 5 | 2E-10 | 2E-01 | 5 | 9E-11 | 9E-02 |
| 6.5 | 3E-10 | 3E-01 | 6.5 | 1E-10 | 1E-01 |
| 25 | 1E-09 | 1E+00 | 25 | 4E-10 | 4E-01 |
| 28.9 | 1E-09 | 1E+00 | 28.9 | 5E-10 | 5E-01 |
| 54 | 2E-09 | 2E+00 | 54 | 9E-10 | 9E-01 |
| 95.1 | 4E-09 | 4E+00 | 95.1 | 2E-09 | 2E+00 |
| 202 | 9E-09 | 9E+00 | 202 | 3E-09 | 3E+00 |

| NONCANCER RISK | | | | | |
|-----------------|-----------|-------|---------------|-----------|-------|
| ONE DAY | | | 3 | | |
| TISSUE CONCENTR | | | | | |
| NG/KG | | | | | |
| ADULT 75 years | | | CHILD 6 years | | |
| DOSE | RATIO | | conc | DOSE | RATIO |
| G/Day | mg/kg/day | | G/Day | mg/kg/day | |
| 0.17 | 7E-12 | 7E-05 | 0.17 | 5E-11 | 5E-04 |
| 0.71 | 3E-11 | 3E-04 | 0.71 | 2E-10 | 2E-03 |
| 5 | 2E-10 | 2E-03 | 5 | 2E-09 | 2E-02 |
| 6.5 | 3E-10 | 3E-03 | 6.5 | 2E-09 | 2E-02 |
| 25 | 1E-09 | 1E-02 | 25 | 8E-09 | 8E-02 |
| 28.9 | 1E-09 | 1E-02 | 28.9 | 9E-09 | 9E-02 |
| 54 | 2E-09 | 2E-02 | 54 | 2E-08 | 2E-01 |
| 95.1 | 4E-09 | 4E-02 | 95.1 | 3E-08 | 3E-01 |
| 202 | 9E-09 | 9E-02 | 202 | 6E-08 | 6E-01 |

AVISO 1E-09
 ADVISO 1E-07 one day
 SLOPE 156000
 Duration Days Body weight Kg
 75 yrs 27375 Adult 70
 30 yrs 10950 Child 10
 6 year 2190

CANCER RISK

| TISSUE CONCENTR | | | 0.4 | | | | | |
|-----------------|-----------|-------|----------------|-----------|-------|---------------|-----------|-------|
| NG/KG | | | | | | | | |
| ADULT 75 years | | | ADULT 30 years | | | CHILD 6 years | | |
| ING | DOSE | RISK | conc | DOSE | RISK | conc | DOSE | RISK |
| G/Day | mg/kg/day | | ng/Kg | mg/kg/day | | ng/Kg | mg/kg/day | |
| 0.17 | 1E-12 | 2E-07 | 0.17 | 4E-13 | 6E-08 | 0.17 | 5E-13 | 8E-08 |
| 0.71 | 4E-12 | 6E-07 | 0.71 | 2E-12 | 3E-07 | 0.71 | 2E-12 | 4E-07 |
| 5 | 3E-11 | 4E-06 | 5 | 1E-11 | 2E-06 | 5 | 2E-11 | 2E-06 |
| 6.5 | 4E-11 | 6E-06 | 6.5 | 1E-11 | 2E-06 | 6.5 | 2E-11 | 3E-06 |
| 25 | 1E-10 | 2E-05 | 25 | 6E-11 | 9E-06 | 25 | 8E-11 | 1E-05 |
| 28.9 | 2E-10 | 3E-05 | 28.9 | 7E-11 | 1E-05 | 28.9 | 9E-11 | 1E-05 |
| 54 | 3E-10 | 5E-05 | 54 | 1E-10 | 2E-05 | 54 | 2E-10 | 3E-05 |
| 95.1 | 5E-10 | 8E-05 | 95.1 | 2E-10 | 3E-05 | 95.1 | 3E-10 | 5E-05 |
| 202 | 1E-09 | 2E-04 | 202 | 5E-10 | 7E-05 | 202 | 6E-10 | 1E-04 |

NONCANCER RISK

| TISSUE CONCENTR | | | 0.4 LIFETIME | | |
|-----------------|-----------|-------|----------------|-----------|-------|
| NG/KG | | | | | |
| ADULT 75 years | | | ADULT 30 years | | |
| ING | DOSE | RATIO | conc | DOSE | RATIO |
| G/Day | mg/kg/day | | ng/Kg | mg/kg/day | |
| 0.17 | 1E-12 | 1E-03 | 0.17 | 1E-12 | 1E-03 |
| 0.71 | 4E-12 | 4E-03 | 0.71 | 4E-12 | 4E-03 |
| 5 | 3E-11 | 3E-02 | 5 | 3E-11 | 3E-02 |
| 6.5 | 4E-11 | 4E-02 | 6.5 | 4E-11 | 4E-02 |
| 25 | 1E-10 | 1E-01 | 25 | 1E-10 | 1E-01 |
| 28.9 | 2E-10 | 2E-01 | 28.9 | 2E-10 | 2E-01 |
| 54 | 3E-10 | 3E-01 | 54 | 3E-10 | 3E-01 |
| 95.1 | 5E-10 | 5E-01 | 95.1 | 5E-10 | 5E-01 |
| 202 | 1E-09 | 1E+00 | 202 | 1E-09 | 1E+00 |

NONCANCER RISK

| ONE DAY | | | 0.4 | | |
|-----------------|-----------|-------|---------------|-----------|-------|
| TISSUE CONCENTR | | | | | |
| NG/KG | | | | | |
| ADULT 75 years | | | CHILD 6 years | | |
| DOSE | RATIO | | conc | DOSE | RATIO |
| G/Day | mg/kg/day | | G/Day | mg/kg/day | |
| 0.17 | 4E-12 | 4E-05 | 0.17 | 7E-12 | 7E-05 |
| 0.71 | 3E-11 | 3E-04 | 0.71 | 3E-11 | 3E-04 |
| 5 | 4E-11 | 4E-04 | 5 | 2E-10 | 2E-03 |
| 6.5 | 1E-10 | 1E-03 | 6.5 | 3E-10 | 3E-03 |
| 25 | 2E-10 | 2E-03 | 25 | 1E-09 | 1E-02 |
| 28.9 | 3E-10 | 3E-03 | 28.9 | 1E-09 | 1E-02 |
| 54 | 5E-10 | 5E-03 | 54 | 2E-09 | 2E-02 |
| 95.1 | 1E-09 | 1E-02 | 95.1 | 4E-09 | 4E-02 |
| 202 | 0E+00 | 0E+00 | 202 | 8E-09 | 8E-02 |

AVISO 1E-09
 ADVISO 1E-07 one day
 SLOPE 156000
 Duration Days Body weight Kg
 75 yrs 27375 Adult 70
 30 yrs 10950 Child 10
 6 year 2190

CANCER RISK

| TISSUE CONCENTR | | 1 | | | | | | | | |
|-----------------|-----------|-------|----------------|-----------|-------|----------------|-----------|-------|---------------|-----------|
| NG/KG | | | ADULT 75 years | | | ADULT 30 years | | | CHILD 6 years | |
| ING | DOSE | RISK | conc | DOSE | RISK | conc | DOSE | RISK | conc | DOSE |
| G/Day | mg/kg/day | | ng/Kg | mg/kg/day | | ng/Kg | mg/kg/day | | ng/Kg | mg/kg/day |
| 0.17 | 2E-12 | 4E-07 | 0.17 | 1E-12 | 2E-07 | 0.17 | 1E-12 | 2E-07 | 0.17 | 1E-12 |
| 0.71 | 1E-11 | 2E-06 | 0.71 | 4E-12 | 6E-07 | 0.71 | 6E-12 | 9E-07 | 0.71 | 6E-12 |
| 5 | 7E-11 | 1E-05 | 5 | 3E-11 | 4E-06 | 5 | 4E-11 | 6E-06 | 5 | 4E-11 |
| 6.5 | 9E-11 | 1E-05 | 6.5 | 4E-11 | 6E-06 | 6.5 | 5E-11 | 8E-06 | 6.5 | 5E-11 |
| 25 | 4E-10 | 6E-05 | 25 | 1E-10 | 2E-05 | 25 | 2E-10 | 3E-05 | 25 | 2E-10 |
| 28.9 | 4E-10 | 6E-05 | 28.9 | 2E-10 | 3E-05 | 28.9 | 2E-10 | 4E-05 | 28.9 | 2E-10 |
| 54 | 8E-10 | 1E-04 | 54 | 3E-10 | 5E-05 | 54 | 4E-10 | 7E-05 | 54 | 4E-10 |
| 95.1 | 1E-09 | 2E-04 | 95.1 | 5E-10 | 8E-05 | 95.1 | 8E-10 | 1E-04 | 95.1 | 8E-10 |
| 202 | 3E-09 | 5E-04 | 202 | 1E-09 | 2E-04 | 202 | 2E-09 | 3E-04 | 202 | 2E-09 |

NONCANCER RISK 1 LIFETIME

| TISSUE CONCENTR | | 1 | | | | | | | | |
|-----------------|-----------|-------|----------------|-----------|-------|----------------|-----------|-------|--|--|
| NG/KG | | | ADULT 75 years | | | ADULT 30 years | | | | |
| ING | DOSE | RATIO | conc | DOSE | RATIO | conc | DOSE | RATIO | | |
| G/Day | mg/kg/day | | ng/Kg | mg/kg/day | | ng/Kg | mg/kg/day | | | |
| 0.17 | 2E-12 | 2E-03 | 0.17 | 1E-12 | 1E-03 | 0.17 | 1E-12 | 1E-03 | | |
| 0.71 | 1E-11 | 1E-02 | 0.71 | 4E-12 | 4E-03 | 0.71 | 4E-12 | 4E-03 | | |
| 5 | 7E-11 | 7E-02 | 5 | 3E-11 | 3E-02 | 5 | 3E-11 | 3E-02 | | |
| 6.5 | 9E-11 | 9E-02 | 6.5 | 4E-11 | 4E-02 | 6.5 | 4E-11 | 4E-02 | | |
| 25 | 4E-10 | 4E-01 | 25 | 1E-10 | 1E-01 | 25 | 1E-10 | 1E-01 | | |
| 28.9 | 4E-10 | 4E-01 | 28.9 | 2E-10 | 2E-01 | 28.9 | 2E-10 | 2E-01 | | |
| 54 | 8E-10 | 8E-01 | 54 | 3E-10 | 3E-01 | 54 | 3E-10 | 3E-01 | | |
| 95.1 | 1E-09 | 1E+00 | 95.1 | 5E-10 | 5E-01 | 95.1 | 5E-10 | 5E-01 | | |
| 202 | 3E-09 | 3E+00 | 202 | 1E-09 | 1E+00 | 202 | 1E-09 | 1E+00 | | |

NONCANCER RISK

| ONE DAY TISSUE CONCENTR | | 1 | | | | | | | | |
|----------------------------|-----------|-------|----------------|-----------|-------|---------------|-----------|-------|--|--|
| NG/KG | | | ADULT 75 years | | | CHILD 6 years | | | | |
| ING | DOSE | RATIO | conc | DOSE | RATIO | conc | DOSE | RATIO | | |
| G/Day | mg/kg/day | | G/Day | mg/kg/day | | G/Day | mg/kg/day | | | |
| 0.17 | 2E-12 | 2E-05 | 0.17 | 2E-11 | 2E-04 | 0.17 | 2E-11 | 2E-04 | | |
| 0.71 | 1E-11 | 1E-04 | 0.71 | 7E-11 | 7E-04 | 0.71 | 7E-11 | 7E-04 | | |
| 5 | 7E-11 | 7E-04 | 5 | 5E-10 | 5E-03 | 5 | 5E-10 | 5E-03 | | |
| 6.5 | 9E-11 | 9E-04 | 6.5 | 7E-10 | 7E-03 | 6.5 | 7E-10 | 7E-03 | | |
| 25 | 4E-10 | 4E-03 | 25 | 3E-09 | 3E-02 | 25 | 3E-09 | 3E-02 | | |
| 28.9 | 4E-10 | 4E-03 | 28.9 | 3E-09 | 3E-02 | 28.9 | 3E-09 | 3E-02 | | |
| 54 | 8E-10 | 8E-03 | 54 | 5E-09 | 5E-02 | 54 | 5E-09 | 5E-02 | | |
| 95.1 | 1E-09 | 1E-02 | 95.1 | 1E-08 | 1E-01 | 95.1 | 1E-08 | 1E-01 | | |
| 202 | 3E-09 | 3E-02 | 202 | 2E-08 | 2E-01 | 202 | 2E-08 | 2E-01 | | |

Table B Soil Risk Calculations

| | | | |
|------|----------------|-------------------|--------------------------------|
| DOSE | noncancer derm | 7.9E-05 kg/kgday | Conc X ADI X abs X 1E-06 mg/ng |
| DOSE | cancer dermal | 3.4E-05 kg/kgday | ng/kg kg/kgday mg/ng |
| | sloper factor | 156000 \mg/kg/day | |
| | one day Hlth | 1.0E-07 mg/kg/day | slope factor X ADI |
| DOSE | noncancer soil | 3.7E-06 kg/kgday | ADI/RFD |
| DOSE | cancer soil | 1.6E-06 kg/kgday | |

DERMAL CANCER RISK ESTIMATE

| | ng/Kg | ng/Kg | ng/Kg | DOSE | DOSE | DOSE | RISK | RISK | RISK |
|--------------|---------|---------|---------|---------|---------|---------|---------|---------|---------|
| TEF-factor | TEC | TEC | TEC | TEC | TEC | TEC | TEC | TEC | TEC |
| LOCATION | DL=0.00 | DL=0.50 | DL=1.00 | DL=0.00 | DL=0.50 | DL=1.00 | DL=0.00 | DL=0.50 | DL=1.00 |
| APC-DUP | 0.21 | 12.07 | 23.92 | 1.4E-12 | 8.2E-11 | 1.6E-10 | 2E-08 | 1E-06 | 3E-06 |
| APC | 2.45 | 6.04 | 9.62 | 1.7E-11 | 4.1E-11 | 6.5E-11 | 3E-07 | 6E-07 | 1E-06 |
| GALANKIN I | | 11.86 | 23.72 | 0 | 8.1E-11 | 1.6E-10 | 0E+00 | 1E-06 | 3E-06 |
| GALANKIN I | 0.03 | 6.20 | 12.37 | 2.4E-13 | 4.2E-11 | 8.4E-11 | 4E-09 | 7E-07 | 1E-06 |
| BLUE LAKE | | 11.86 | 23.72 | 0 | 8.1E-11 | 1.6E-10 | 0E+00 | 1E-06 | 3E-06 |
| JAMESTOWN | 0.90 | 13.40 | 25.90 | 6.1E-12 | 9.1E-11 | 1.8E-10 | 1E-07 | 1E-06 | 3E-06 |
| THIMBLEBERRY | 2.59 | 14.34 | 26.09 | 1.8E-11 | 9.7E-11 | 1.8E-10 | 3E-07 | 2E-06 | 3E-06 |
| DEEP INLET | | 21.87 | 43.74 | 0 | 1.5E-10 | 3.0E-10 | 0E+00 | 2E-06 | 5E-06 |
| CITY-N | 0.37 | 3.44 | 6.51 | 2.5E-12 | 2.3E-11 | 4.4E-11 | 4E-08 | 4E-07 | 7E-07 |
| CITY-C | 0.18 | 3.11 | 6.05 | 1.2E-12 | 2.1E-11 | 4.1E-11 | 2E-08 | 3E-07 | 6E-07 |
| CITY-JAPO | 2.59 | 6.19 | 9.78 | 1.8E-11 | 4.2E-11 | 6.7E-11 | 3E-07 | 7E-07 | 1E-06 |
| CITY-S | 1.98 | 4.65 | 7.33 | 1.3E-11 | 3.2E-11 | 5.0E-11 | 2E-07 | 5E-07 | 8E-07 |
| F. RESIDUE | 25 | 26 | 26 | 1.7E-10 | 1.8E-10 | 1.8E-10 | 3E-06 | 3E-06 | 3E-06 |
| F. RESIDUE | 10 | 11 | 12 | 6.8E-11 | 7.5E-11 | 8.2E-11 | 1E-06 | 1E-06 | 1E-06 |
| F. RESIDUE | 12 | 13 | 15 | 8.2E-11 | 8.8E-11 | 1.0E-10 | 1E-06 | 1E-06 | 2E-06 |

SOIL INGESTION CANCER RISK

| | ng/Kg | ng/Kg | ng/Kg | DOSE | DOSE | DOSE | RISK | RISK | RISK |
|--------------|---------|---------|---------|---------|---------|---------|---------|---------|---------|
| TEF-factor | TEC | TEC | TEC | TEC | TEC | TEC | TEC | TEC | TEC |
| LOCATION | DL=0.00 | DL=0.50 | DL=1.00 | DL=0.00 | DL=0.50 | DL=1.00 | DL=0.00 | DL=0.50 | DL=1.00 |
| APC-DUP | 0.21 | 12.07 | 23.92 | 3.4E-13 | 2.0E-11 | 3.9E-11 | 5E-08 | 3E-06 | 6E-06 |
| APC | 2.45 | 6.04 | 9.62 | 4.0E-12 | 9.8E-12 | 1.6E-11 | 6E-07 | 2E-06 | 2E-06 |
| GALANKIN I | | 11.86 | 23.72 | 0 | 1.9E-11 | 3.9E-11 | 0E+00 | 3E-06 | 6E-06 |
| GALANKIN I | 0.03 | 6.20 | 12.37 | 5.7E-14 | 1.0E-11 | 2.0E-11 | 9E-09 | 2E-06 | 3E-06 |
| BLUE LAKE | | 11.86 | 23.72 | 0 | 1.9E-11 | 3.9E-11 | 0E+00 | 3E-06 | 6E-06 |
| JAMESTOWN | 0.90 | 13.40 | 25.90 | 1.5E-12 | 2.2E-11 | 4.2E-11 | 2E-07 | 3E-06 | 7E-06 |
| THIMBLEBERRY | 2.59 | 14.34 | 26.09 | 4.2E-12 | 2.3E-11 | 4.3E-11 | 7E-07 | 4E-06 | 7E-06 |
| DEEP INLET | | 21.87 | 43.74 | 0 | 3.6E-11 | 7.1E-11 | 0E+00 | 6E-06 | 1E-05 |
| CITY-N | 0.37 | 3.44 | 6.51 | 6.0E-13 | 5.6E-12 | 1.1E-11 | 9E-08 | 9E-07 | 2E-06 |
| CITY-C | 0.18 | 3.11 | 6.05 | 3.0E-13 | 5.1E-12 | 9.9E-12 | 5E-08 | 8E-07 | 2E-06 |
| CITY-JAPO | 2.59 | 6.19 | 9.78 | 4.2E-12 | 1.0E-11 | 1.6E-11 | 7E-07 | 2E-06 | 2E-06 |
| CITY-S | 1.98 | 4.65 | 7.33 | 3.2E-12 | 7.6E-12 | 1.2E-11 | 5E-07 | 1E-06 | 2E-06 |
| F. RESIDUE | 25 | 26 | 26 | 4.1E-11 | 4.2E-11 | 4.2E-11 | 6E-06 | 7E-06 | 7E-06 |
| F. RESIDUE | 10 | 11 | 12 | 1.6E-11 | 1.8E-11 | 2.0E-11 | 3E-06 | 3E-06 | 3E-06 |
| F. RESIDUE | 12 | 13 | 15 | 2.0E-11 | 2.1E-11 | 2.4E-11 | 3E-06 | 3E-06 | 4E-06 |

