WASTEWATER TREATMENT BY NATURAL AND ARTIFICIAL MARSHES



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WASTEWATER TREATMENT BY NATURAL AND ARTIFICIAL MARSHES

by

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ABSTRACT

Investigations were conducted on the use of artificial and natural marshes as purifiers of effluent from municipal treatment plants. Observations were made on marsh influent and effluent quality. Phosphorus distribution in the ecosystem and removal by harvesting were studied. Responses of the vegetation to repeated harvesting were recorded.

Artificial marshes consisted of plastic-lined excavations containing emergent vegetation, especially <u>Scirpus validus</u>, growing in gravel. Various combinations of retention time, primary effluent, secondary effluent, basin shape, and depth of planting medium were studied. A polluted natural marsh was studied simultaneously.

The degree of improvement in water quality suggests that the process may be acceptable for certain treatment applications. Harvesting was not a practical phosphorus removal technique. Marshes remove phosphorus in the growing season, but release it at other times. Development of management techniques for successful use of marshes for wastewater treatment is thought possible.

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SECTION I

CONCLUSIONS

MAJOR CONCLUSION

Emergent vegetation has been used to treat wastewater biologically to a degree of purity which suggests that continued research could lead to widespread applicability of the process.

MINOR CONCLUSIONS

General

- 1. Natural and artificial marshes are more aesthetically pleasing than most conventional treatment facilities.
- 2. Small natural or artificial marshes could be employed to polish effluent from septic tanks of single buildings or small clusters of buildings.
- 3. Conventional treatment plants which discharge effluent into wetlands possibly should not be required to provide the high degree of wastewater purification required of other treatment plants as long as other beneficial uses of the wetlands are not interfered with.
- 4. Marshes remove phosphorus from water during some periods and release it at others thus acting as buffers which may be managed to great advantage.

- 5. Emergent vegetation can be used seasonally in Wisconsin for wastewater treatment in applications involving temporary high waste loads in summer.
- 6. Harvesting of emergent vegetation is not a potential method of removing a large portion of phosphorus entering a marsh.

Specific

Phosphorus Removal From Brillion Marsh --

- 1. Older plant tissue contains little or no phosphorus.
- 2. Approximately 0.6 g P m^{-2} can be removed by a single late season harvest.
- 3. Only a small portion of the phosphorus retained by a marsh system is incorporated into harvestable plant tissue.
- 4. Repeated harvesting of a natural \underline{Typha} marsh removed only about 1 q P m⁻².
- 5. Plants at the downstream end of the natural marsh did not recover or retain the productivity of those at the upstream end of the marsh.
- 6. Harvesting greatly reduced the standing crop during mid-summer.

 Loss of cover could be detrimental to mammals and birds using the area.
- 7. There was replacement of <u>Typha</u> by <u>Sparganium</u> in at least one harvested quadrat.

- 8. The natural marsh was a moderately effective phosphorus removal system during the growing season, but the annual phosphorus output was about equal to the input.
- 9. Harvesting plants from Brillion Marsh would remove only about 6.5% of the phosphorus input.
- 10. Most of the phosphorus from the Brillion sewage treatment plant effluent was removed from the water in the first 100 m or so below the outfall.
- 11. The Spring Creek sediments immediately below the Brillion sewage treatment plant and those in a secondary stream receiving agricultural runoff contained 10 to 20 times as much phosphorus as the marsh sediments. Presumably, most of the phosphorus was precipitated or otherwise removed non-biologically in the stream channel.

Phosphorus Removal From Artificial Marshes --

- 1. Harvesting does not affect the quality of the effluent produced by the area on which the plants grew.
- 2. Older plant tissue contains little or no phosphorus.
- 3. Repeated harvesting of <u>Scirpus</u> during the growing season can result in the removal of 3.5 g P m^{-2} .
- 4. Only a small portion of the phosphorus retained by a marsh system is incorporated into harvestable plant tissue.

- 5. The artificial marshes retained 20 or more g P m-2 of which up to 80% is associated with the microflora substrate regime.
- 6. Most of the phosphorus associated with the microflora substrate regime is lost from the system over winter.
- 7. Harvesting reduced the total productivity (standing crop) in artificial systems, but to a lesser extent than in a natural marsh.