TABLE B SOIL CUMULATIVE RISK CALCULATIONS

| | DL=0.00 | | | DL=0.50 | | | DL=1.0 | | |
|--------------|---------|--------|-------|---------|--------|-------|--------|--------|-------|
| LOCATION | DERMAL | INGEST | SUM | DERMAL | INGEST | SUM | DERMAL | INGEST | SUM |
| APC-DUP | 2E-08 | 5E-08 | 8E-08 | 1E-06 | 5E-08 | 1E-06 | 3E-06 | 6E-06 | 9E-06 |
| APC | 3E-07 | 6E-07 | 9E-07 | 6E-07 | 6E-07 | 1E-06 | 1E-06 | 2E-06 | 3E-06 |
| GALANKIN I | 0E+00 | 0E+00 | 0E+00 | 1E-06 | 0E+00 | 1E-06 | 3E-06 | 6E-06 | 9E-06 |
| GALANKIN I | 4E-09 | 9E-09 | 1E-08 | 7E-07 | 9E-09 | 7E-07 | 1E-06 | 3E-06 | 4E-06 |
| BLUE LAKE | 0E+00 | 0E+00 | 0E+00 | 1E-06 | 0E+00 | 1E-06 | 3E-06 | 6E-06 | 9E-06 |
| JAMESTOWN | 1E-07 | 2E-07 | 3E-07 | 1E-06 | 2E-07 | 2E-06 | 3E-06 | 7E-06 | 9E-06 |
| THIMBLEBERRY | 3E-07 | 7E-07 | 9E-07 | 2E-06 | 7E-07 | 2E-06 | 3E-06 | 7E-06 | 9E-06 |
| DEEP INLET | 0E+00 | 0E+00 | 0E+00 | 2E-06 | 0E+00 | 2E-06 | 5E-06 | 1E-05 | 2E-05 |
| CITY-N | 4E-08 | 9E-08 | 1E-07 | 4E-07 | 9E-08 | 5E-07 | 7E-07 | 2E-06 | 2E-06 |
| CITY-C | 2E-08 | 5E-08 | 7E-08 | 3E-07 | 5E-08 | 4E-07 | 6E-07 | 2E-06 | 2E-06 |
| CITY-JAPO | 3E-07 | 7E-07 | 9E-07 | 7E-07 | 7E-07 | 1E-06 | 1E-06 | 2E-06 | 4E-06 |
| CITY-S | 2E-07 | 5E-07 | 7E-07 | 5E-07 | 5E-07 | 1E-06 | 8E-07 | 2E-06 | 3E-06 |
| F RESIDUE | 3E-06 | 6E-06 | 9E-06 | 3E-06 | 6E-06 | 9E-06 | 3E-06 | 7E-06 | 9E-06 |
| F RESIDUE | 1E-06 | 3E-06 | 4E-06 | 1E-06 | 3E-06 | 4E-06 | 1E-06 | 3E-06 | 4E-06 |
| F RESIDUE | 1E-06 | 3E-06 | 4E-06 | 1E-06 | 3E-06 | 4E-06 | 2E-06 | 4E-06 | 5E-06 |

TABLE 5. SOIL NON-CANCER LIFETIME RISK ESTIMATES

| | | | | | |
|------|----------------|---------|------------|--------------------------|-------|
| DOSE | noncancer derm | 7.9E-05 | kg/kgday | Conc X ADI X abs X 1E-06 | mg/ng |
| DOSE | cancer dermal | 3.4E-05 | kg/kgday | ng/kg kg/kgday mg/ng | |
| | sloper factor | 156000 | \mg/kg/day | | |
| | lifetime | 1.0E-09 | mg/kg/day | slope factor X ADI | |
| | one day Hlth | 1.0E-07 | mg/kg/day | ADI/RFD | |
| DOSE | noncancer soil | 3.7E-06 | kg/kgday | | |
| DOSE | cancer soil | 1.6E-06 | kg/kgday | | |

DERMAL NON-CANCER RISK ESTIMATE

| TEF- LOCATION | ng/Kg TEC DL=0.00 | ng/Kg TEC DL=0.50 | ng/Kg TEC DL=1.00 | DOSE TEC DL=0.00 | DOSE TEC DL=0.50 | DOSE TEC DL=1.00 | RATIO TEC DL=0.00 | RATIO TEC DL=0.50 | RATIO TEC DL=1.00 |
|------------------|-------------------------|-------------------------|-------------------------|------------------------|------------------------|------------------------|-------------------------|-------------------------|-------------------------|
| APC-DUP | 0.21 | 12.07 | 23.92 | 1.4E-12 | 8.2E-11 | 1.6E-10 | 1E-04 | 8E-03 | 2E-02 |
| APC | 2.45 | 6.04 | 9.62 | 1.7E-11 | 4.1E-11 | 6.5E-11 | 2E-03 | 4E-03 | 7E-03 |
| GALANKIN I | | 11.86 | 23.72 | 0 | 8.1E-11 | 1.6E-10 | 0E+00 | 8E-03 | 2E-02 |
| GALANKIN | 0.03 | 6.20 | 12.37 | 2.4E-13 | 4.2E-11 | 8.4E-11 | 2E-05 | 4E-03 | 8E-03 |
| BLUE LAKE | | 11.86 | 23.72 | 0 | 8.1E-11 | 1.6E-10 | 0E+00 | 8E-03 | 2E-02 |
| JAMESTOWN | 0.90 | 13.40 | 25.90 | 6.1E-12 | 9.1E-11 | 1.8E-10 | 6E-04 | 9E-03 | 2E-02 |
| THIMBLEBE | 2.59 | 14.34 | 26.09 | 1.8E-11 | 9.7E-11 | 1.8E-10 | 2E-03 | 1E-02 | 2E-02 |
| DEEP INLET | | 21.87 | 43.74 | 0 | 1.5E-10 | 3.0E-10 | 0E+00 | 1E-02 | 3E-02 |
| CITY-N | 0.37 | 3.44 | 6.51 | 2.5E-12 | 2.3E-11 | 4.4E-11 | 3E-04 | 2E-03 | 4E-03 |
| CITY-C | 0.18 | 3.11 | 6.05 | 1.2E-12 | 2.1E-11 | 4.1E-11 | 1E-04 | 2E-03 | 4E-03 |
| CITY-JAPO | 2.59 | 6.19 | 9.78 | 1.8E-11 | 4.2E-11 | 6.7E-11 | 2E-03 | 4E-03 | 7E-03 |
| CITY-S | 1.98 | 4.65 | 7.33 | 1.3E-11 | 3.2E-11 | 5.0E-11 | 1E-03 | 3E-03 | 5E-03 |
| F. RESIDU | 25 | 26 | 26 | 1.7E-10 | 1.8E-10 | 1.8E-10 | 2E-02 | 2E-02 | 2E-02 |
| F. RESIDU | 10 | 11 | 12 | 6.8E-11 | 7.5E-11 | 8.2E-11 | 7E-03 | 7E-03 | 8E-03 |
| F. RESIDU | 12 | 13 | 15 | 8.2E-11 | 8.8E-11 | 1.0E-10 | 8E-03 | 9E-03 | 1E-02 |

SOIL INGESTION NON-CANCER RISK

| TEF- LOCATION | ng/Kg TEC DL=0.00 | ng/Kg TEC DL=0.50 | ng/Kg TEC DL=1.00 | DOSE TEC DL=0.00 | DOSE TEC DL=0.50 | DOSE TEC DL=1.00 | RATIO TEC DL=0.00 | RATIO TEC DL=0.50 | RATIO TEC DL=1.00 |
|------------------|-------------------------|-------------------------|-------------------------|------------------------|------------------------|------------------------|-------------------------|-------------------------|-------------------------|
| APC-DUP | 0.21 | 12.07 | 23.92 | 7.8E-13 | 4.5E-11 | 8.9E-11 | 0.00 | 0.04 | 0.09 |
| APC | 2.45 | 6.04 | 9.62 | 9.1E-12 | 2.2E-11 | 3.6E-11 | 0.01 | 0.02 | 0.04 |
| GALANKIN I | | 11.86 | 23.72 | 0 | 4.4E-11 | 8.8E-11 | 0.00 | 0.04 | 0.09 |
| GALANKIN | 0.03 | 6.20 | 12.37 | 1.3E-13 | 2.3E-11 | 4.6E-11 | 0.00 | 0.02 | 0.05 |
| BLUE LAKE | | 11.86 | 23.72 | 0 | 4.4E-11 | 8.8E-11 | 0.00 | 0.04 | 0.09 |
| JAMESTOWN | 0.90 | 13.40 | 25.90 | 3.3E-12 | 5.0E-11 | 9.6E-11 | 0.00 | 0.05 | 0.10 |
| THIMBLEBE | 2.59 | 14.34 | 26.09 | 9.6E-12 | 5.3E-11 | 9.7E-11 | 0.01 | 0.05 | 0.10 |
| DEEP INLET | | 21.87 | 43.74 | 0 | 8.1E-11 | 1.6E-10 | 0.00 | 0.08 | 0.16 |
| CITY-N | 0.37 | 3.44 | 6.51 | 1.4E-12 | 1.3E-11 | 2.4E-11 | 0.00 | 0.01 | 0.02 |
| CITY-C | 0.18 | 3.11 | 6.05 | 6.7E-13 | 1.2E-11 | 2.2E-11 | 0.00 | 0.01 | 0.02 |
| CITY-JAPO | 2.59 | 6.19 | 9.78 | 9.6E-12 | 2.3E-11 | 3.6E-11 | 0.01 | 0.02 | 0.04 |
| CITY-S | 1.98 | 4.65 | 7.33 | 7.3E-12 | 1.7E-11 | 2.7E-11 | 0.01 | 0.02 | 0.03 |
| F. RESIDU | 25 | 26 | 26 | 9.3E-11 | 9.6E-11 | 9.6E-11 | 0.09 | 0.10 | 0.10 |
| F. RESIDU | 10 | 11 | 12 | 3.7E-11 | 4.1E-11 | 4.4E-11 | 0.04 | 0.04 | 0.04 |
| F. RESIDU | 12 | 13 | 15 | 4.4E-11 | 4.8E-11 | 5.6E-11 | 0.04 | 0.05 | 0.06 |

| | DL=0 | | | DL=.5 | | | DL=1 | | |
|--------------|--------|--------|------|--------|--------|------|--------|--------|------|
| LOCATION | DERMAL | INGEST | SUM | DERMAL | INGEST | SUM | DERMAL | INGEST | SUM |
| APC-DUP | 0.00 | 0.00 | 0.00 | 0.01 | 0.04 | 0.05 | 0.02 | 0.09 | 0.10 |
| APC | 0.00 | 0.01 | 0.01 | 0.00 | 0.02 | 0.03 | 0.01 | 0.04 | 0.04 |
| GALANKIN I | 0.00 | 0.00 | 0.00 | 0.01 | 0.04 | 0.05 | 0.02 | 0.09 | 0.10 |
| GALANKIN I | 0.00 | 0.00 | 0.00 | 0.00 | 0.02 | 0.03 | 0.01 | 0.05 | 0.05 |
| BLUE LAKE | 0.00 | 0.00 | 0.00 | 0.01 | 0.04 | 0.05 | 0.02 | 0.09 | 0.10 |
| JAMESTOWN | 0.00 | 0.00 | 0.00 | 0.01 | 0.05 | 0.06 | 0.02 | 0.10 | 0.11 |
| THIMBLEBERRY | 0.00 | 0.01 | 0.01 | 0.01 | 0.05 | 0.06 | 0.02 | 0.10 | 0.11 |
| DEEP INLET | 0.00 | 0.00 | 0.00 | 0.01 | 0.08 | 0.10 | 0.03 | 0.16 | 0.19 |
| CITY-N | 0.00 | 0.00 | 0.00 | 0.00 | 0.01 | 0.02 | 0.00 | 0.02 | 0.03 |
| CITY-C | 0.00 | 0.00 | 0.00 | 0.00 | 0.01 | 0.01 | 0.00 | 0.02 | 0.03 |
| CITY-JAPO | 0.00 | 0.01 | 0.01 | 0.00 | 0.02 | 0.03 | 0.01 | 0.04 | 0.04 |
| CITY-S | 0.00 | 0.01 | 0.01 | 0.00 | 0.02 | 0.02 | 0.00 | 0.03 | 0.03 |
| F. RESIDUE | 0.02 | 0.09 | 0.11 | 0.02 | 0.10 | 0.11 | 0.02 | 0.10 | 0.11 |
| F. RESIDUE | 0.01 | 0.04 | 0.04 | 0.01 | 0.04 | 0.05 | 0.01 | 0.04 | 0.05 |
| F. RESIDUE | 0.01 | 0.04 | 0.05 | 0.01 | 0.05 | 0.06 | 0.01 | 0.06 | 0.07 |

Table B Soil Risk Calculations

| | | | | | | |
|------|----------------|---------|------------|-------|--------------------|-------|
| DOSE | cancer dermal | 3.4E-05 | kg/kgday | ng/kg | kg/kgday | mg/ng |
| | sloper factor | 156000 | \mg/kg/day | | | |
| | lifetime | 1.0E-09 | mg/kg/day | | slope factor X ADI | |
| | one day Hlth | 1.0E-07 | mg/kg/day | | ADI/RFD | |
| DOSE | noncancer soil | 3.7E-06 | kg/kgday | | | |
| DOSE | cancer soil | 1.6E-06 | kg/kgday | | | |

CHILD ADI 200 mg/day x 1e-12kg/ng /15 Kg

1.3E-11 mg/ng/day

| TEF-factor | ng/Kg | | | DOSE | | | RATIO | | |
|--------------|---------|---------|---------|---------|---------|---------|---------|---------|---------|
| | TEC | TEC | TEC | TEC | TEC | TEC | TEC | TEC | TEC |
| LOCATION | DL=0.00 | DL=0.50 | DL=1.00 | DL=0.00 | DL=0.50 | DL=1.00 | DL=0.00 | DL=0.50 | DL=1.00 |
| APC-DUP | 0.21 | 12.07 | 23.92 | 2.8E-12 | 1.6E-10 | 3.2E-10 | 0.00 | 0.00 | 0.00 |
| APC | 2.45 | 6.04 | 9.62 | 3.3E-11 | 8.0E-11 | 1.3E-10 | 0.00 | 0.00 | 0.00 |
| GALANKIN I | | 11.86 | 23.72 | 0.0E+00 | 1.6E-10 | 3.2E-10 | 0.00 | 0.00 | 0.00 |
| GALANKIN I | 0.03 | 6.20 | 12.37 | 4.6E-13 | 8.3E-11 | 1.6E-10 | 0.00 | 0.00 | 0.00 |
| BLUE LAKE | | 11.86 | 23.72 | 0.0E+00 | 1.6E-10 | 3.2E-10 | 0.00 | 0.00 | 0.00 |
| JAMESTOWN | 0.90 | 13.40 | 25.90 | 1.2E-11 | 1.8E-10 | 3.5E-10 | 0.00 | 0.00 | 0.00 |
| THIMBLEBERRY | 2.59 | 14.34 | 26.09 | 3.4E-11 | 1.9E-10 | 3.5E-10 | 0.00 | 0.00 | 0.00 |
| DEEP INLET | | 21.87 | 43.74 | 0.0E+00 | 2.9E-10 | 5.8E-10 | 0.00 | 0.00 | 0.01 |
| CITY-N | 0.37 | 3.44 | 6.51 | 4.9E-12 | 4.6E-11 | 8.7E-11 | 0.00 | 0.00 | 0.00 |
| CITY-C | 0.18 | 3.11 | 6.05 | 2.4E-12 | 4.2E-11 | 8.1E-11 | 0.00 | 0.00 | 0.00 |
| CITY-JAPO | 2.59 | 6.19 | 9.78 | 3.5E-11 | 8.3E-11 | 1.3E-10 | 0.00 | 0.00 | 0.00 |
| CITY-S | 1.98 | 4.65 | 7.33 | 2.6E-11 | 6.2E-11 | 9.8E-11 | 0.00 | 0.00 | 0.00 |
| F. RESIDUE | 25 | 26 | 26 | 3.3E-10 | 3.5E-10 | 3.5E-10 | 0.00 | 0.00 | 0.00 |
| F. RESIDUE | 10 | 11 | 12 | 1.3E-10 | 1.5E-10 | 1.6E-10 | 0.00 | 0.00 | 0.00 |
| F. RESIDUE | 12 | 13 | 15 | 1.6E-10 | 1.7E-10 | 2.0E-10 | 0.00 | 0.00 | 0.00 |