Water Quality Improvement In Brillion Marsh --

- 1. Spring Creek above the Brillion sewage treatment plant receives a variable loading of pollutants from industrial sources.
- 2. The effluent from the Brillion sewage treatment plant has, in general, high concentrations of BOD, COD, orthophosphate, total phosphorus and coliform bacteria.
- 3. During periods of low flow, as much as one-fourth to one-third of the total flow of Spring Creek where it enters Brillion Marsh is sewage treatment plant effluent.
- 4. During periods of low flow of Spring Creek the concentrations of BOD, COD, orthophosphate, total phosphate and coliform bacteria tend to be higher at Station II than at other times.
- 5. The average concentrations of BOD, COD, orthophosphate, total phosphorus and coliform bacteria were greater in Spring Creek below the sewage treatment plant than above.

- 6. The average value of conductivity, turbidity, nitrate, and total, suspended, and dissolved solids in Spring Creek were greater above the sewage treatment plant than below.
- 7. The average concentrations of BOD, COD, turbidity, nitrate, coliform bacteria, and suspended solids in Spring Creek were reduced by 29% or more by passage through some 1,900 m of Brillion Marsh from Station II to Station III.
- 8. The average concentrations of orthophosphate, total phosphorus, conductivity and total solids were reduced by 13% or less by passage through some 1,900 m of Brillion Marsh from Station II to Station III.
- 9. The observed reduction of orthophosphate, total phosphorus, conductivity and total solids could be primarily due to dilution from other, non-polluted waters entering Brillion Marsh.
- 10. Some of the observed reduction in the value of BOD, COD, turbidity, nitrate, coliform bacteria, and suspended solids is due to physical, chemical, and biological reactions in the marsh. Dilution also accounted for some of the observed reductions.
- 11. The concentration of phosphorus in the water draining from the marsh does not seem to follow any discernible pattern related to either temperature, rainfall, or season. It ranged from a low of 0.43 mg 1^{-1} in September to a high of 11.85 mg 1^{-1} in July.

- 12. The concentration of phosphorus in Spring Creek above the wastewater treatment plant follows a pattern. The higher values (1.14 to 3.61 mg P 1^{-1}) occurred in October and November, when fall frosts were occurring. The values were lowest (0.03 to 0.08 mg P 1^{-1}) in the winter months of December through March. Late spring, summer, and early fall values are intermediate. One very high reading (12.76 mg P 1^{-1}) of July 29, 1975 seems to be an anomaly, possibly due to a slug of contaminant.
- 13. A special intensive study showed the concentrations of the following constituents tended to be highly variable above the marsh but relatively steady below the marsh: suspended solids, total solids, BOD, nitrate, coliform bacteria, ammonia, and total phosphorus.
- 14. During the special intensive study, significant reduction in concentration of the following parameters were noted: nitrate, ammonia, coliform bacteria, total phosphorus, BOD, and suspended solids.
- 15. A mass balance study of total phosphorus, using estimated streamflow data and average phosphorus concentrations, shows the same order of magnitude of phosphorus leaving the marsh as entering it.

Water Quality In Artificial Marshes --

- 1. Evapotranspiration may remove a significant fraction of the wastewater (45%) from the system, thus, strongly influencing concentrations.
- 2. Scirpus validus, softstem bulrush, was the best species of those

tested for use in artificial marshes. This choice was made primarily because of the favorable response to harvesting. Since harvesting is not a good method of phosphorus removal, perhaps other species may prove to be more desirable because of other characteristics.

- 3. Phosphorus removal of over 80% should be attainable since 84% was attained in the greenhouse and 64% was attained under less than optimum field conditions for a wastewater that was very high in total phosphorus.
- 4. BOD removal of well over 90% should be attainable since over 90% was attained with field conditions that could only be poorly controlled.
- 5. Retention time of 5 10 days may be required although some studies using 5-hour retention time gave quite good results except for phosphorus removal and 16-hour retention time gave nearly as good results as 10-day retention in some cases.
- 6. Effluent quality is not influenced by harvesting the plant shoots.
- 7. The artificial marsh behaved in the same manner as Brillion Marsh with regard to phosphorus removal and responses of vegetation to harvesting and other treatments.
- 8. In plastic-lined systems the substrate, whether gravel or soil, should be probably only about 15 cm deep and wastewater should flow through it, not over it.

- 9. Percent reductions for most parameters were not greatly different when primary effluent was treated than when secondary effluent was treated.
- 10. Special intensive studies showed the marsh system effluent to be quite constant in quality over a 24 hour period.