CHILD (3900cm2 x 1mg/cm2 x 1e-12 kg/ng)/15Kg

2.6E-10 mg/ng/day

| TEF-factor | ng/Kg | | | DOSE | | | RATIO | | |
|--------------|---------|---------|---------|---------|---------|---------|---------|---------|---------|
| | TEC | TEC | TEC | TEC | TEC | TEC | TEC | TEC | TEC |
| LOCATION | DL=0.00 | DL=0.50 | DL=1.00 | DL=0.00 | DL=0.50 | DL=1.00 | DL=0.00 | DL=0.50 | DL=1.00 |
| APC-DUP | 0.21 | 12.07 | 23.92 | 5.5E-11 | 3.1E-09 | 6.2E-09 | 0.00 | 0.00 | 0.01 |
| APC | 2.45 | 6.04 | 9.62 | 6.4E-10 | 1.6E-09 | 2.5E-09 | 0.00 | 0.00 | 0.00 |
| GALANKIN I | | 11.86 | 23.72 | 0.0E+00 | 3.1E-09 | 6.2E-09 | 0.00 | 0.00 | 0.01 |
| GALANKIN I | 0.03 | 6.20 | 12.37 | 9.0E-12 | 1.6E-09 | 3.2E-09 | 0.00 | 0.00 | 0.00 |
| BLUE LAKE | | 11.86 | 23.72 | 0.0E+00 | 3.1E-09 | 6.2E-09 | 0.00 | 0.00 | 0.01 |
| JAMESTOWN | 0.90 | 13.40 | 25.90 | 2.3E-10 | 3.5E-09 | 6.7E-09 | 0.00 | 0.00 | 0.01 |
| THIMBLEBERRY | 2.59 | 14.34 | 26.09 | 6.7E-10 | 3.7E-09 | 6.8E-09 | 0.00 | 0.00 | 0.01 |
| DEEP INLET | | 21.87 | 43.74 | 0.0E+00 | 5.7E-09 | 1.1E-08 | 0.00 | 0.01 | 0.01 |
| CITY-N | 0.37 | 3.44 | 6.51 | 9.6E-11 | 8.9E-10 | 1.7E-09 | 0.00 | 0.00 | 0.00 |
| CITY-C | 0.18 | 3.11 | 6.05 | 4.7E-11 | 8.1E-10 | 1.6E-09 | 0.00 | 0.00 | 0.00 |
| CITY-JAPO | 2.59 | 6.19 | 9.78 | 6.7E-10 | 1.6E-09 | 2.5E-09 | 0.00 | 0.00 | 0.00 |
| CITY-S | 1.98 | 4.65 | 7.33 | 5.1E-10 | 1.2E-09 | 1.9E-09 | 0.00 | 0.00 | 0.00 |
| F. RESIDUE | 25 | 26 | 26 | 6.5E-09 | 6.8E-09 | 6.8E-09 | 0.01 | 0.01 | 0.01 |
| F. RESIDUE | 10 | 11 | 12 | 2.6E-09 | 2.9E-09 | 3.1E-09 | 0.00 | 0.00 | 0.00 |
| F. RESIDUE | 12 | 13 | 15 | 3.1E-09 | 3.4E-09 | 3.9E-09 | 0.00 | 0.00 | 0.00 |

ONE DAY CHILD HEALTH ADVISORY

| LOCATION | DL=0 | | | DL=.5 | | | DL=1 | | |
|--------------|--------|--------|------|--------|--------|------|--------|--------|------|
| | DERMAL | INGEST | SUM | DERMAL | INGEST | SUM | DERMAL | INGEST | SUM |
| APC-DUP | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.01 | 0.00 |
| APC | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| GALANKIN I | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.01 | 0.00 | 0.01 |
| GALANKIN I | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| BLUE LAKE | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.01 | 0.00 | 0.01 |
| JAMESTOWN | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.01 | 0.01 | 0.00 | 0.01 |
| THIMBLEBERRY | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.01 | 0.01 | 0.00 | 0.01 |
| DEEP INLET | 0.00 | 0.00 | 0.00 | 0.01 | 0.00 | 0.01 | 0.01 | 0.01 | 0.02 |
| CITY-N | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| CITY-C | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| CITY-JAPO | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| CITY-S | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| F. RESIDUE | 0.01 | 0.00 | 0.01 | 0.01 | 0.00 | 0.01 | 0.01 | 0.00 | 0.01 |
| F. RESIDUE | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| F. RESIDUE | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.01 | 0.00 | 0.00 | 0.01 |

APPENDIX C. RAW DATA

- C-1. DIOXINS AND FURANS
- C-2. ORGANICS
- C-3. METALS
- C-4. CONVENTIONALS
- C-5. AQUATIC ORGANISMS
- C-6. LOCATION OF SOIL SAMPLES

C-1. DIOXINS AND FURANS

EPA August 1990 investigation

| L a Type b | Sample | | Units | 2378 | 2378 | 2378 | 2378 | OCDD | 2378 | 12378 | 23478 | 2378 | 2378 | OCDF |
|------------------|--------|-----------------------|-------|---------------|----------------|----------------|----------------|------|---------------|----------------|----------------|----------------|----------------|------|
| | No. | Descrip TEF-factor | | TCDD 1.000 | PeCDD 0.500 | HxCDD 0.100 | HpCDD 0.010 | | TCDF 0.100 | PeCDF 0.050 | PeCDF 0.500 | HxCDF 0.100 | HpCDF 0.010 | |

Description of Data qualifiers is attached.

| | | | | | | | | | | | | | | |
|-------------|------|-------------|-------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-------|
| CL Ash | 4269 | HOGF-BOTTOM | ng/Kg | 2.17 U | 3.90 U | 6.42 | 18.30 | 28.80 | 54.70 | 9.48 | 7.12 | 6.34 U | 14.20 | 9.50 |
| CL Ash | 4270 | MULTI CLON | ng/Kg | 2.32 | 3.22 | 5.36 U | 4.42 U | 9.90 U | 31.50 | 5.72 | 4.08 | 6.49 U | 5.62 U | 9.90 |
| CL Soil | 4721 | DEEP INLET | ng/Kg | 17.90 UJ3 | 14.40 UJ3 | 39.80 UJ3 | 40.80 UJ3 | 99.10 UJ3 | 16.30 UJ3 | 14.60 UJ3 | 14.60 UJ3 | 40.00 UJ3 | 39.10 UJ3 | 99.10 |
| CL Soil | 4730 | GALANKIN I | ng/Kg | 3.89 U | 2.97 U | 35.80 U | 8.37 U | 34.70 | 3.86 U | 3.16 U | 3.16 U | 10.60 U | 9.25 U | 18.20 |
| CL Soil | 4731 | APC | ng/Kg | 3.06 U | 2.32 U | 10.50 | 83.70 | 531.00 | 5.40 U | 2.77 U | 2.77 U | 7.67 U | 12.50 UJ | 30.20 |
| CL Leachate | 4740 | SITKA LAND | pg/L | 24.20 UJ3 | 18.40 UJ3 | 57.30 UJ3 | 44.30 UJ3 | 92.70 UJ3 | 19.10 UJ3 | 20.20 UJ3 | 20.20 UJ3 | 61.40 UJ3 | 58.10 UJ3 | 92.70 |
| CL Soil | 4741 | CITY-N | ng/Kg | 2.53 U | 2.06 U | 6.07 U | 22.50 | 143.00 | 2.52 U | 1.97 U | 1.97 U | 5.69 U | 5.97 U | 14.10 |
| CL Soil | 4742 | CITY-C | ng/Kg | 2.48 U | 1.79 U | 5.45 U | 11.20 | 69.00 | 2.26 U | 1.97 U | 1.97 U | 5.79 U | 4.92 U | 9.41 |
| CL Soil | 4743 | CITY-JAPO | ng/Kg | 2.76 U | 2.29 U | 7.06 U | 120.00 | 946.00 | 2.41 U | 2.71 U | 2.71 U | 8.46 U | 35.90 | 89.00 |
| CL Soil | 4749 | CITY-S | ng/Kg | 2.53 U | 1.86 U | 5.37 U | 57.60 | 247.00 | 1.98 U | 2.11 U | 2.11 U | 7.92 | 32.80 | 33.10 |

U=practical quantitation limit

| | | | | | | | | | | | | | | |
|-------------|------|---------------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-----|
| R7 Water | 4744 | FIELD BLANK | pg/L | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 |
| R7 Water | 4746 | BLUE LAKE | pg/L | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 |
| R7 Water | 4746 | BLUE LAKE-DUP | pg/L | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 |
| R7 Leachate | 4267 | HOGF STOR | pg/L | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 |
| R7 Leachate | 4267 | HOGF STOR-DUP | pg/L | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 U | 100 |
| R7 Ash | 4269 | HOGF-BOTTOM | ng/Kg | 10 U | 10 U | 10 U | 10 U | 10 U | 79 U | 26 | 12 | 20 U | 20 U | 10 |
| R7 Soil | 4730 | GALANKIN I | ng/Kg | 10 U | 10 U | 10 U | 10 U | 10 U | 10 U | 10 U | 10 U | 10 U | 10 U | 10 |
| R7 Soil | 4732 | APC-DUP | ng/Kg | 10 U | 10 U | 10 U | 10 U | 210 | 10 U | 10 U | 10 U | 10 U | 10 U | 10 |
| R7 Soil | 4733 | BLUE LAKE | ng/Kg | 10 U | 10 U | 10 U | 10 U | 10 U | 10 U | 10 U | 10 U | 10 U | 10 U | 10 |
| R7 Soil | 4747 | JAMESTOWN | ng/Kg | 10 U | 10 U | 10 U | 150 U | 580 | 10 U | 10 U | 10 U | 10 U | 31 | 12 |
| R7 Soil | 4748 | THIMBLEBERRY | ng/Kg | 10 U | 10 U | 10 U | 170 | 560 | 10 U | 10 U | 10 U | 10 U | 29 | 36 |

U=Detection Limit

| | | | | | | | | | | | | | | |
|-------------|------|---------------|-------|--------|--------|----------|--------|---------|--------|--------|--------|--------|--------|---------|
| DU Tissue | 0179 | BL LK-R TROUT | ng/Kg | 0.67 U | 1.27 U | 1.66 U | 7.12 U | No data | 0.47 U | 1.68 U | 1.68 U | 2.75 U | 5.3 U | Few Oct |
| DU Tissue | 0181 | H COVE-QUILLB | ng/Kg | 0.67 U | 1.27 U | 1.67 U | 7.12 U | No data | 2.36 | 1.68 U | 1.68 U | 2.77 U | 5.32 U | No data |
| DU Tissue | 0183 | SAW B-QUILLB | ng/Kg | 0.67 U | 1.27 U | 1.66 U | 7.12 U | No data | 3.48 | 1.68 U | 1.68 U | 2.75 U | 5.3 U | No data |
| DU Tissue | 0176 | SILV B-SCULP | ng/Kg | 0.67 U | 1.27 U | 1.66 U | 7.29 U | No data | 3.83 | 1.68 U | 1.68 U | 2.75 U | 5.3 U | No data |
| DU Tissue | 0185 | SILV B-E SOLE | ng/Kg | 1.17 | 1.27 U | 1.66 U | 0 U | No data | 1.76 | 1.68 U | 1.68 U | 2.75 U | 5.3 U | No data |
| DU Tissue | 0175 | SILV B-SHRIMP | ng/Kg | 0.67 U | 1.27 U | 1.67 U | 7.12 U | No data | 1.10 | 1.68 U | 1.68 U | 2.76 U | 5.31 U | No data |
| DU Tissue | 0184 | TH B-SANDDAB | ng/Kg | 0.68 U | 1.27 U | 1.67 U | 7.14 U | No data | 1.37 | 1.68 U | 1.68 U | 2.76 U | 5.32 U | No data |
| DU Tissue | 0182 | TH B-QUILLB | ng/Kg | 0.67 U | 1.27 U | 1.67 U | 7.12 U | No data | 2.16 | 1.68 U | 1.68 U | 2.76 U | 5.31 U | No data |
| DU Tissue | 0180 | TH L-E BR TRT | ng/Kg | 0.67 U | 1.27 U | 1.66 U | 7.12 U | No data | 0.47 U | 1.67 U | 1.67 U | 2.75 U | 5.29 U | No data |
| DU Tissue | 0178 | SAW B-CRB HEP | ng/Kg | 0.76 U | 1.27 U | 2.03 U | 8.15 U | 9.6 U | 11.38 | 1.68 U | 1.68 U | 2.76 U | 5.31 U | 6.41 |
| DU Tissue | 0178 | SAW B-CRB GHS | ng/Kg | 0.68 U | 1.28 U | 1.67 U | 7.7 U | 9.6 U | 7.51 | 1.69 U | 1.69 U | 2.77 U | 5.33 U | 6.41 |
| DU Tissue | 0178 | SAW B-CRB MUS | ng/Kg | 0.67 U | 1.27 U | 1.88 U | 7.37 U | 9.6 U | 2.73 | 1.68 U | 1.68 U | 2.76 U | 5.31 U | 6.41 |
| DU Tissue | 0177 | TH B-MUSSELS | ng/Kg | 0.68 U | 1.27 U | 1.67 U | 7.71 U | 2.4 | 3.22 | 1.68 U | 1.68 U | 2.76 U | 5.31 U | 6.41 |
| DU Sediment | 4711 | H COVE | ng/Kg | 1.35 U | 3.09 U | 6.42 U | 54 QR | No data | 4.14 | 3.77 U | 3.38 U | 5.54 U | 20 QR | No data |
| DU Sediment | 4704 | E CHANNEL | ng/Kg | 1.35 U | 3.15 U | 4.12 U | 25 QR | No data | 8.13 | 3.39 U | 3.44 U | 6.23 U | 9 QR | No data |
| DU Sediment | 4710 | SAWMILL COVE | ng/Kg | 1.35 U | 3.39 U | 11 QR | 174 QR | No data | 46.1 | 2.88 | 3.43 | 3.13 | 53 QR | No data |
| DU Sediment | 4712 | BLUE LAKE | ng/Kg | 2.09 U | 3.85 | 16.31 | 28 QR | No data | 3.98 | 4.59 U | 4.06 | 14.54 | 55 QR | No data |
| DU Sediment | 4745 | LANDF-LEACH | ng/Kg | 1.35 U | 2.61 U | 6.58 UQR | 96 QR | No data | 1.34 | 3.38 U | 4.27 U | 7 QR | 99 QR | No data |

| L a b | Type | Sample No. | Descrip TEF-factor | Units | 123478 | 123789 | 1234678 | 123478 | 123678 | 123789 | 234678 | 1234678 | 1234789 |
|-------------|------|---------------|-----------------------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|
| | | | | | HxCDD | HxCDD | HpCDD | HxCDF | HxCDF | HxCDF | HxCDF | HpCDF | HpCDF |
| CL Ash | 4269 | HOGF-BOTTOM | ng/Kg | 5.57U | 6.42* | 18.3* | 6.34U | 6.34U | 6.34U | 6.34U | 6.34U | 14.2* | 5.8U |
| CL Ash | 4270 | MULTI CLON | ng/Kg | 5.36U | 5.36U | 4.42U | 6.49U | 6.49U | 6.49U | 6.49U | 6.49U | 5.62U | 5.62U |
| CL Soil | 4721 | DEEP INLET | ng/Kg | 39.8UJ3 | 39.8UJ3 | 40.8UJ3 | 40UJ3 | 40UJ3 | 40UJ3 | 40UJ3 | 40UJ3 | 39.1UJ3 | 39.1UJ3 |
| CL Soil | 4730 | GALANKIN I | ng/Kg | 35.8U | 8.61U | 8.37U | 10.6U | 10.6U | 10.6U | 10.6U | 10.6U | 9.25U | 9.25U |
| CL Soil | 4731 | APC | ng/Kg | 7.61U | 10.5 | 83.7 | 7.67U | 7.67U | 7.67U | 7.67U | 7.67U | 12.5UJ | 12.5UJ |
| CL Leach | 4740 | SITKA LAND | pg/L | 57.3UJ3 | 57.3UJ3 | 44.3UJ3 | 61.4UJ3 | 61.4UJ3 | 61.4UJ3 | 61.4UJ3 | 61.4UJ3 | 58.1UJ3 | 58.1UJ3 |
| CL Soil | 4741 | CITY-N | ng/Kg | 6.07U | 6.07U | 22.5 | 5.69U | 5.69U | 5.69U | 5.69U | 5.69U | 5.97U | 5.97U |
| CL Soil | 4742 | CITY-C | ng/Kg | 5.45U | 5.45U | 11.2 | 5.79U | 5.79U | 5.79U | 5.79U | 5.79U | 4.92U | 4.92U |
| CL Soil | 4743 | CITY-JAPO | ng/Kg | 7.06U | 7.06U | 120 | 8.46U | 8.46U | 8.46U | 8.46U | 8.46U | 35.9 | 6.89U |
| CL Soil | 4749 | CITY-S | ng/Kg | 5.37U | 5.37U | 57.6 | 6.67U | 6.67U | 6.67U | 6.67U | 7.92 | 32.8 | 6.21U |

ADEC investigation

| L a Type | Sample No. | Descrip TEF-factor | Units | 2378 | 2378 | 2378 | 2378 | OCDD | 2378 | 12378 | 23478 | 2378 | 2378 | OCDF |
|------------------------------------|---------------|-----------------------|-------|-------|-------|-------|-------|-------|-------|--------|--------|-------|-------|-------|
| | | | | TCDD | PeCDD | HxCDD | HpCDD | | TCDF | PeCDF | PeCDF | HxCDF | HpCDF | |
| b | | | | 1.000 | 0.500 | 0.100 | 0.010 | 0.001 | 0.100 | 0.050 | 0.500 | 0.100 | 0.010 | 0.001 |
| U=Detection Limit | | | | | | | | | | | | | | |
| AD Ash | JR190 | FLY ASH - ESP | ng/Kg | 99 | 600 | 6500 | 12000 | 1900 | 1500 | 320 | 230 | 803 | 77 | 290 |
| AD Solid | JR290 | FIELD BLANK | ng/Kg | 2.2 U | 2.9 U | 6.4 U | 12 | 45 | 1.5 U | 0.97 U | 0.78 U | 1.1 U | 4.5 U | 2.6 |
| AD Ash | JR390 | FLY ASH - ESP | ng/Kg | 42 U | 2000 | 16400 | 17000 | 12000 | 3100 | 700 | 540 | 569 | 413 | 70 |
| AD Ash | JR590 | FLY ASH - ESP | ng/Kg | 80 U | 780 | 6800 | 12000 | 5900 | 2100 | 420 | 330 | 831 | 78 | 100 |
| Sample Collected July 15, 1990 | | | | | | | | | | | | | | |
| AD Sludge | DEC05 | SAWMILL COVE | ng/Kg | 3.8 | 3.5 | 22.7 | 57 | 330 | 140 | 2.7 | 3.9 | 6.7 | 4.4 U | 13 |
| Sample Collected October 15, 1990; | | | | | | | | | | | | | | |
| AD Sludge | 0004 | EASTERN CHANL | ng/Kg | 3 | 1.8 U | 4.8 U | 31 | 220 | 61 | 1.5 U | 2 U | 3.3 U | 5 | 13 |
| AD Sludge | 0009 | EASTERN CHANL | ng/Kg | 2.6 | 1.6 U | 7.6 U | 28 | 230 | 84 | 1.5 U | 2.1 U | 2.8 U | 2.5 U | 10 |

C-2. ORGANICS

Below are the definitions for qualifiers used in the GC/MS Organics area when qualifying Semivolatile, Volatile, PAH, Resin Acids, and Guaiacol/Catechol/Phenolics analysis results.