SECTION II

RECOMMENDATIONS

- 1. Nutrient cycles should be studied extensively in marshes so that management techniques can be applied effectively.
- 2. Artificial marshes which are lined should be designed with a shallow (15 cm) substrate and wastewater should flow through the substrate rather than over it. Unlined systems using soil instead of gravel should be tested.
- 3. Artificial marshes should take the form of trenches of considerable length.
- 4. A method should be sought for controlling the fate of phosphorus which is flushed out of a marsh after killing frosts in autumn.
- 5. Research should be done on the possibility that vigor of emergent vegetation is lessened by lack of some limiting factor in both natural and artificial marshes.

SECTION III

INTRODUCTION

GOALS

The concept of using marshes for waste treatment or for polishing treated wastewater is an obvious outcome of the need for new, inexpensive technology. Application of the concept has been tested in Europe with varying degrees of success. The project reported on here, from June 1972, to June 1976, had the goal of showing that wastewater passed over and/or through masses of emergent vegetation would be improved in quality biologically and chemically. It was anticipated that nutrient removal could be affected by periodic harvesting of the upper portions of the plants (Figures 1 and 2). Secondary goals were to discover the effects of varying several parameters including water depth, retention time or loading rate, nature of wastewater applied (primary or secondary effluent) and frequency of harvesting of the vegetation on quality of effluent produced.

APPLICABILITY IN WISCONSIN CLIMATE

Wetlands are widely distributed in Wisconsin and would be readily available at comparatively low cost to a significant number of users. Application of the method would have to be seasonal for climate reasons, but a number of seasonal needs exist.



Figure 1. Pilot Plant With Bulrushes Prior to Harvest.

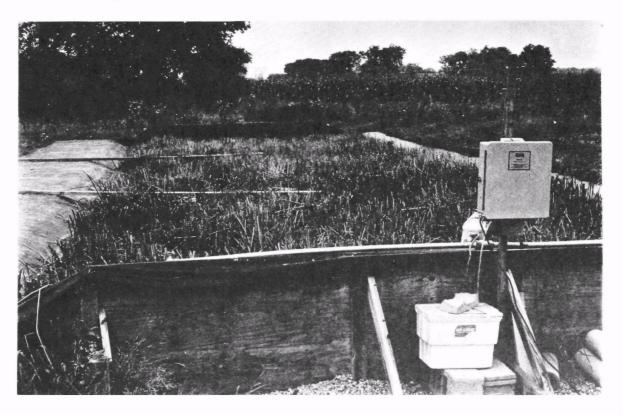


Figure 2. Pilot Plant After Bulrushes Have Been Harvested.

Needs include treatment of wastes produced by campgrounds, resort communities (with temporary high populations in summer), canning companies, agriculture, summer recreation camps, and trailer parks among others.

APPROACH

Experimental Basins and Pilot Plant

The approach used in this project involved use of plastic lined excavations in which vegetation was planted. Observations were also made on a natural marsh receiving water polluted by the outfall from a municipal treatment plant. Excavations were of two sizes. Small square ones (constructed in late summer, 1972) are referred to in this report as experimental basins. There was one large excavation, in the form of a trench, which is referred to as the pilot plant. Water depth in all excavations was controllable from zero to one meter. Flows into and out of the excavations were metered. Retention time could be varied by changing water depth and inflow rate. Analyses were performed one or more times per week on grab samples of influent and effluent. In some studies (1974 and 1975), samples were collected as frequently as every four hours for three to four days in order to discover short term temporal variation. In the later parts of the project a composite sampler was used for some sampling. Specimens of plant tissue were analyzed for phosphorus content, and cores were taken from the substrate for the purpose of studying nutrient distribution in the system.

Culture Techniques and Greenhouse Studies

Prior to field studies (summer 1972), some work was done on techniques for obtaining a number of kinds of vegetation from the natural habitat and propagating them in the study area. Also, some greenhouse observations were made (1972, 1973) in order to narrow the choice of species to be used in field applications. Following culture techniques and greenhouse experiments, field work using the experimental basins was begun (summer 1973).

In early field studies two kinds of planting media were tried. In some cases vegetation was rooted in gravel and in others it was tied to a hardware cloth on moveable frames. The frames offered the advantage of flexibility of experimental design but were abandoned because plants did not grow well on them and lodged easily. Accumulation of solids did not occur and, thus, moveable frames were not necessary for the purpose of flushing away solids.