Data Qualifiers

- | | | |
|-----|---|--|
| U | - | The analyte was analyzed for, but not detected. the associated numerical value is the sample quantitation limit. |
| J | - | The analyte was analyzed for, and was positively identified. However, the associated numerical value is an <u>estimate only</u> . |
| REJ | - | The data are <u>unusable</u> for all purposes. The analyte was analyzed for, but the presence or absence of the analyte has not been verified. |
| N | - | There is <u>presumptive</u> evidence the analyte is present. |
| NJ | - | A combination of "N" and "J" qualifiers. There is <u>presumptive evidence</u> that the analyte is present. The associated numerical value is an estimate of the concentration of the analyte in this sample. |
| UJ | - | A combination of "U" and "J" qualifiers. The analyte was analyzed for, and was not present above the level of the associated value. The associated numerical value is an estimate of the quantitation limit of the analyte in this sample. |
| NAR | - | There is <u>no analysis result</u> for this analyte. |
| * | - | The analyte was present in the sample. |

| Sample #: | 90354267 | 90354704 | 90354710 | 90354711 | 90354712 | 90354744 | 90354746 | 90354700 | 90354701 | 90354702 | 90354709 |
|----------------------------------|------------|------------|------------|------------|-----------|------------|-----------|------------|------------|------------|------------|
| MEDIA | Leachate | Sediment | Sediment | Sediment | Sediment | Water | Water-DW | Water-Mar | Water-Mar | Water-Mar | Water-Mar |
| Descrip: | HOG FUEL S | EAST CHANN | SAWMILL CO | HERRING CO | BLUE LAKE | SITKA CATC | BLUE LAKE | HERRING CO | THIMLEBERR | EAST CHANN | SAWMILL CO |
| Matrix: | Water-To | Sediment | Sediment | Sediment | Sediment | Water-To | Water-To | Water-To | Water-To | Water-To | Water-To |
| Units: | ug/l | ug/kg | ug/kg | ug/kg | ug/kg | ug/l | ug/l | ug/l | ug/l | ug/l | ug/l |
| TENTATIVELY IDENTIFIED VOLATILES | | | | | | | | | | | |
| RX 52 1,1,2 Trichlorotrifluoro | | 2.8NJ* | | | | | | | | | |
| RX 52 1H-INDENE, 1-ETHYLIDENE- | | | | | | 0.083NJ* | | | | | |
| RX 52 BENZENE, 1-ETHYL-2-METHY | | | | | | 0.084NJ* | | | | | |
| RX 52 BICYCLO[4.1.0]HEPTANE, 3 | 0.72NJ* | | | | | | | | | | |
| RX 52 CAMPHOR (ACN) | 2.4NJ* | | | | | | | | | | |
| RX 52 CINEOLE (VAN) | 0.51NJ* | | | | | | | | | | |
| RX 52 DECANE, 2,5,6-TRIMETHYL- | | | 75NJ* | | | | | | | | |
| RX 52 Decane, 2,6,6-Trimethyl- | | | 58NJ* | | | | | | | | |
| RX 52 DECANE, 2,6,7-TRIMETHYL- | | | 97NJ* | | | | | | | | |
| RX 52 DISULFIDE, DIMETHYL | | 0.89NJ* | 6.4NJ* | | | | | | | | |
| RX 52 FORMAMIDE, N,N-DIBUTYL- | | | | | | | | | | | |
| RX 52 METHANE, THIOBIS | | 59NJ* | 150NJ* | | | | | | | | |
| RX 52 NAPHTHALENE, 1,4-DIMETHY | | | | | | 0.21NJ* | | | | | |
| BASE/NEUTRALS/ACIDS | | | | | | | | | | | |
| RX 68 1,2,4-Trichlorobenzene | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 1,2-Dichlorobenzene | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 1,3-Dichlorobenzene | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 1,4-Dichlorobenzene | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 2,4,5-Trichlorophenol | 18UJ | 3100U | 12000U | 14000U | 5000U | 42UJ | 42UJ | 9UJ | 12UJ | 10UJ | 15UJ |
| RX 68 2,4,6-Trichlorophenol | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 2,4-Dichlorophenol | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 2,4-Dimethylphenol | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 2,4-Dinitrophenol | 18UJ | 3100UJ | 12000UJ | 14000UJ | 5000UJ | 42UJ | 42UJ | 9UJ | 12UJ | 10UJ | 15UJ |
| RX 68 2,4-Dinitrotoluene | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | REJ | REJ | REJ |
| RX 68 2,6-Dinitrotoluene | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 2-Chloronaphthalene | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 2-Methylnaphthalene | 4UJ | 30J* | 610J* | 83J* | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 2-Methylphenol | 52J* | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 2-Nitroaniline | 18UJ | 3100U | 12000U | 14000U | 5000U | 42UJ | 42UJ | 9UJ | 12UJ | 10UJ | 15UJ |
| RX 68 2-Nitrophenol | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 3,3'-Dichlorobenzidine | REJ | REJ | REJ | REJ | REJ | REJ | REJ | 2UJ | REJ | REJ | REJ |
| RX 68 3-Nitroaniline | 18UJ | 3100UJ | 12000UJ | 14000UJ | 5000UJ | 42UJ | 42UJ | 9UJ | 12UJ | 10UJ | 15UJ |
| RX 68 4,6-Dinitro-2-methylphen | 18UJ | 3100U | 12000U | 14000U | 5000U | 42UJ | 42UJ | 9UJ | 12UJ | 10UJ | 15UJ |
| RX 68 4-Bromophenyl-phenylethe | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 4-Chloroaniline | REJ | REJ | REJ | REJ | REJ | REJ | REJ | REJ | REJ | REJ | REJ |
| RX 68 4-Chlorophenyl-phenyleth | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 4-Chloro-3-Methylphenol | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 4-Methylphenol | 340J* | 4400* | 180000* | 540J* | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 4-Nitroaniline | 18UJ | 3100U | 12000U | 14000U | 5000U | 42UJ | 42UJ | 9UJ | 12UJ | 10UJ | 15UJ |
| RX 68 4-Nitrophenol | 18UJ | 3100U | 12000U | 14000U | 5000U | 42UJ | 42UJ | 9UJ | 12UJ | 10UJ | 15UJ |
| RX 68 Acenaphthene | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Acenaphthylene | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Anthracene | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Benzoic acid | 18UJ | 880J* | 12000J* | 14000UJ | 100J* | 42UJ | 42UJ | 9UJ | 12UJ | 10UJ | 15UJ |
| RX 68 Benzo(a)anthracene | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Benzo(a)pyrene | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |

| Sample #: | 90354267 | 90354704 | 90354710 | 90354711 | 90354712 | 90354744 | 90354746 | 90354700 | 90354701 | 90354702 | 90354709 |
|---------------------------------------|------------|------------|------------|------------|-----------|------------|-----------|------------|------------|------------|------------|
| MEDIA | Leachate | Sediment | Sediment | Sediment | Sediment | Water | Water-DW | Water-Mar | Water-Mar | Water-Mar | Water-Mar |
| Descrip: | HOG FUEL S | EAST CHANN | SAWMILL CO | HERRING CO | BLUE LAKE | SITKA CATC | BLUE LAKE | HERRING CO | THIMLEBERR | EAST CHANN | SAWMILL CO |
| Matrix: | Water-To | Sediment | Sediment | Sediment | Sediment | Water-To | Water-To | Water-To | Water-To | Water-To | Water-To |
| Units: | ug/l | ug/kg | ug/kg | ug/kg | ug/kg | ug/l | ug/l | ug/l | ug/l | ug/l | ug/l |
| RX 68 Benzo(b)fluoranthene | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Benzo(ghi)perylene | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Benzo(k)fluoranthene | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Benzyl Alcohol | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | REJ | REJ | REJ | REJ |
| RX 68 bis(2-Chloroethoxy)Metha | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 bis(2-Chloroethyl)Ether | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 bis(2-Chloroisopropyl)Et | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 BIS(2-ETHYLHEXYL) PHTHAL | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 0.4J* | 0.2J* | 0.4J* | 0.1J* |
| RX 68 Butylbenzylphthalate | 4U | 650U | 2500U | 2900U | 1000U | 8UJ | 0.7J* | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Carbazole | 4UJ | 650UJ | 2500UJ | 2900UJ | 1000UJ | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Chrysene | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Dibenzo(furan | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Dibenzo(a,h)anthracene | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Diethylphthalate | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Dimethylphthalate | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Di-n-Butylphthalate | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Di-n-Octyl Phthalate | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Fluoranthene | 4UJ | 650U | 2300J* | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Fluorene | 4UJ | 650U | 570J* | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Hexachlorobenzene | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Hexachlorobutadiene | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Hexachlorocyclopentadien | 7UJ | 1300U | 5000U | 5700U | 2100U | 17UJ | 17UJ | 4UJ | 5UJ | 4UJ | 6UJ |
| RX 68 Hexachloroethane | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Indeno(1,2,3-cd)pyrene | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Isophorone | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Naphthalene | 4UJ | 27J* | 790J* | 76J* | 1000U | 8UJ | 8UJ | 2UJ | 0.09J* | 2UJ | 0.1J* |
| RX 68 Naphthalene, 1-Methyl- | 4UJ | 33J* | 800J* | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Nitrobenzene | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 N-Nitrosodiphenylamine | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 2J* | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 N-Nitroso-di-n-Propylami | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 o-Chlorophenol | 4UJ | 650U | 2500U | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Pentachlorophenol | 3J* | 3100U | 12000U | 14000U | 5000U | 42UJ | 42UJ | 9UJ | 12UJ | 10UJ | 15UJ |
| RX 68 Phenanthrene | 4UJ | 650U | 2600* | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Phenol | 58J* | 310J* | 13000* | 2900U | 1000U | 2J* | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Pyrene | 4UJ | 650U | 1600J* | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 Retene | 4UJ | 170J* | 2500J* | 2900U | 1000U | 8UJ | 8UJ | 2UJ | 2UJ | 2UJ | 3UJ |
| RX 68 %RECOV:PYRENE-D10 (SS) | | 92 | 83 | 70 | 96 | 101 | 148 | 122 | 131 | 109 | 115 |
| RX 68 %RECOV:Surrog: 2-Fluorobiphenyl | | 78 | 68 | 72 | 78 | 67 | 54 | 80 | 27 | 19 | 46 |
| RX 68 %RECOV:Surrog: 2-Fluorophenol | | 54 | 54 | 70 | 73 | 68 | 103 | 93 | 69 | 85 | 68 |
| RX 68 %RECOV:Surrog: D14-Terphenyl | | 61 | 89 | 76 | 101 | 106 | 158 | 130 | 140 | 122 | 125 |
| RX 68 %RECOV:Surrog: D5-Nitrobenzene | | 91 | 50 | 84 | 68 | 68 | 78 | 112 | 60 | 59 | 56 |
| RX 68 %RECOV:Surrog: D5-Phenol | | 44 | 40 | 55 | 40 | 50 | 45 | 55 | 26 | 24 | 21 |

TENTATIVELY IDENTIFIED BASE/NEUTRALS/ACIDS

| | | |
|--------------------------------|--------|---------|
| RX 6A 1H-INDOLE-2,3-DIONE | | 6400NJ* |
| RX 6A 1-PHENANTHRENECARBOXALDE | 9.2NJ* | |
| RX 6A 2,4-PENTANEDIONE, 3-METH | | |
| RX 6A 2H-1-BENZOPYRAN, 3,4-DIH | 78NJ* | |

| Sample #: | 90354267 | 90354704 | 90354710 | 90354711 | 90354712 | 90354744 | 90354746 | 90354700 | 90354701 | 90354702 | 90354709 |
|--|------------|------------|------------|------------|-----------|------------|-----------|------------|------------|------------|------------|
| MEDIA | Leachate | Sediment | Sediment | Sediment | Sediment | Water | Water-DW | Water-Mar | Water-Mar | Water-Mar | Water-Mar |
| Descrip: | HOG FUEL S | EAST CHANN | SAWMILL CO | HERRING CO | BLUE LAKE | SITKA CATC | BLUE LAKE | HERRING CO | THIMLEBERR | EAST CHANN | SAWMILL CO |
| Matrix: | Water-To | Sediment | Sediment | Sediment | Sediment | Water-To | Water-To | Water-To | Water-To | Water-To | Water-To |
| Units: | ug/l | ug/kg | ug/kg | ug/kg | ug/kg | ug/l | ug/l | ug/l | ug/l | ug/l | ug/l |
| PESTICIDES/PCBs | | | | | | | | | | | |
| RX 71 4,4'-DDD | 0.008UJ | 6U | 40U | 25U | 9U | 0.02U | 0.011U | 0.008U | 0.01U | 0.008U | 0.012U |
| RX 71 4,4'-DDE | 0.008UJ | 6U | 40U | 25U | 9U | 0.02U | 0.011U | 0.008U | 0.01U | 0.008U | 0.012U |
| RX 71 4,4'-DDT | 0.016UJ | 6U | 60UJ | 25U | 9U | 0.02U | 0.011U | 0.008U | 0.01U | 0.008U | 0.012U |
| RX 71 Aldrin | 0.008UJ | 6U | 40U | 25U | 9U | 0.02U | 0.011U | 0.008U | 0.01U | 0.008U | 0.012U |
| RX 71 alpha-BHC | 0.016UJ | 6U | 20U | 25U | 9U | 0.02U | 0.011U | 0.008U | 0.01U | 0.008U | 0.012U |
| RX 71 beta-BHC | 0.016UJ | 6U | 40U | 25U | 9U | 0.02U | 0.011U | 0.008U | 0.01U | 0.008U | 0.012U |
| RX 71 Chlordane (Tech) | 0.08UJ | 60U | 400U | 250U | 90U | 0.2U | 0.11U | 0.08U | 0.1U | 0.08U | 0.12U |
| RX 71 delta-BHC | 0.016UJ | 6U | 40U | 25U | 9U | 0.02U | 0.011U | 0.008U | 0.01U | 0.008U | 0.012U |
| RX 71 Dieldrin | 0.008UJ | 6U | 40U | 25U | 9U | 0.02U | 0.011U | 0.008U | 0.01U | 0.008U | 0.012U |
| RX 71 Endosulfan I | 0.016UJ | 6U | 40U | 25U | 9U | 0.02U | 0.011U | 0.008U | 0.01U | 0.008U | 0.012U |
| RX 71 Endosulfan II | 0.008UJ | 6U | 40U | 25U | 9U | 0.02U | 0.011U | 0.008U | 0.01U | 0.008U | 0.012U |
| RX 71 Endosulfan sulfate | 0.016UJ | 6U | 80U | 25U | 9U | 0.02U | 0.011U | 0.008U | 0.01U | 0.008U | 0.012U |
| RX 71 Endrin | 0.016UJ | 6U | 20U | 25U | 9U | 0.02U | 0.011U | 0.008U | 0.01U | 0.008U | 0.012U |
| RX 71 Endrin aldehyde | 0.008UJ | 6U | 40U | 25U | 9U | 0.02U | 0.011U | 0.008U | 0.01U | 0.008U | 0.012U |
| RX 71 Endrin Ketone | 0.016UJ | 12U | 80U | 50U | 18U | | | | | | |
| RX 71 gamma-BHC (Lindane) | 0.008UJ | 6U | 40U | 25U | 9U | 0.02U | 0.011U | 0.008U | 0.01U | 0.008U | 0.012U |
| RX 71 Heptachlor | 0.016UJ | 6U | 40U | 25U | 9U | 0.02U | 0.011U | 0.008U | 0.01U | 0.008U | 0.012U |
| RX 71 Heptachlor Epoxide | 0.016UJ | 6U | 20U | 25U | 9U | 0.02U | 0.011U | 0.008U | 0.01U | 0.008U | 0.012U |
| RX 71 Methoxychlor | 0.016UJ | 12U | 80U | 50U | 18U | 0.04U | 0.022U | 0.015U | 0.02U | 0.016U | 0.024U |
| RX 71 PCB - 1016 | 0.3UJ | 60U | 820U | 250U | 90U | 0.2U | 0.11U | 0.08U | 0.1U | 0.08U | 0.12U |
| RX 71 PCB - 1221 | 0.3UJ | 60U | 820UJ | 250U | 90U | 0.2U | 0.11U | 0.08U | 0.1U | 0.08U | 0.12U |
| RX 71 PCB - 1232 | 0.3UJ | 60U | 820U | 250U | 90U | 0.2U | 0.11U | 0.08U | 0.1U | 0.08U | 0.12U |
| RX 71 PCB - 1242 | 0.3UJ | 60U | 820U | 250U | 90U | 0.2U | 0.11U | 0.08U | 0.1U | 0.08U | 0.12U |
| RX 71 PCB - 1248 | 0.3UJ | 60U | 820U | 250U | 90U | 0.2U | 0.11U | 0.08U | 0.1U | 0.08U | 0.12U |
| RX 71 PCB - 1254 | 0.3UJ | 60U | 820U | 250U | 90U | 0.2U | 0.11U | 0.08U | 0.1U | 0.08U | 0.12U |
| RX 71 PCB - 1260 | 0.5UJ | 60U | 820U | 250U | 90U | 0.2U | 0.11U | 0.08U | 0.1U | 0.08U | 0.12U |
| RX 71 Toxaphene | 0.45UJ | 160U | 2000U | 700U | 250U | 0.5U | 0.33U | 0.25U | 0.3U | 0.25U | 0.35U |
| RX 71 %RECOV:4,4-Dibromooctafluorobip | 42 | 66 | 78 | 44 | 39 | 41 | 53 | 37 | 46 | 48 | 51 |
| RX 71 %RECOV:DIBUTYLCHLORENDATE (SS) 17J | | 80 | 69 | 58 | 62 | 79 | 88 | 77 | 79 | 73 | 64 |
| RX 71 %RECOV:OCTACHLORONAPHTHALENE (S | 24 | 8J | 7J | 2U | 15 | 71 | 77 | 75 | 85 | 70 | 68 |
| VOLATILES | | | | | | | | | | | |
| RX 51 1,1,1-Trichloroethane | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 1,1,2-Trichloroethane | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 1,1-Dichloroethane | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 1,1-Dichloroethene | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 1,1-Dichloropropene | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 1,2,3-Trichlorobenzene | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 1,2,3-Trichloropropane | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 1,2,4-Trichlorobenzene | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 1,2,4-Trimethylbenzene | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 1,2-Dibromoethane (EDB) | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 1,2-Dichlorobenzene | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 1,2-Dichloroethane | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 1,2-Dichloropropane | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 1,3,5-Trimethylbenzene | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |

| Sample #: | 90354267 | 90354704 | 90354710 | 90354711 | 90354712 | 90354744 | 90354746 | 90354700 | 90354701 | 90354702 | 90354709 |
|--------------------------------|------------|------------|------------|------------|-----------|------------|-----------|------------|------------|------------|------------|
| MEDIA | Leachate | Sediment | Sediment | Sediment | Sediment | Water | Water-DW | Water-Mar | Water-Mar | Water-Mar | Water-Mar |
| Descrip: | HOG FUEL S | EAST CHANN | SAWMILL CO | HERRING CO | BLUE LAKE | SITKA CATC | BLUE LAKE | HERRING CO | THIMLEBERR | EAST CHANN | SAWMILL CO |
| Matrix: | Water-To | Sediment | Sediment | Sediment | Sediment | Water-To | Water-To | Water-To | Water-To | Water-To | Water-To |
| Units: | ug/l | ug/kg | ug/kg | ug/kg | ug/kg | ug/l | ug/l | ug/l | ug/l | ug/l | ug/l |
| RX 51 1,3-Dichlorobenzene | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 1,3-Dichloropropane | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 1,4-Dichlorobenzene | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 2,2-Dichloropropane | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 2-Butanone | 9J* | 4UJ | 18J* | 31UJ | 64J* | 2U | 1U | 1UJ | | | |
| RX 51 2-Chlorotoluene | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 2-Hexanone | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 4-Chlorotoluene | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 4-Methyl-2-Pentanone | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Acetone | 10J* | 9UJ | 52UJ | 550J* | 320J* | 6* | 1U | 1UJ | | | |
| RX 51 Benzene | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Benzene, 1,2-Dimethyl | | 4UJ | 17UJ | 11UJ | 6UJ | | | | | | |
| RX 51 BENZENE, ETHENYL-(STYREN | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 BENZENE, ETHYL- | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 BENZENE, PROPYL- | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Bromobenzene | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Bromochloromethane | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Bromodichloromethane | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Bromoform | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Bromomethane | 1UJ | 4U | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Butylbenzene | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Carbon Disulfide | 1UJ | 4UJ | 70J* | 11J* | 1J* | 1U | 1U | 1UJ | | | |
| RX 51 Carbon Tetrachloride | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Chlorobenzene | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Chloroethane | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Chloroform | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Chloromethane | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Cis-1,2-Dichloroethene | 0.5J* | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 cis-1,3-Dichloropropene | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 d8-Toluene | | | | | | | | | | | |
| RX 51 DBCP | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Dibromochloromethane | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Dibromomethane | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Ethane, 1,1,1,2-Tetrachl | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 ETHANE, 1,1,2,2-TETRACHL | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Hexachlorobutadiene | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Isopropylbenzene (Cumene | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Methane, Dichlorodifluor | 1UJ | REJ | REJ | REJ | REJ | 1U | 1U | 1UJ | | | |
| RX 51 Methylene Chloride | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Naphthalene | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 p-Isopropyltoluene | 4UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Sec-Butylbenzene | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Tert-Butylbenzene | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Tetrachloroethene | 0.03J* | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Toluene | 670J* | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Total Xylenes | 1UJ | | | | | 1U | 1U | 1UJ | | | |
| RX 51 trans-1,2-Dichloroethene | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 trans-1,3-Dichloropropen | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |

Organics analysis I. - Page 6

| Sample #: | 90354267 | 90354704 | 90354710 | 90354711 | 90354712 | 90354744 | 90354746 | 90354700 | 90354701 | 90354702 | 90354709 |
|---------------------------------------|------------|------------|------------|------------|-----------|------------|-----------|------------|------------|------------|------------|
| MEDIA | Leachate | Sediment | Sediment | Sediment | Sediment | Water | Water-DW | Water-Mar | Water-Mar | Water-Mar | Water-Mar |
| Descrip: | HOG FUEL S | EAST CHANN | SAWMILL CO | HERRING CO | BLUE LAKE | SITKA CATC | BLUE LAKE | HERRING CO | THIMLEBERR | EAST CHANN | SAWMILL CO |
| Matrix: | Water-To | Sediment | Sediment | Sediment | Sediment | Water-To | Water-To | Water-To | Water-To | Water-To | Water-To |
| Units: | ug/l | ug/kg | ug/kg | ug/kg | ug/kg | ug/l | ug/l | ug/l | ug/l | ug/l | ug/l |
| RX 51 Trichloroethene | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 1U | 1U | 1UJ | | | |
| RX 51 Trichlorofluoromethane | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 0.5J* | 1U | 1UJ | | | |
| RX 51 Vinyl Acetate | REJ | REJ | REJ | REJ | REJ | REJ | REJ | REJ | | | |
| RX 51 Vinyl Chloride | 1UJ | 4UJ | 17UJ | 11UJ | 6UJ | 0.1J* | 1U | 1UJ | | | |
| RX 51 %RECOV:1,2-Dichloroethane-d4 (| 80 | 99 | 97 | 124 | 93 | 96 | 88 | 124 | | | |
| RX 51 %RECOV:d8-Toluene | 94 | 101 | 104 | 104 | 96 | 104 | 94 | 103 | | | |
| RX 51 %RECOV:p-Bromofluorobenzene | 92 | 77 | 75 | 79 | 92 | 106 | 96 | 99 | | | |
| RX 51 %RECOV:Surrog: 1-Bromo-2-Fluoro | 82 | 122 | 119 | 109 | 104 | 96 | 92 | 107 | | | |

| Sample #: | 90354269 | 90354740 |
|----------------------------------|------------|------------|
| MEDIA | Ash | Leachate |
| Descrip: | BOTTOM ASH | SITKA LAND |
| Matrix: | Sediment | Water-To |
| Units: | ug/kg | ug/l |
| TENTATIVELY IDENTIFIED VOLATILES | | |
| RX 52 1,1,2 Trichlorotrifluoro | 9.0NJ* | |
| RX 52 1H-INDENE, 1-ETHYLIDENE- | | 3.0NJ* |
| RX 52 1H-INDENE, 2,3-DIHYDRO-1 | | 0.38NJ* |
| RX 52 1H-INDENE, 2,3-DIHYDRO-1 | | 0.64NJ* |
| RX 52 1H-INDENE, 2,3-DIHYDRO-1 | | 0.67NJ* |
| RX 52 1H-INDENE, 2,3-DIHYDRO-4 | | 1.1NJ* |
| RX 52 1-PROPANOL, 2,2-DIMETHYL | | |
| RX 52 2-PENTANONE, 3-METHYL- | | |
| RX 52 3-PENTANONE, 2,4-DIMETHY | | 1.5NJ* |
| RX 52 ACETIC ACID, 1-METHYLETH | | |
| RX 52 ACETIC ACID, PROPYL ESTE | | |
| RX 52 BENZENE, 1,2,3-TRIMETHYL | | 5.6NJ* |
| RX 52 BENZENE, 1-ETHENYL-2-MET | | 4.6NJ* |
| RX 52 BENZENE, 1-ETHYL-2,3-DIM | | 0.64NJ* |
| RX 52 BENZENE, 1-ETHYL-2-METHY | | 5.6NJ* |
| RX 52 BENZENE, 1-ETHYL-4-METHY | | 4.2NJ* |
| RX 52 BENZENE, 1-PROPYNYL- | | 1.8NJ* |
| RX 52 BENZENE, 2-ETHYL-1,4-DIM | | 1.8NJ* |
| RX 52 BENZENE, PENTAMETHYL- | | 0.17NJ* |
| RX 52 BENZENE, (1-METHYL-1-PRO | | 2.8NJ* |
| RX 52 BUTANOIC ACID, 1-METHYLE | | |
| RX 52 CARBON OXIDE SULFIDE (CO | | |
| RX 52 Methane, Dimethoxy- | | 0.79NJ* |
| RX 52 NAPHTHALENE, 1,2,3,4-TET | | 0.94NJ* |
| RX 52 Naphthalene, 1-Methyl- | | 2.1NJ* |
| RX 52 p-Isopropyltoluene | | 1.7NJ* |
| RX 52 Tert-Butylbenzene | 4.1NJ* | |
| RX 52 Tetrahydrofuran | | 3.5NJ* |
| RX 52 Thiophene, Benzo[b]- | | 0.47NJ* |
| BASE/NEUTRALS/ACIDS | | |
| RX 68 1,2,4-Trichlorobenzene | 490U | 4UJ |
| RX 68 1,2-Dichlorobenzene | 490U | 0.3J* |
| RX 68 1,3-Dichlorobenzene | 490U | 4UJ |
| RX 68 1,4-Dichlorobenzene | 490U | 2J* |
| RX 68 2,4,5-Trichlorophenol | 2400U | 18UJ |
| RX 68 2,4,6-Trichlorophenol | 490U | 4UJ |
| RX 68 2,4-Dichlorophenol | 490U | 4UJ |
| RX 68 2,4-Dimethylphenol | 490U | 4UJ |
| RX 68 2,4-Dinitrophenol | 2400UJ | 18UJ |
| RX 68 2,4-Dinitrotoluene | 490U | 4UJ |
| RX 68 2,6-Dinitrotoluene | 490U | 4UJ |
| RX 68 2-Chloronaphthalene | 490U | 4UJ |
| RX 68 2-Methylnaphthalene | 490U | 1J* |
| RX 68 2-Methylphenol | 490U | 4UJ |
| RX 68 2-Nitroaniline | 2400U | 18UJ |
| RX 68 2-Nitrophenol | 490U | 4UJ |
| RX 68 3,3'-Dichlorobenzidine | REJ | REJ |
| RX 68 3-Nitroaniline | 2400UJ | 18UJ |
| RX 68 4,6-Dinitro-2-methylphen | 2400U | 18UJ |
| RX 68 4-Bromophenyl-phenylethe | 490U | 4UJ |
| RX 68 4-Chloroaniline | REJ | REJ |
| RX 68 4-Chlorophenyl-phenyleth | 490U | 4UJ |
| RX 68 4-Chloro-3-Methylphenol | 490U | 4UJ |
| RX 68 4-Methylphenol | 10J* | 4UJ |
| RX 68 4-Nitroaniline | 2400U | 18UJ |
| RX 68 4-Nitrophenol | 2400U | 18UJ |
| RX 68 Acenaphthene | 490U | 2J* |
| RX 68 Acenaphthylene | 490U | 4UJ |
| RX 68 Anthracene | 490U | 4UJ |
| RX 68 Benzoic acid | 51J* | 18UJ |
| RX 68 Benzo(a)anthracene | 490U | 4UJ |
| RX 68 Benzo(a)pyrene | 490U | 4UJ |
| RX 68 Benzo(b)fluoranthene | 25J* | 0.1J* |
| RX 68 Benzo(ghi)perylene | 490U | 4UJ |

| Sample #: | | 90354269 | 90354740 |
|--|---------------------------------|------------|------------|
| MEDIA | | Ash | Leachate |
| Descrip: | | BOTTOM ASH | SITKA LAND |
| Matrix: | | Sediment | Water-To |
| Units: | | ug/kg | ug/l |
| RX 68 | Benzo(k)fluoranthene | 490U | 0.2J* |
| RX 68 | Benzyl Alcohol | 490U | 4UJ |
| RX 68 | bis(2-Chloroethoxy)Metha | 490U | 4UJ |
| RX 68 | bis(2-Chloroethyl)Ether | 490U | 4UJ |
| RX 68 | bis(2-Chloroisopropyl)Et | 490U | 4UJ |
| RX 68 | BIS(2-ETHYLHEXYL) PHTHAL | 490U | 0.5J* |
| RX 68 | Butylbenzylphthalate | 490U | 0.7J* |
| RX 68 | Carbazole | 490UJ | 1J* |
| RX 68 | Chrysene | 17J* | 0.3J* |
| RX 68 | Dibenzofuran | 490U | 1J* |
| RX 68 | Dibenzo(a,h)anthracene | 490U | 4UJ |
| RX 68 | Diethylphthalate | 490U | 2J* |
| RX 68 | Dimethylphthalate | 490U | 4UJ |
| RX 68 | Di-n-Butylphthalate | 800U | 0.2J* |
| RX 68 | Di-n-Octyl Phthalate | 490U | 0.2J* |
| RX 68 | Fluoranthene | 490U | 0.2J* |
| RX 68 | Fluorene | 490U | 1J* |
| RX 68 | Hexachlorobenzene | 490U | 4UJ |
| RX 68 | Hexachlorobutadiene | 490U | 4UJ |
| RX 68 | Hexachlorocyclopentadien | 980U | 7UJ |
| RX 68 | Hexachloroethane | 490U | 4UJ |
| RX 68 | Indeno(1,2,3-cd)pyrene | 490U | 4UJ |
| RX 68 | Isophorone | 490U | 4UJ |
| RX 68 | m-Cresol | | |
| RX 68 | Naphthalene | 490U | 6J* |
| RX 68 | Naphthalene, 1-Methyl- | 490U | 3J* |
| RX 68 | Nitrobenzene | 490U | 4UJ |
| RX 68 | N-Nitrosodiphenylamine | 490U | 4UJ |
| RX 68 | N-Nitroso-di-n-Propylami | 490U | 4UJ |
| RX 68 | o-Chlorophenol | 490U | 4UJ |
| RX 68 | Pentachlorophenol | 2400U | 18UJ |
| RX 68 | Phenanthrene | 490U | 1J* |
| RX 68 | Phenol | 490U | 4UJ |
| RX 68 | Pyrene | 490U | 0.3J* |
| RX 68 | Retene | 490U | 0.3J* |
| RX 68 | %RECOV:PYRENE-D10 (SS) | 80 | 97 |
| RX 68 | %RECOV:Surrog: 2-Fluorobiphenyl | 60 | 67 |
| RX 68 | %RECOV:Surrog: 2-Fluorophenol | 55 | 68 |
| RX 68 | %RECOV:Surrog: D14-Terphenyl | 85 | 108 |
| RX 68 | %RECOV:Surrog: D5-Nitrobenzene | 55 | 79 |
| RX 68 | %RECOV:Surrog: D5-Phenol | 50 | 35 |
| TENTATIVELY IDENTIFIED BASE/NEUTRALS/ACIDS | | | |
| RX 6A | 1,1'-BIPHENYL | | 0.54NJ* |
| RX 6A | 1,2,4-Trimethylbenzene | | 3.6NJ* |
| RX 6A | 1,3,5-Trimethylbenzene | | 1.8NJ* |
| RX 6A | 1H-INDENE, 2,3-DIHYDRO-4 | | 0.95NJ* |
| RX 6A | BENZAMIDE, N,N-DIETHYL-3 | | 9.9NJ* |
| RX 6A | Benzene, 1,2,3,4-Tetrame | | 0.64NJ* |
| RX 6A | BENZENE, 1,2,3-TRIMETHYL | | 5.6NJ* |
| RX 6A | BENZENE, 1,2,4,5-TETRA | | 0.48NJ* |
| RX 6A | Benzene, 1,2-Dimethyl | | 5.2NJ* |
| RX 6A | BENZENE, 1-ETHENYL-2-MET | | 2.2NJ* |
| RX 6A | BENZENE, 1-ETHYL-2-METHY | | 3.2NJ* |
| RX 6A | BENZENE, ETHYL- | | 13NJ* |
| RX 6A | Benzoic acid | | |
| RX 6A | CAMPOR (ACN) | | 1.4NJ* |
| RX 6A | Chlorobenzene | | 4.6NJ* |
| RX 6A | ETHANOL, 2-BUTOXY-, PHOS | | 2.7NJ* |
| RX 6A | ETHANONE, 1-(METHYLPHENY | | 0.68NJ* |
| RX 6A | m-Xylene | | 13NJ* |
| RX 6A | PHOSPHORIC ACID TRIBUTYL | | 2.6NJ* |
| PESTICIDES/PCBs | | | |
| RX 71 | 4,4'-DDD | 5U | 0.008U |
| RX 71 | 4,4'-DDE | 5U | 0.008U |

| | | Sample #: | 90354269 | 90354740 |
|----|----|---------------------------------|------------|------------|
| | | MEDIA | Ash | Leachate |
| | | Descrip: | BOTTOM ASH | SITKA LAND |
| | | Matrix: | Sediment | Water-To |
| | | Units: | ug/kg | ug/l |
| RX | 71 | 4,4'-DDT | 5U | 0.008U |
| RX | 71 | Aldrin | 5U | 0.008U |
| RX | 71 | alpha-BHC | 5U | 0.008U |
| RX | 71 | beta-BHC | 5U | 0.008U |
| RX | 71 | Chlordane (Tech) | 50U | 0.08U |
| RX | 71 | delta-BHC | 5U | 0.008U |
| RX | 71 | Dieldrin | 5U | 0.0185* |
| RX | 71 | Endosulfan I | 5U | 0.008U |
| RX | 71 | Endosulfan II | 5U | 0.008U |
| RX | 71 | Endosulfan sulfate | 8U | 0.008U |
| RX | 71 | Endrin | 5U | 0.008U |
| RX | 71 | Endrin aldehyde | 5U | 0.008U |
| RX | 71 | Endrin Ketone | 8U | |
| RX | 71 | gamma-BHC (Lindane) | 5U | 0.008U |
| RX | 71 | Heptachlor | 5U | 0.008U |
| RX | 71 | Heptachlor Epoxide | 5U | 0.008U |
| RX | 71 | Methoxychlor | 8U | 0.016U |
| RX | 71 | PCB - 1016 | 50U | 0.08U |
| RX | 71 | PCB 1221 | 50U | 0.08U |
| RX | 71 | PCB - 1232 | 50U | 0.08U |
| RX | 71 | PCB - 1242 | 50U | 0.08U |
| RX | 71 | PCB - 1248 | 50U | 0.08U |
| RX | 71 | PCB 1254 | 50U | 0.08U |
| RX | 71 | PCB - 1260 | 50U | 0.08U |
| RX | 71 | Toxaphene | 150U | 0.25U |
| RX | 71 | %RECOV:4,4-Dibromooctafluorobip | 54 | 60 |
| RX | 71 | %RECOV:DIBUTYLCHLORENDATE (SS) | 86 | 68 |
| RX | 71 | %RECOV:OCTACHLORONAPHTHALENE (S | 31 | 69 |

PESTICIDES

| | | |
|----|----|--------------------------|
| RX | 72 | 4,4-Dibromooctafluorobip |
| RX | 72 | Chlordane (Tech) |
| RX | 72 | DECACHLOROBIPHENYL |
| RX | 72 | DIBUTYLCHLORENDATE (SS) |
| RX | 72 | Endrin |
| RX | 72 | gamma-BHC (Lindane) |
| RX | 72 | Heptachlor |
| RX | 72 | Heptachlor Epoxide |
| RX | 72 | Methoxychlor |
| RX | 72 | OCTACHLORONAPHTHALENE (S |
| RX | 72 | Toxaphene |
| RX | 72 | TRANS-CHLORDANE (GAMMA) |