Natural Marsh Study

Concurrent with the artificial marsh work in 1974 and 1975, observations were carried out on the Brillion Marsh with the idea of eventually deciding whether using existing natural marshes or constructing artificial ones would be the most appropriate (Section XI).

CALENDAR

Four full summer growing seasons (1972-1975) were consumed by the project. In 1972 the study site was readied for work. The summer of

1973 was consumed by preliminary sampling and was regarded largely as a period of establishment of vegetative communities in the plastic-lined excavations. Probably a longer start-up time should be used but the summers of 1974 and 1975 were regarded as producing data typical of established artificial marsh communities.

SECTION IV

METHODS AND MATERIALS

CHEMICAL AND MICROBIOLOGICAL WATER QUALITY

In all cases U.S. Environmental Protection Agency methods were followed (EPA, 1971) where applicable. Standard Methods (APHA, 1971) was also used. Samples collected at the field sites (Sections V and XI) were transported in ice chests to the laboratory where they were analyzed within 24 hours. A walk-in refrigerator was used for short-term storage as needed.

Coliform (total) densities were determined using the membrane filter procedure described by APHA (1971). The stannous chloride method was used for orthophosphate phosphorus and persulfate digestion was used to obtain total phosphorus. One experiment comparing the results of the persulfate digestion with results of the sulfuric acid - nitric acid digestion gave considerably higher results for the latter, more rigorous digestion (Table 1). The more rigorous digestion, however, was not routinely used because of the hazardous nature. Analyses of sediments and plant tissues are discussed below. The biochemical oxygen demand (BOD) test used the standard five-day incubation period.

Suction Sampling Device

In order to withdraw samples of water from known, discrete depths in the pilot plant, pipes of various lengths were permanently inserted into

Table 1. PHOSPHORUS CONCENTRATION AS DETERMINED BY THREE METHODS^a (mg 1-1)

	Sampling Location		
Method	Influent to Ponds	Effluent Bed 1	Effluent Bed 5
SnCl ₂ (Orthophosphate)	34.78	23.17	22.19
Persulfate (Total Phosphorus)	48.51	23.37	21.84
H ₂ SO ₄ -HNO ₃ (Total Phosphorus)	60.21	24.41	24.57

aAPHA, Standard Methods, 13th ed. 1971.

the gravel. Water was withdrawn from them with the suction sampling device shown in Figure 3. The apparatus was rinsed with alcohol prior to collection of samples for microbiological analyses.

Analysis of Data

Direct comparison of concentrations of a material in the influent and effluent of experimental basins or pilot plant leads to erroneous conclusions about treatment efficiency. A significant fraction of the water flowing through a basin is lost from the system by evapotranspiration thus resulting in a tendency toward increasing the concentration of a material rather than decreasing it. To combat this problem we have avoided comparing concentrations as a means of judging results in most instances. Instead, we have compared the number of grams of a substance entering a basin via the influent to the number of grams leaving via the effluent. The difference is the number of grams removed by the system and can be expressed as a percent reduction as follows:

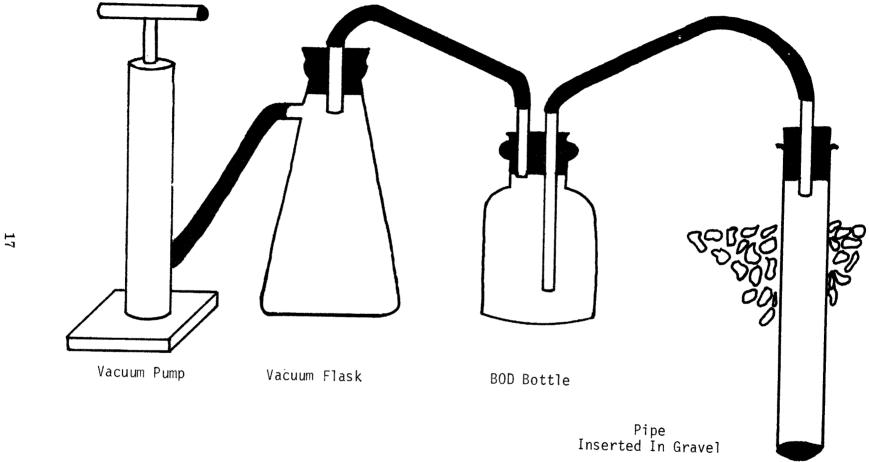


Figure 3. Sampling Device for Obtaining Water Samples from Desired Depths in Pilot Plant.