HERBICIDES

| | | |
|----|----|--------------------------|
| RX | 73 | 2,4,5-TP (Silvex) |
| RX | 73 | 2,4-D |
| RX | 73 | Surrog: 2,4,6-Tribromoph |

VOLATILES

| | | | | |
|----|----|-------------------------|------|-------|
| RX | 51 | 1,1,1-Trichloroethane | 4J* | 1U |
| RX | 51 | 1,1,2-Trichloroethane | 4UJ | 1U |
| RX | 51 | 1,1-Dichloroethane | 4UJ | 1* |
| RX | 51 | 1,1-Dichloroethene | 3J* | 1U |
| RX | 51 | 1,1-Dichloropropene | 4UJ | 1U |
| RX | 51 | 1,2,3-Trichlorobenzene | 4UJ | 1U |
| RX | 51 | 1,2,3-Trichloropropane | 4UJ | 1U |
| RX | 51 | 1,2,4-Trichlorobenzene | 4UJ | 1U |
| RX | 51 | 1,2,4-Trimethylbenzene | 2J* | 10* |
| RX | 51 | 1,2-Dibromoethane (EDB) | 4UJ | 1U |
| RX | 51 | 1,2-Dichlorobenzene | 4UJ | 0.5J* |
| RX | 51 | 1,2-Dichloroethane | 4UJ | 1U |
| RX | 51 | 1,2-Dichloropropane | 4UJ | 1U |
| RX | 51 | 1,3,5-Trimethylbenzene | 24J* | 3* |
| RX | 51 | 1,3-Dichlorobenzene | 4UJ | 1U |
| RX | 51 | 1,3-Dichloropropane | 4UJ | 1U |
| RX | 51 | 1,4-Dichlorobenzene | 4UJ | 3* |

| | | Sample #: | 90354269 | 90354740 |
|----|----|---------------------------------|------------|------------|
| | | MEDIA | Ash | Leachate |
| | | Descrip: | BOTTOM ASH | SITKA LAND |
| | | Matrix: | Sediment | Water-To |
| | | Units: | ug/kg | ug/l |
| RX | 51 | 2,2-Dichloropropane | 4UJ | 1U |
| RX | 51 | 2-Butanone | 10UJ | 1U |
| RX | 51 | 2-Chlorotoluene | 4UJ | 1U |
| RX | 51 | 2-Hexanone | 4UJ | 1U |
| RX | 51 | 4-Chlorotoluene | 4UJ | 1U |
| RX | 51 | 4-Methyl-2-Pentanone | 4UJ | 1U |
| RX | 51 | Acetone | 88J* | 2* |
| RX | 51 | Benzene | 40J* | 10* |
| RX | 51 | Benzene, 1,2-Dimethyl | | 25* |
| RX | 51 | BENZENE, ETHENYL-(STYREN | 4UJ | 1* |
| RX | 51 | BENZENE, ETHYL- | 4UJ | 25* |
| RX | 51 | BENZENE, PROPYL- | 4UJ | 2* |
| RX | 51 | Bromobenzene | 4UJ | 1U |
| RX | 51 | Bromochloromethane | 4UJ | 1U |
| RX | 51 | Bromodichloromethane | 4UJ | 1U |
| RX | 51 | Bromoform | 4UJ | 1U |
| RX | 51 | Bromomethane | 4UJ | 1U |
| RX | 51 | Butylbenzene | 4UJ | 1U |
| RX | 51 | Carbon Disulfide | 10J* | 1U |
| RX | 51 | Carbon Tetrachloride | 4UJ | 1U |
| RX | 51 | Chlorobenzene | 4UJ | 7* |
| RX | 51 | Chloroethane | 2J* | 5* |
| RX | 51 | Chloroform | 4UJ | 1U |
| RX | 51 | Chloromethane | 4J* | 1U |
| RX | 51 | Cis-1,2-Dichloroethene | 4UJ | 2* |
| RX | 51 | cis-1,3-Dichloropropene | 4UJ | 1U |
| RX | 51 | d8-Toluene | | |
| RX | 51 | DBCP | 4UJ | 1U |
| RX | 51 | Dibromochloromethane | 4UJ | 1U |
| RX | 51 | Dibromomethane | 4UJ | 1U |
| RX | 51 | Ethane, 1,1,1,2-Tetrachl | 4UJ | 1U |
| RX | 51 | ETHANE, 1,1,2,2-TETRACHL | 4UJ | 1U |
| RX | 51 | Hexachlorobutadiene | 4UJ | 1U |
| RX | 51 | Isopropylbenzene (Cumene | 4UJ | 2* |
| RX | 51 | Methane, Dichlorodifluor | 1700J* | 5* |
| RX | 51 | Methylene Chloride | 140J* | 1U |
| RX | 51 | Naphthalene | 4UJ | 26* |
| RX | 51 | p-Isopropyltoluene | 4UJ | 1U |
| RX | 51 | Sec-Butylbenzene | 4UJ | 1U |
| RX | 51 | Tert-Butylbenzene | 4UJ | 1U |
| RX | 51 | Tetrachloroethene | 4UJ | 0.09J* |
| RX | 51 | Toluene | 6J* | 2* |
| RX | 51 | Total Xylenes | 7J* | |
| RX | 51 | trans-1,2-Dichloroethene | 4UJ | 0.08J* |
| RX | 51 | trans-1,3-Dichloropropen | 4UJ | 1U |
| RX | 51 | Trichloroethene | 4UJ | 0.3J* |
| RX | 51 | Trichlorofluoromethane | 38J* | 1U |
| RX | 51 | Vinyl Acetate | REJ | REJ |
| RX | 51 | Vinyl Chloride | 4UJ | 1U |
| RX | 51 | %RECOV:1,2-Dichloroethane-d4 (| 136 | 92 |
| RX | 51 | %RECOV:d8-Toluene | 131 | 91 |
| RX | 51 | %RECOV:p-Bromofluorobenzene | 62 | 105 |
| RX | 51 | %RECOV:Surrog: 1-Bromo-2-Fluoro | 320 | 87 |

C-3. METALS

Below are the definitions for qualifiers used in the Metals area when qualifying data from metals analysis.

Data Qualifiers

- | | | |
|-----|---|--|
| U | - | Element was analyzed for but not detected. The associated numerical value is the instrument detection limit/method detection limit. |
| J | - | The analyte was detected above the instrument detection limit but not quantified within expected limits of precision. The laboratory has established minimum quantitation limits having a relative standard deviation of no more than 10%. |
| E | - | The reported value is an estimate because of the presence of interference. |
| B | - | Analyte found in the analytical blank as well as the sample, indicating possible/probable contamination. "B" accompanies those analytical results within 10 (10x) times the instrument detection limit for the analyte of interest. |
| N | - | Spike sample recovery not within control limits. |
| NAR | - | There is <u>no analysis result</u> for this analyte. |
| NA | - | Not Applicable/Not Required. |
| * | - | The analyte was present in the sample. |

| Type Request SampleNo Description | Ash-Boiler Metals 90354269 BOTTOM ASH | Ash-Boiler Metals 90354269 BOTTOM ASH Lab duplicate | Soil Metals 90354731 APC-1 | Soil Metals 90354732 APC-2 Field duplicate | Soil Metals 90354733 BLUE LAKE | Soil Metals 90354742 CITY-C Central | Soil Metals 90354743 CITY-JAPO Japanski I. | Soil Metals 90354741 CITY-N North | Soil Metals 90354749 CITY-S South | Soil Metals 90354721 DEEP INLET Background | Soil Metals 90354730 GALANKIN I |
|--|--|--|-------------------------------------|---|---|---|--|---|---|--|--|
| Station Taken QA_Code Matrix Units | 900828 | 900828 LDP1 | 900830 | 900830 | 900830 | 900831 | 900831 | 900831 | 900831 | 900829 | 900830 |
| Ag | .20U | .20U | 0.20J* | 0.20U | 0.20U | 0.20U | 0.20U | 0.20U | 0.20U | 0.20U | 0.20U |
| Al | 18800.* | 19400.* | 10900.* | 12800* | 25500* | 11100.* | 18000.* | 9620.* | 13500.* | 4680.* | 34900* |
| As | 29.8* | 28.4* | 146.* | 118* | 18.7* | 6.7J* | 24.1* | 5.1J* | 23.9* | 9.3J* | 14J* |
| Ba | 305E* | 164.E* | 174.* | 196* | 110* | 73.6* | 43.0* | 100.* | 77.2* | 7.08* | 23.6* |
| Be | .504* | .48J* | 0.15J* | 0.13J* | 0.10U | 0.10U | 0.18J* | 0.10U | 0.26J* | 0.10U | 0.40J* |
| Ca | 112000.* | 95500.* | 12300.* | 12700* | 4360* | 21900.* | 5190.* | 20100.* | 4920.* | 5890.* | 4050* |
| Cd | .20U | .20U | 0.20U | 0.20U | 0.20U | 0.20U | 0.20U | 0.20U | 0.20U | 0.51J* | 0.20U |
| Co | 11.8* | 11.0* | 12.8* | 13.2* | 17.6* | 5.84* | 8.53* | 4.25* | 9.33* | 7.8* | 9.31* |
| Cr | 122.* | 108.* | 88.3* | 94.2* | 63.3* | 24.6* | 33.7* | 18.1* | 28.8* | 19.3* | 11.2* |
| Cu | 191.* | 180.* | 400.* | 340* | 55.2* | 13.5* | 26.9* | 11.4* | 22.3* | 18.0* | 19.0* |
| Fe | 30700.* | 29700.* | 65200.* | 58000* | 42300* | 17200.* | 29900.* | 12700.* | 27500.* | 5890.* | 25900* |
| Hg | 0.004U | | .021J* | .027J* | .028J* | .018J* | .051* | .024J* | .039J* | .033J* | .029J* |
| K | 8940.* | 8740.* | 1040.* | 1010* | 1740* | 1010.* | 775.* | 1270.* | 775.* | 1250* | 199* |
| Mg | 65300.* | 60600.* | 7830.* | 9460* | 13600* | 7330.* | 9430.* | 4750.* | 6650.* | 5650* | 5290* |
| Mn | 4020.* | 3450.* | 640.* | 576* | 943* | 279.* | 470.* | 190.* | 358.* | 123.* | 325* |
| Mo | 29.6* | 23.5* | 22.9* | 30.5* | 0.20U | 0.20U | 0.20U | 0.20U | 0.20U | 1.2J* | 0.20U |
| Na | 63900.* | 56100.* | 484.* | 358* | 59.3* | 302.* | 170.* | 281.* | 170.* | 4310* | 854* |
| Ni | 216.* | 180.* | 57.0* | 56.5* | 36.7* | 12.2* | 20.1* | 8.30* | 18.1* | 109.* | 10.9* |
| Pb | 18.8E* | 31.1E* | 144.* | 260* | 12.2* | 3.6J* | 52.8* | 3.2J* | 8.5J* | 8.1J* | 2.7J* |
| Sb | 9.9J* | 9.1J* | 27.4* | 22.4* | 3.0U | 3.0U | 3.0U | 3.0U | 3.0U | 3.0U | 3.0U |
| Se | 1.0U | 1.0U | 0.20U | 0.20U | 0.20U | 0.20U | 0.20U | 0.20U | 3.42* | 0.63J* | 0.20U |
| Tl | 5.0U | 5.0U | 0.25UN | 0.25UN | 0.25UN | 0.25UN | 0.25UN | 0.25UN | 3.71N* | 0.25UN | 0.25UN |
| V | 350.* | 296.* | 56.9* | 56.3* | 89.0* | 41.5* | 51.3* | 34.2* | 40.6* | 14.3* | 64.0* |
| Zn | 196.* | 190* | 1447.* | 1150* | 72.8* | 56.2* | 241.* | 31.3* | 172.* | 20.1* | 24.4* |

| Type Request SampleNo Description | Soil Metals 90354747 JAMESTOWN Bay | Soil Metals 90354748 THIMBLEBER Bay | Leach-Sed TCL 90354745 SITKA LAND Sludge in catch basin | Sediment TCL 90354710 SAWMILL CO Cove | Sediment TCL 90354710 SAWMILL CO Lab duplicate | Sediment TCL 90354711 HERRING CO Cove | Sediment TCL 90354712 BLUE LAKE | Sediment TCL 90354704 EAST CHANN Eastern Channel | Leachate TCL 90354267 HOG FUEL S Storage at Herring Cove | Leachate TCL 90354740 SITKA LAND | Leachate TCL 90354744 SITKA CATC Field blank (QA) | Water TCL 90354746 BLUE LAKE |
|--|--|---|---|--|---|--|--|--|--|---|---|---------------------------------------|
| Station Taken QA_Code Matrix Units | 900831 | 900831 | 900831 | 900829 | 900829 LOP1 | 900827 | 900830 | 900828 | 900829 | 900831 | 900831 | 900830 |
| Ag | 0.20U | 0.20U | 4.0U | 0.28J* | 0.34J* | 0.21J* | 0.20U | 0.20U | 2.0U | 2.0U | 2.0U | 2.0U |
| Al | 23100* | 18500* | 4060* | 8270* | 7990* | 11700* | 38700* | 11800* | 810.* | 150* | 10U | 24J* |
| As | 21.3* | 20.0* | 87.9* | 31.8* | 36.7* | 27.1* | 46.5* | 20.8* | 15.8* | 1.5U | 1.5U | 3.0U |
| Ba | 32.5* | 32.2* | 224* | 64.5* | 61.6* | 57.7* | 204* | 50.8* | 109.* | 155* | 1.0U | 10.5* |
| Be | 0.22J* | 0.22J* | 0.703* | 0.10U | 0.10U | 0.10* | 0.16J* | 0.10U | 1.0U | 1.0U | 1.0U | 1.0U |
| Ca | 5520* | 4980* | 6780* | 6800* | 6600* | 7130* | 9140* | 5170* | 132000.* | 112000.* | 30.0* | 3910.* |
| Cd | 0.20U | 0.20U | 15U | 3.73* | 4.09* | 1.2J* | 0.20U | 1.4JB* | 2.0U | 2.0U | 2.0U | 2.0U |
| Co | 10.1* | 10.4* | 40.6* | 6.65* | 5.86* | 5.69* | 40.5* | 7.82* | 5.0U | 5.0U | 5.0U | 5.0U |
| Cr | 54.7* | 51.5* | 21.7* | 56.6* | 55.7* | 41.8* | 105* | 55.2* | 5.0U | 5.0U | 5.0U | 5.0U |
| Cu | 43.2* | 24.3* | 308* | 64.9E* | 61.6E* | 32.3E* | 134E* | 58.0E* | 13.5* | 3.7J* | 2.0U | 2.0U |
| Fe | 31600* | 29400* | 444000* | 10600* | 10300* | 16900* | 64000* | 18900* | 5640.* | 76500* | 2.8J* | 30.9* |
| Hg | .029J* | .021J* | .024J* | .016J* | | .004U | .022J* | .017J* | 0.08U | .013J* | .04U | .04U |
| K | 674* | 657* | 346* | 3390* | 3280* | 3190* | 2400* | 2240* | 62100.* | 50300.* | 300U | 640J* |
| Mg | 10400* | 10200* | 2000* | 12800* | 12600* | 12900* | 24400* | 11300* | 249000.* | 28800* | 3.5J* | 290.* |
| Mn | 582* | 522* | 633* | 149E* | 140E* | 196E* | 2010E* | 322E* | 6600.* | 891.* | 1.0U | 11.0* |
| Mo | 0.20U | 0.20U | 60U | 26.0* | 26.6* | 20.9* | 0.20U | 11.1* | 2.0U | 2.0U | 2.0U | 2.0U |
| Na | 167* | 209* | 1390* | 47200* | 46700* | 45500* | 463E* | 18300* | 503000.* | 249000* | 425.* | 1360.* |
| Ni | 25.5* | 24.7* | 39.9* | 43.9* | 44.1* | 23.2* | 73.0* | 30.4* | 43J* | 10U | 10U | 10U |
| Pb | 87.8* | 24.3* | 21.8* | 10.9* | 12.3* | 7.9J* | 6.6J* | 5.7J* | 3.0J* | 2.0J* | 1.0U | 10U |
| Sb | 3.0U | 3.0U | 19.3* | 3.0U | 3.0U | 3.0U | 3.0U | 3.0U | 30U | 30U | 30U | 30U |
| Se | 0.20U | 0.20U | 48.8* | 1.61* | 1.77* | 1.71* | 0.36J* | 0.66J* | 2.0U | 2.0U | 2.0U | 8.0U |
| Tl | 0.25UN | 0.25UN | 0.25UN | 0.28J* | 0.25U | 0.25U | 0.25U | 0.25U | 2.5U | 2.5U | 2.5U | 5.0UE |
| V | 64.5* | 59.8* | 100U | 96.0* | 92.9* | 63.5* | 190* | 61.2* | 4.9J* | 2.0U | 2.0U | 2.0U |
| Zn | 118* | 86.9* | 7200* | 93.8E* | 96.1E* | 245E* | 364E* | 68.9E* | 34.6* | 81.3* | 5.8J* | 16J* |

| | | | | |
|------------------|--------------------|-------------------|------------|--------------------|
| Type | Water-Mar | Water-Mar | Water-Mar | Water-Mar |
| Request | TCL(mtls) | TCL(mtls) | TCL(mtls) | TCL(mtls) |
| SampleNo | 90354705 | 90354706 | 90354707 | 90354708 |
| Description | HERRING CO Cove | THIMBLEBER Bay | E. CHANNEL | SAWMILL CO Cove |
| Station Taken | 900827 | 900828 | 900828 | 900829 |
| QA_Code | | | | |
| Matrix | Water | Water | Water | Water |
| Units | ug/l | ug/l | ug/l | ug/l |
| Ag | 2.0U | 10U | 2.0U | 2.0U |
| Al | 635.* | 672.* | 592.* | 761E* |
| As | 3.0U | 3.0U | 3.0U | 3.0U |
| Ba | 6.5* | 5.06* | 5.73* | 5.6* |
| Be | 1.0U | 1.0U | 1.0U | 1.0U |
| Ca | 316000* | 336000.* | 336000* | 375000* |
| Cd | 10U | 20U | 20U | 2.0U |
| Co | 25U | 25.U | 25U | 25.U |
| Cr | 5.0U | 5.0U | 5.0U | 5.0U |
| Cu | 16.2* | 2.0U | 2.3J* | 181* |
| Fe | 98.8* | 56.9* | 32.8* | 200J* |
| Hg | .19J* | .04U | .16J* | 0.15U |
| K | 293000* | 314000.* | 314000* | 352000* |
| Mg | 966000* | 1030000.* | 1030000* | 1150000* |
| Mn | 50U | 50U | 40U | 50U |
| Mo | 20U | 40U | 20U | 40U |
| Na | 8010000* | 7690000* | 8060000* | 6960000* |
| Ni | 50U | 50U | 50U | 50U |
| Pb | 18J* | 12J* | 10U | 10U |
| Sb | 30U | 30U | 30U | 30U |
| Se | 8.0U | 8.0U | 8.0U | 8.0U |
| Tl | 5.0UE | 5.0UE | 5.0UE | 5.0UE |
| V | 2.0U | 2.0U | 2.0U | 22J* |
| Zn | 25J* | 10U | 10U | 267.* |

C-4. CONVENTIONALS

1. T, Salinity, DO, pH
2. TOC, particle size

APPENDIX C-4. Table 1.

Hydrolab data

| Station | Date | Time | Depth (m) | Temp (C) | Salinity (o/oo) | DO (mg/L) | %Sat-DO | pH |
|----------------|-----------|-------|--------------|-------------|--------------------|--------------|---------|------|
| 1-Thimbleberry | 28-Aug-90 | 10:21 | 1.0 | 15.5 | 26 | 5.0 | 58 | 6.00 |
| 1-Thimbleberry | 28-Aug-90 | 11:21 | 1.0 | 15.2 | | 5.3 | 61 | 7.50 |
| 1-Thimbleberry | 28-Aug-90 | 10:45 | 1.0 | 15.3 | | 5.3 | 61 | 6.75 |
| 1-Thimbleberry | 28-Aug-90 | 10:48 | 10.0 | 13.6 | | 5.7 | 65 | 7.33 |
| 1-Thimbleberry | 28-Aug-90 | 10:54 | 19.9 | 12.4 | | 5.6 | 63 | 7.54 |
| 1-Thimbleberry | 28-Aug-90 | 10:57 | 29.9 | 11.2 | 32 | 5.5 | 61 | 7.61 |
| 1-Thimbleberry | 28-Aug-90 | 11:00 | 40.1 | 9.6 | | 4.8 | 51 | 7.61 |
| 1-Thimbleberry | 28-Aug-90 | 11:05 | 50.0 | 8.3 | | 3.9 | 40 | 8.30 |
| 1-Thimbleberry | 28-Aug-90 | 11:12 | 57.8 | 7.8 | 32 | 3.3 | 34 | 7.47 |
| 2-E.Channel | 28-Aug-90 | 16:22 | 1.0 | 15.6 | 22 | 6.9 | 79 | 7.53 |
| 2-E.Channel | 28-Aug-90 | 15:54 | 1.0 | 15.6 | | 4.8 | 55 | 7.44 |
| 2-E.Channel | 28-Aug-90 | 15:57 | 10.0 | 13.7 | | 6.3 | 70 | 7.67 |
| 2-E.Channel | 28-Aug-90 | 16:01 | 20.0 | 12.7 | | 5.3 | 59 | 7.72 |
| 2-E.Channel | 28-Aug-90 | 16:04 | 30.0 | 11.5 | 32 | 6.0 | 67 | 7.75 |
| 2-E.Channel | 28-Aug-90 | 16:07 | 40.0 | 10.0 | | 5.6 | 60 | 7.75 |
| 2-E.Channel | 28-Aug-90 | 16:10 | 50.0 | 8.3 | | 4.3 | 44 | 7.70 |
| 2-E.Channel | 28-Aug-90 | 16:13 | 60.0 | 7.3 | | 2.8 | 28 | 7.70 |
| 2-E.Channel | 28-Aug-90 | 16:17 | 66.4 | 7.0 | 32 | 2.0 | 20 | 7.50 |
| 3-Sawmill Cove | 29-Aug-90 | 10:07 | 1 | 15.1 | 20 | 9.07 | 101 | 5.49 |
| 3-Sawmill Cove | 29-Aug-90 | 10:49 | 1 | 13.9 | | 10.26 | 111 | 7.81 |
| 3-Sawmill Cove | 29-Aug-90 | 10:46 | 10 | 13.6 | | 5.39 | 60 | 7.68 |
| 3-Sawmill Cove | 29-Aug-90 | 10:12 | 10 | 13.8 | | 4.84 | 54 | 6.4 |
| 3-Sawmill Cove | 29-Aug-90 | 10:16 | 20 | 12.4 | | 5.43 | 60 | 7.15 |
| 3-Sawmill Cove | 29-Aug-90 | 10:43 | 20 | 12.2 | | 5.52 | 61 | 7.55 |
| 3-Sawmill Cove | 29-Aug-90 | 10:20 | 30 | 11 | | 4.29 | 47 | 7.47 |
| 3-Sawmill Cove | 29-Aug-90 | 10:39 | 30 | 10.5 | 32 | 5.32 | 58 | 7.41 |
| 3-Sawmill Cove | 29-Aug-90 | 10:24 | 40 | 9.5 | | 4.35 | 46 | 7.54 |
| 3-Sawmill Cove | 29-Aug-90 | 10:27 | 50.1 | 8.1 | | 3.07 | 32 | 7.52 |
| 3-Sawmill Cove | 29-Aug-90 | 10:35 | 59.4 | 6.7 | | 1.26 | 13 | 7.41 |
| 3-Sawmill Cove | 29-Aug-90 | 10:31 | 60 | 6.9 | 32 | 1.34 | 13 | 7.47 |
| 4-Herring Cove | 27-Aug-90 | 15:28 | 0.9 | 16.3 | 22 | 7.3 | 84 | 5.50 |
| 4-Herring Cove | 27-Aug-90 | 15:30 | 10.2 | 13.2 | | 6.6 | 73 | 6.46 |
| 4-Herring Cove | 27-Aug-90 | 15:32 | 19.9 | 12.0 | | 5.7 | 63 | 6.81 |
| 4-Herring Cove | 27-Aug-90 | 15:34 | 30.1 | 10.6 | 32 | 5.4 | 59 | 7.06 |
| 4-Herring Cove | 27-Aug-90 | 15:38 | 40.0 | 9.1 | | 5.5 | 58 | 7.26 |
| 4-Herring Cove | 27-Aug-90 | 15:41 | 50.0 | 7.6 | | 4.0 | 41 | 7.34 |
| 4-Herring Cove | 27-Aug-90 | 15:45 | 60.1 | 6.7 | | 2.6 | 26 | 7.34 |
| 4-Herring Cove | 27-Aug-90 | 15:49 | 63.3 | 6.5 | 32 | 2.1 | 21 | 7.28 |

APPENDIX C-4. Table 2. Results from Total Organic Carbon and grain size analysis of sediment samples.

| <u>Location</u> | <u>TOC</u> | <u>% Sand</u> | <u>% Silt</u> | <u>%Clay</u> | <u>%Silt + Clay</u> |
|----------------------|------------|---------------|---------------|--------------|---------------------|
| Silver Bay | | | | | |
| Eastern Channel | 8.2% | 27.0% | 6.3% | 4.8% | 11.1% |
| Sawmill Cove | 30.1% | 30.9% | 28.5% | 24.8% | 53.3% |
| (duplicate) | 26.2% | 34.6% | 29.7% | 23.7% | 53.4% |
| Herring Cove | 17.5% | 25.1% | 36.7% | 31.8% | 68.5% |
| Blue Lake | 2.7% | 5.2% | 46.9% | 47.5% | 94.4% |
| City Landfill-sludge | 3.8% | 29.8% | 16.5% | 52.7% | 69.2% |

C-5. AQUATIC ORGANISMS

| SAMPLE# | FISH LOCATION ID # | DATE COLLECTED | GEAR | SPECIES | LENGTH(mm) | WEIGHT(g) | ABNORMAL- ITIES |
|---------|-----------------------|-------------------|------------|--------------------------|------------|-------------|--------------------|
| 1 | 82 Blue Lake | 30-Aug-90 | Trap | Trout-Rainbow | 217 | 124 | |
| 1 | 81 Blue Lake | 30-Aug-90 | Trap | Trout-Rainbow | 161 | 50 | |
| 1 | 83 Blue Lake | 30-Aug-90 | Trap | Trout-Rainbow | 208 | 108 | |
| 2 | 2 Herring Cove | 27-Aug-90 | Set line | Rockfish-quillback | 261 | 310 | |
| 2 | 5 Herring Cove | 27-Aug-90 | Set line | Rockfish-quillback | 392 | 1065 | |
| 3 | 77 Sawmill Bay | 30-Aug-90 | Pot-tanner | Crab-Dungeness-2 | 178 | | |
| 4 | 64 Sawmill Bay | 29-Aug-90 | Set line | Rockfish-quillback | 331 | 665 | |
| 4 | 62 Sawmill Bay | 29-Aug-90 | Set line | Rockfish-quillback | 407 | 1310 | |
| 4 | 65 Sawmill Bay | 29-Aug-90 | Set line | Rockfish-quillback | 264 | 360 | |
| 5 | 67 Sawmill Bay | 29-Aug-90 | Set line | Sculpin-Pacific staghorn | 247 | 190 | |
| 5 | 66 Sawmill Bay | 29-Aug-90 | Set line | Sculpin-Pacific staghorn | 278 | 335 | |
| 5 | 34 Sawmill Bay | 29-Aug-90 | Set line | Sculpin-Pacific staghorn | 314 | 430 | |
| 5 | 29 Sawmill Bay | 29-Aug-90 | Gill net | Sculpin-Pacific staghorn | 255 | 245 | |
| 5 | 30 Sawmill Bay | 29-Aug-90 | Gill net | Sculpin-Pacific staghorn | 300 | 389 | |
| 6 | 55 Sawmill/Herring | 29-Aug-90 | Trawl | Flatfish-Sole-english | 426 | 790 | |
| 6 | 57 Sawmill/Herring | 29-Aug-90 | Trawl | Flatfish-Sole-english | 438 | 755 | |
| 6 | 56 Sawmill/Herring | 29-Aug-90 | Trawl | Flatfish-Sole-english | 479 | 1095 | |
| 6 | 54 Sawmill/Herring | 29-Aug-90 | Trawl | Flatfish-Sole-english | 406 | 835 | |
| 6 | 58 Sawmill/Herring | 29-Aug-90 | Trawl | Flatfish-Sole-english | 324 | 345 | |
| 7 | 59 Sawmill/Herring | 29-Aug-90 | Trawl | Shrimp-mixed (pinks) | #--150 | 735 | |
| 8 | 24 Thimbleberry Bay | 28-Aug-90 | Set line | Flatfish-Pacific sanddab | 250 | 126 | |
| 8 | 17 Thimbleberry Bay | 28-Aug-90 | Set line | Flatfish-Pacific sanddab | 258 | 165 | |
| 8 | 20 Thimbleberry Bay | 28-Aug-90 | Set line | Flatfish-Pacific sanddab | 240 | 130 | |
| 8 | 19 Thimbleberry Bay | 28-Aug-90 | Set line | Flatfish-Pacific sanddab | 267 | 170 | |
| 8 | 15 Thimbleberry Bay | 28-Aug-90 | Set line | Flatfish-Pacific sanddab | 306 | 230 | |
| 8 | 16 Thimbleberry Bay | 28-Aug-90 | Set line | Flatfish-Pacific sanddab | 250 | 68 | |
| 8 | 23 Thimbleberry Bay | 28-Aug-90 | Set line | Flatfish-Pacific sanddab | 245 | 98 | |
| 8 | 21 Thimbleberry Bay | 28-Aug-90 | Set line | Flatfish-Pacific sanddab | 309 | 265 | |
| 8 | 18 Thimbleberry Bay | 28-Aug-90 | Set line | Flatfish-Pacific sanddab | 311 | 110 | |
| 8 | 22 Thimbleberry Bay | 28-Aug-90 | Set line | Flatfish-Pacific sanddab | 290 | 215 | |
| 9 | 14 Thimbleberry Bay | 28-Aug-90 | Set line | Rockfish-quillback | 378 | 1055 | |
| 10 | 80 Thimbleberry Bay | 30-Aug-90 | Hand | Shellfish-mussels | | 1/2 gal jar | |
| 11 F-3 | Thimbleberry Lake | 28-Aug-90 | Trap | Trout-Eastern brook | 186 | 70 | |
| 11 | 60 Thimbleberry Lake | 29-Aug-90 | Trap | Trout-Eastern brook | 187 | 76 | |
| 11 F-2 | Thimbleberry Lake | 28-Aug-90 | Trap | Trout-Eastern brook | 214 | 118 | |
| 11 F-1 | Thimbleberry Lake | 28-Aug-90 | Trap | Trout-Eastern brook | 188 | 78 | |

C-6. LOCATION OF SOIL SAMPLES

APPENDIX C-6. Locations of soil samples.

| <u>Sample Name</u> | <u>Date</u> | <u>Subsamples and their Locations</u> |
|--------------------|-------------|---|
| Sitka City | | |
| -North | 8/31/90 | <ol style="list-style-type: none"> 1. Women's ball field, home plate. 2. Men's ball field, home plate. 3. Verstovia school ball field, home plate. 4. Residence at the southeast corner of Kostrometinoff and Edgecumbe Streets; garden soil just above the sidewalk. |
| -Central | 8/31/90 | <ol style="list-style-type: none"> 1. Moller Park, northern playground, middle of the swingset. 2. Moller Park, southeast playground, middle of the swingset. 3. Moller Park, ball field, home plate. 4. Blatchley school, ball field, home plate. |
| -South | 8/31/90 | <ol style="list-style-type: none"> 1. Etolin street school ballfield, home plate. 2. Sheldon Jackson Harbor park, swingset on south side of tennis courts. 3. Residence (# 106, end of Barlow Road), soil near roadside. 4. South side of jogging trail near the City incinerator. |
| -Japonski Island | 8/31/90 | <ol style="list-style-type: none"> 1. Coast Guard housing playground, swingset. 2. Mt. Edgecumbe preschool (430 Fairway) playground, near base of tree, away from beauty bark. 3. Mt. Edgecumbe elementary school (Charcoal & Alice Loop Roads), bus loading area. 4. Residence one block west of the elementary school (house # 480A), soil near sidewalk. |
| Galankin Island | 8/30/90 | <ol style="list-style-type: none"> 1. Back yard of residence in middle of northeast side. 2. Mt. side of side yard of residence at southeast end. 3. Side of trail between the middle residence and the quarry. 4. Bare patch of soil on the northeast side of the quarry pit. |
| Jamestown Bay | 8/31/90 | <ol style="list-style-type: none"> 1. Residence (1320 Raven Island) west end of the Bay (Price Street exit off of Sawmill Road), soils at edge of driveway. 2. Residence (107 Wolff Drive), soil at edge of driveway. 3. Residence (1618 Sawmill Road), soil near edge of sidewalk. 4. Residence (# 3 at a Trailer park) at east end of the Bay, soil between back door and driveway. |
| Thimbleberry Bay | 8/31/90 | <ol style="list-style-type: none"> 1. Residential lot at west end (off of Shotgun Alley at approximately 2103 Sawmill Road), soil near side of road. 2. Residence (205 Blueberry Lane, at approximately 2507 Sawmill Road), soil from south part of yard. 3. Thimbleberry Lake trailhead, soil from base of tree to the east of the sign. 4. Roadside near Thimbleberry Creek, soil near west end of guardrail on south side of road. |
| Deep Inlet | 8/29/90 | 1-5. Bare patches of soil on trail near the south shoreline at the head of the inlet |
| Alaska Pulp Corp | 8/30/90 | <ol style="list-style-type: none"> 1. About 5 ft southeast of the east leg of the pylon supporting the chip conveyor (near the weak red liquor tank). 2. Between the tracks and the curb near the SO₂ solution tank and the nearby drain. 3. Near the base of the pylon supporting cables and hoses (around the corner from the location of subsample #2). 4. Across from the toe of the concrete near the 3B absorption tower; between the tracks and the nearby drain. |
| Blue Lake | 8/30/90 | 1-4. Two from the northwest edge of the launching area. Two from the southeast edge of the launch/parking area. |

APPENDIX D. QUALITY ASSURANCE REPORTS

1. Quality assurance reports are available from EPA, Region 10, for analyses completed by the following laboratories:

EPA, Region 10, laboratory
Contract Laboratory Program
ADEC ash and floating residue samples
(completed by ENSECO)

2. Table of laboratory methods used including details on TOC, particle size, and salinity
3. Quality assurance plan for the survey

**D-2. Laboratory methods used
including details on TOC, particle size, and salinity**

LABORATORY METHODS

| <u>Laboratory</u> | <u>Analyte</u> | <u>Method</u> | <u>Comment</u> |
|----------------------|--------------------------|-------------------------|--|
| USEPA Region 7 | Dioxin/furan | EPA 8290 | Without recovery standard |
| USEPA Duluth | Dioxin/furan | EPA 8290 | EPA 600-3-90-022, Analytical procedures and quality assurance plan for determination of PCDD and PCDF in fish. |
| Contract Lab Program | Dioxin/furan TOC | EPA 8290 PS Protocol | Puget Sound Protocol for sediment TOC (EPA 910/9-88-200) |
| | Particle size | ASTM D422-63 | (See attached) |
| USEPA Region 10 | VOAs (volatiles) | EPA 8260 | Region 10 lab modifications |
| | B/N/Acid (semis) | EPA 8270 | Region 10 lab modifications |
| | Pesticides/PCBs | EPA 608 | |
| | Salinity | | See attached |
| | Metals-ICP scan | EPA 200.7 | (Total Recoverable) |
| | <u>Specified Metals:</u> | | |
| | Arsenic | EPA 206.2 | |
| | Lead | EPA 239.2 | |
| | Mercury (water) | EPA 245.1 | |
| | Mercury (sed/soil) | EPA 245.5 | |
| | Selenium | EPA 270.2 | |
| | Thallium | EPA 279.2 | |

Scope of Work for TOC and Particle Size Analyses

1. General description of analytical service requested:

Total Organic Carbon (TOC) analysis in sediment by combustion, using the attached method, dated March 1986.

The samples will also be analyzed for particle size using method ASTM D422.

2. Definition and number of work units:

There will be 6 low level sediment samples submitted for TOC and particle size analyses.

3. Estimated date(s) of collection/shipping:

The samples have already been collected.

The samples will be shipped via Federal Express as soon as laboratory arrangements can be made.

4. Number of days analysis and data required after laboratory receipt of samples:

The complete data package is required 35 days from receipt of the samples. The samples will be analyzed for TOC within 7 days of Validated Time of Sample Receipt and are to be stored at < 4 degrees Celsius.

5. Analytical protocol required:

The attached method, dated March 1986, will be used for the TOC analyses. Follow Method ASTM D422-63, Particle Size Analysis of Soils for the particle size analyses.

6. Special technical instructions:

Priority of analyses:

Sample size may be limited for some or all of the samples. Therefore, TOC shall be analyzed before particle size.

For TOC analyses:

* An infrared CO2 analyzer should be used if available, rather than a gravimetric method.

* Dried samples shall be weighed prior to acidification.

* Report organic carbon results as a percentage of the dry weight of the unacidified sample to the nearest 0.1 units.

* An initial five point calibration curve that covers the dynamic range of the detector will be performed at the beginning of each analytical batch or whenever a calibration check exceeds specifications. The low calibration standard shall be at the Quantitation Limit specified in section 9 of this scope of work. In addition, the initial calibration will also include a blank. The percent relative standard deviation (RSD) of each calibration factor is not to exceed 25% RSD.

* Matrix Spike and Matrix Spike Duplicate analyses are to be performed on 10% of the field samples utilizing potassium hydrogen phthalate as the spiking compound. Spiking concentrations should be approximately half the instrument calibration range.

* Matrix Spike and Matrix Spike Duplicate results target is RPD < 35% and the target mean recovery is 75 - 125% .

* Corrective action is required when method blank results exceed target detection limits.

* Triplicate sample analysis will be performed on sample 90354710. The %RPD for the sample triplicates shall be reported and shall be $\leq 35\%$.

* A calibration check standard will be analyzed after every ten field or QC samples analyzed. The calibration check standard will have a concentration equal to the mid-range standard of the established calibration curve. The response factor percent differences (%D) between the calibration check standard and the initial calibration standard will be $\leq 25\%$.

For particle size analyses:

Analyze one sample in duplicate. Follow method ASTM D422. Initially weigh each sample aliquot prior to analysis and determine % recovery relative to the cumulative weights after fractionating. Note: both sieve and hydrometer analyses are required.

7. Analytical results required:

The data package for TOC analyses shall include the following:

1. Chain-of-Custody sheets.

2. The bench sheets for sample preparation indicating dates, times and methods of preparation, standard information, spike volumes/amounts added, instrument run time/date, etc.
3. All sample and blank results reported as percentage on the dry weight and all supporting raw data including: run logs, computer printouts, calibration factors and QC results. All computer printouts will be labeled with a minimum of the sample number, and date/time of analysis.
4. All standard data including: preparation logs, traceability of standards, calibration factors (and RSD where applicable) for all standards analyzed.
5. All QC data including: summary forms, raw data and calculations.
6. An example calculation with the formula and all definitions of parameters in the formula in sufficient detail to allow an independent third party reconstruction of the results from the raw data.

The data package for particle size analyses shall include the following:

1. All Method D422-63 data reporting and documentation requirements.
2. The final results for each sample and it's duplicate.
3. All information used in calculating sample results.
4. Chain-of-Custody sheets.
4. Data on balance calibration checks for each day that samples are weighed.

8. Send the data packages to:

Laura Castrilli
USEPA Region 10
1200 Sixth Ave. MS/ES-095
Seattle, WA 98101

(Specify 9th. Floor if an express delivery service is used)

Phone: (206) 553-4323

9. Data Requirements

| <u>Parameter</u> | <u>Quantitation Limit</u> | <u>Precision Desired (percent or Concentration)</u> |
|---|---------------------------|---|
| <u>Particle size as per method ASTM D422-63</u> | <u>0.001 mm</u> | <u>< 35% RPD</u> |
| <u>TOC</u> | <u>5.0 mg/Kg</u> | <u>75-125% R</u> |

10. QC Requirements

| <u>Audits required</u> | <u>Frequency of Audits</u> | <u>Limits (percent or Concentration)</u> |
|------------------------------------|----------------------------|--|
| Particle size Analyses: | | |
| <u>Per method ASTM D422-63</u> | | |
| <u>Duplicate</u> | <u>one</u> | <u>< 35% RPD</u> |
| TOC analyses: | | |
| <u>Method Blanks</u> | <u>10 % Field Samples</u> | <u>< Detection Limit</u> |
| <u>MS/MSD</u> | <u>10 % Field Samples</u> | <u>RPD < 35 %, R=75- 125%</u> |
| <u>Calibration</u> | <u>1 per 10 samples</u> | <u>% Difference</u> |
| <u>Check Standard</u> | <u>analyzed</u> | <u>< 25%</u> |
| <u>Triplicate Analyses</u> | <u>on sample 90354711</u> | <u>RPD < 35 %</u> |

11. Action Required if Limits are Exceeded

Call Bruce Woods, QA chemist, at 206-553-1193, immediately,
for problem resolution.



UNITED STATES ENVIRONMENTAL PROTECTION AGENCY
REGION 10 LABORATORY
MANCHESTER, WASHINGTON 98353

REPLY TO
ATTN OF

M/S LAB

September 19, 1990

MEMORANDUM

SUBJECT: APC Salinity Analysis

FROM: Katherine York, CSC *KY*
Phil Davis *P. Davis*

TO: Leigh Woodruff
Bruce Duncan
Dan Tangarone

At the request of Bruce Duncan, four APC samples were analyzed for salinity on 9/11/90 by Phil Davis. These sample numbers are 90354700, 4701, 4702, and 4709. The request was for the analysis to be done on the top, middle, and bottom of the sample, which Phil did. Unfortunately, there is no simple way to report three values for each sample number with the data management system we presently use.

Rather than assign new sample numbers to the middle and bottom results, for example, we decided it would be best to report the results in the form of a memo.

| <u>Sample number</u> | <u>Top</u> | <u>Results in ppt</u> | |
|----------------------------|------------|-----------------------|---------------|
| | | <u>Middle</u> | <u>Bottom</u> |
| 90354700 | 22. | 32. | 32. |
| 90353701 | 26. | 32. | 32. |
| 90354702 | 22. | 32. | 32. |
| 90354709 | 20. | 32. | 32. |
| 90354701-Duplicate | 26. | 32. | 32. |
| Blank #1-BK0254 | | 0.0 | |
| Blank #2+HCl-BK0254A | | 0.0 | |
| Standard #1-34.995 ppt | | 35. | |
| Standard #2-20.ppt | | 20. | |
| Standard #3+HCl-34.995 ppt | | 35. | |

Phil Davis used a Refractometric method to do his analysis. Please contact him or myself if you have any questions.

cc: Carolyn Wilson, RSCC
Arthur Dan Baker, RSCC

D-3. Quality assurance plan for the survey

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QUALITY ASSURANCE PROJECT PLAN AND SAMPLING PLAN

Project Name: Alaska Pulp Corporation Multimedia & Risk Assessment Study,
Sitka, Alaska

Project Manager: Dan Tangarone, Multimedia Inspection
Bruce Duncan, Risk Assessment Study

Field Operations: Dan Tangarone, ESD, EPA & ADEC Inspectors
Bruce Duncan, ESD • Dave Terpening, ESD • Andy Hess, ESD
Fish & Wildlife Service (2)

QA Office Concurrence: *Paul G. [Signature]* Date 22 Aug 90

Peer Review: _____ Date _____

Account Number: _____ Sample Numbers: _____

PROJECT DESCRIPTION, OBJECTIVES, AND SITE LOCATION

APC has been selected for a multimedia compliance inspection which will involve EPA and ADEC program inspectors (Attachment A) during the week of August 27, 1990.

Previous studies of APC air emissions, wastewater discharges, and fly-ash disposal practices have raised concerns about dioxin contamination in soil, sediment, air, water, and biota in the vicinity of APC. Because of this, additional sampling (Attachment B) will be conducted during the same week to obtain data for a preliminary human health risk assessment study. Proposed sampling locations associated with this risk assessment phase are also listed in general terms in Attachment B.

The primary objectives of the August 27-31, 1990 APC survey are:

- Media program compliance inspections
- Human health risk assessment. The details of the toxicity and exposure assessments are shown on Attachment A.

SCHEDULE OF TASKS:

| | |
|---------------------|---|
| Sample Collection | August 27-31, 1990 |
| Analysis Completion | Variable depending upon analysis type and laboratory. Dioxin sample results are estimated to require 45 days after receipt by the laboratory. Analyses to be performed by the Region 10 EPA Laboratory involving the Target Compound |

| | |
|--|---|
| Data Summarization Report Preparation | List (TCL) will take approximately 45 days. Final results should be available by October 31, 1990. October 31, 1990 All compliance inspection / program reports will be generated by the program inspectors and will be submitted to the ESD APC multimedia coordinator (Dan Tangarone) for compilation and summary. Reports associated with the risk assessment will be generated by the risk assessment group of ESD. |
|--|---|

DATA USAGE:

Data will be used to determine: (1) program compliance and (2) initial evaluation of risks to human health via various media and exposure routes.

QA OBJECTIVES FOR MEASUREMENT DATA

Precision and Accuracy Protocols / Limits:

Lab: Accuracy will be monitored by matrix /matrix spike duplicates, and laboratory control samples.

Laboratory replicate samples will be analyzed at a frequency of five percent for each sample matrix received for all parameters.

Rinsate blanks used for the final rinse of sampling equipment (Van Veen dredge and Automatic samplers) and are analyzed for all parameters of interest..

DATA REPRESENTATIVENESS

Sample selection will be based primarily on biased and composite sampling.

DATA COMPARABILITY:

These data are collected as a stand alone investigation and will not be compared to any other data set.

DATA COMPLETENESS:

The target goal for completeness is 100 percent.

DETECTION LIMITS:

Method detection limits required are based on human health risk assessment criteria, best available technology criteria, matrix type, and location. Where detection limits required for risk assessment are lower than attainable by current technology, SAS will be employed when appropriate to reduce the uncertainty in the risk assessment. Detection limits for CLP RAS will employ contract required quantitation limits specified in the CLP Statements of Work for organic and inorganic analyses.

Soil Dioxin and Furans

tcdd and tcdf 2.0 ng/kg

ocdd and ocdf 20 ng/kg

others 10.0 ng/kg

Water Dioxin and Furans

tcdd and tcdf 0.02 ng/l

ocdd and ocdf 0.2 ng/l

others 0.1 ng/l

Tissue Dioxin and Furans

1 pg/kg for all 2,3,7,8 isomers

For all other parameters see attachment C.

SAMPLING PROGRAM

Samples to be obtained for the risk assessment phase and some NPDES related samples are shown in Attachment B. Other samples may be obtained during the conduct of the various media inspections. The programs which will involve the collection of samples are NPDES, Air, and Drinking Water. Samples may be obtained during the TSCA and RCRA inspections.

SAMPLE TYPES: See attachment B for sample types.

SAMPLE COLLECTION PROCEDURES:

Wastewater: One 24-Hour composite final effluent sample will be obtained by automatic sampler. In addition, 12-hour manual composite samples will be taken from the bleach plant discharges, the treatment plant influent and effluent and the final effluent.

Other water samples from Silver Bay and Blue Lake will be as shown on Attachment B and will also include TOC, pH, Conductivity, TSS, and TCL.

Water The marine water composite samples will be made up of approximately equal aliquots from the surface (5 Ft depth), mid point and bottom locations at each station using a Kemmerer sampler (Stainless or Teflon) or similar sampling device. The same sampler will be used at each location following a thorough ambient water rinse. A Hydrolab will be used for field analytical measurements.

Leachate Leachate samples will be obtained manually from the APC and city landfills.

Ash: Bottom ash from the APC and KPC hog fuel boilers will be collected using a stainless steel scoop or other appropriate sampling device as determined on location.

Sediment: Open water and near shore sediment samples will be collected from 4 areas in marine waters and from Thimbleberry Lake, Beaver Lake.

Debris or other objects larger than 0.5 inch diameter will be removed before placing the sample in the container.

Marine sediments will be collected using a stainless steel Van Veen grab sampler or other appropriate device. The upper 5 cm will be sampled from the center of each grab using clean stainless steel spoons. Two 8-oz wide mouth glass jar will be collected at each site for archiving. In addition, approximately equal portions of sediment from each of the four area locations will be composited into a half-gallon wide mouth glass jar or stainless steel pan. Sediments in this jar or pan will be mixed using a stainless spoon and a portion of this sample will be placed into 3, 8-oz and 1, 120 ml glass jars. One of the 8-oz jars will be archived, one will be sent

the the appropriate laboratory for dioxin analysis, and the third will be sent to Region 10 lab along with the 120 ml jar for TCL analysis. Sample suitability will be judged according to the Puget Sound Protocols.

Freshwater lake sediment samples will be collected at the shoreline directly into appropriate containers using stainless steel spoons.

Soil: Surface soils will be sampled using a stainless steel scoop or other appropriate sampling device. Compositing of discrete samples will be done in stainless steel bowls. The depth of the sample will not exceed 2 inches below the ground surface. Objects larger than 0.5 inch diameter will be removed before placing the sample in the sample jar. If the sampling location is covered by vegetation, the turf will be separated from the soil and discarded. The thickness of the discarded turf will not be included in the 2-inch depth determination.

Tissue: Whole organisms - bottomfish, crab, clams, trout - will be obtained by normal methods and will be frozen aboard the boat, identified and wrapped in aluminum foil for shipment to the laboratory.

Following the procedures used in the National Bioaccumulation Study (1986), whole fish, crab, clams and mussel samples will be wrapped in aluminum foil and frozen ASAP for shipment to the analytical laboratory.

Tissue compositing will be done at the lab as part of the analytical procedure. The composite sample should not exceed 20 pounds. The weight, length, and species will be included on the sample data sheet. For sport or high utilization fish, the lab will analyze fillets. For bottom feeder fish, the whole fish will be analyzed.

Air: Certain air streams within APC will be sampled using evacuated canisters during the air compliance inspection. Sources to be sampled and analyzed for chloroform will be determined on site during the inspection but are thought to be from the vent system from the bleaching operation. Approximately ten in-plant air samples will be obtained.

DECONTAMINATION PROCEDURES

Sampling equipment will be thoroughly cleaned with alconox , carbon free water and methanol when appropriate.

SAMPLE CUSTODY PROCEDURES:

Region 10 chain-of-custody procedures will be used in the field and during sample shipment to the laboratory. Laboratory custody and sample control procedures will be in accordance with procedures described in the CLP SOWs for organics (EPA 1988a) and inorganics (EPA 1988b).

SAMPLE PRESERVATION AND HOLDING TIMES

Samples will be preserved as required according to procedures presented in the CLP organic (EPA 1988a) and inorganic (EPA 1988b) statements of work, SW-846 (EPA 1986a), Methods for the Chemical Analysis of Water and Wastes (EPA 1983). The sample preservation and holding time requirements for this study are summarized in the attached table.

SAMPLE CONTAINERS

Type and number of sample containers required for the sampling program are shown in the attached tables.

SHIPPING REQUIREMENTS

Packaging, marking, labelling, and shipping of samples will comply with all regulations promulgated by the U. S. Department of Transportation (DOT) in the Code of Federal Regulations, 49 CFR 171 – 177 and International Air Transport Association (IATA) regulations. Detailed requirements are discussed in the CLP User's Guide (EPA 1986b).

CALIBRATION PROCEDURES:

All field instruments will be operated, calibrated and maintained according to manufacturer's guidelines and recommendations by qualified personnel. Field maintenance and calibration records will be recorded in the field notebook.

ANALYTICAL METHODS:

See Attachment B.

DOCUMENTATION:

Samples will be documented by use of the Region 10 Field Sample Data Sheet, a log book, photographs, traffic reports and shipping documents.

Each sample will be assigned a unique identifying number and documentation will include the following information:

- Name of sampler
- Date and time of sample collection
- Sample number
- Sample matrix and how collected (i.e., grab, composite)
- Preservation method
- Analyses required

DATA REDUCTION, VALIDATION, AND REPORTING:

All data generated by the laboratory will undergo a comprehensive quality assurance data validation. Data validation will assess laboratory performance in meeting the quality control specifications.

The project officer will oversee field measurements and data recording. Reduction and transfer of raw data from field notebooks to data tables will be independently checked and verified.

Data validation will assess laboratory performance in meeting the quality control specifications.

PERFORMANCE/SYSTEM AUDITS:

No additional audits are planned for this project.

CORRECTIVE ACTION:

The laboratory QA Officer will be responsible for ensuring that analytical results meet quality control criteria described in the CLP SOWs for organics (EPA 1988a) and inorganics (EPA 1988b) analyses or the appropriate EPA analytical method for SAS analyses and for implementing corrective actions as specified in the analytical methods and SOWs.

Appropriate corrective actions will be determined when and if required.

REPORTS:

Report development will be the responsibility of the Project Managers and/or program compliance inspectors.

| Parameter | Media | Method | Samples | QA Samples | Parameters |
|-----------------|------------|--------------|---------|------------|----------------------------|
| Chloroform | Air | TO-14 | 10 | | |
| Dioxin/Furans | Soil | 8290 | 8 | 1-Blank | |
| Metals | | CLP SOW | 8 | 1-Blank | |
| Particle Size | | ASTM D422-63 | 8 | 1-Blank | |
| Target Compound | Ash | CLP SOW | 1 | | BNA, Pest/PCB, Metals |
| Dioxin/Furans | | 8290 | 3 | | |
| TCLP | | FR 3/29/90 | 1 | | BNA, Pest/PCB, Metals |
| Dioxin/Furans | Leachate | 8290 | 2 | | |
| Target Compound | | CLP SOW | 2 | | BNA, Pest/PCB, Metals, VOA |
| Dioxin/Furans | Wastewater | 8290 | 4 | 1-Blank | |
| Chloroform | | CLP SOW | 4 | 1-Blank | |
| TOX | | 9020 | 4 | 1-Blank | |
| BOD | | 405.1 | 2 | 1-Blank | |
| TSS | | 160.3 | 2 | 1-Blank | |
| Free Chlorine | | 330.3 | 1 | | |
| | | | | | |
| Fecal Coliform | | 9132 | 1 | | |
| Dioxin/Furans | Sediment | 8290 | 9 | | |
| Target Compound | | CLP SOW | 13 | | BNA, Pest/PCB, Metals, VOA |
| Particle Size | | ASTM D422-63 | 6 | | |
| TOC | | 9060 | 5 | | |
| Dioxin/Furans | Water | 8290 | 1 | | |
| Target Compound | | CLP SOW | 5 | | BNA, Pest/PCB, Metals, VOA |
| Dioxin/Furans | Tissue | 8290 | 7 | | |
| lipids | | ?? | 7 | | |