Known volumes of influent and effluent were multiplied by mean concentrations of a material to obtain values for grams in and grams out. For other kinds of data ($\underline{i}.\underline{e}$. biomass, flow rates, \underline{etc} .) no special techniques are required.

LIVING MATERIALS

Experiments on propagation techniques and on substrate suitability are described in Section VI. Actual handling of living materials was as follows. During construction of the experimental basins, rhizomes of Scirpus acutus and S. validus and tubers of S. fluviatilis were obtained locally and spread on the university campus lawn. They were covered with severed tops and kept moist until new shoots developed. The plants were then transplanted into the experimental basins. Iris were collected and placed directly into the experimental basins after excising about half of the leaf material to reduce transpiration and induce formation of new shoots.

Planting was done in September 1972. Half of the plants of each species were tied onto the wire-covered frames in four of the basins. The other half were planted in pea gravel in four basins. The two remaining basins served as controls (Figure 4):

Basin #1 - control, no plants.

Basin #3 - Softstem bulrush, 540 rhizome sections having 4-5 shoots per clump.

- Basin #5 Hardstem bulrush, 150 root clumps having new shoots.
- Basin #7 River bulrush, 540 10-14 in tall sprouts already beginning to become chlorotic; condition marginal.
- Basin #9 <u>Iris</u>, 540 roots, each having a sheath of 4-10 healthy new leaves.

The remainder of the plants were tied on the wire racks with nylon cord as follows:

- Basin #2 control, no plants.
- Basin #4 River bulrush, 1024 tubers with chlorotic shoots.
- Basin #6 Hardstem bulrush, 480 root clumps having new shoots.
- Basin #8 Softstem bulrush, 480 rhizomes sections having new shoots.
- Basin #10 Iris, 540 roots, each having 4-10 new leaves.

In an attempt to ward off damage by early frost the basins were covered with polyethylene supported by wood ridge poles. The transparent covers had the effect of raising the temperature by several degrees. Floods then inundated the basins on September 29, severely damaging the basins and uprooting many of the plants. The plants were re-set and some data taken, but it was obvious that little could be accomplished before winter. Inasmuch as the plants were not yet established and had little opportunity to store adequate reserves for over-wintering, we decided (November 10) to transplant the Iris, softstem, and hardstem from the pea gravel to the greenhouse to maintain them until spring. The river bulrush appeared to be dead, and all plants were left in the pond. The materials on racks were moved into basins that still held water and were covered with one foot of water until spring.

PHOSPHORUS DISTRIBUTION AND REMOVAL BY HARVESTING

Plant Harvesting Techniques and Laboratory Analyses

For the purposes of nutrient removal, the emergent plant shoots were harvested from each of the basins and Brillion Marsh by cutting with a hand sickle at about 20 cm above the substrate surface (above the intercalary meristem). The samples were placed into pre-weighed plastic bags and sealed. Fresh weight was measured in the laboratory. Then 100 g of intact shoots were cut into small sections, dried at 60 C, ground in a Wiley Mill over a #20 mesh screen and stored. Aliquots of ground tissue were analyzed for dry weight, organic weight, and phosphorus content (AOAC, 1960).

Plant Harvesting Quadrat Selection and Sizes

Experimental Basins --

Quadrats were selected for harvesting by tossing a golf ball over the shoulder. The point of impact became the NE corner of a $0.093~\text{m}^2$ (1 ft²) quadrat. The entire basin was harvested and aliquots taken for analyses at the end of the growing season and when standing crops were very small.

Pilot Plant --

Quadrats were selected as for the experimental basins, but 1 $\rm m^2$ samples were taken. Three quadrats were harvested from each of the three sections of the pilot plant.

Natural (Brillion) Marsh --

Two permanent 1 m² quadrats were established in typical vegetation near the stream at each end of the marsh (Figure 8, Quadrats 1-4). At the upriver end of the marsh, both quadrats were located in nearly unispecific stands of Typha. At the downstream end of the marsh, there was a 15 - 30 m band of Sparganium adjacent to the stream and a wide expanse of Typha extending laterally. One quadrat was established in the Sparganium and the other in Typha. These quadrats were harvested at various intervals during the growing season and were referred to as experimental quadrats. At the end of each growing season in 1974, and 1975, a similar quadrat, referred to as a control quadrat, was harvested near each experimental quadrat.

Substrate Core Sampling Techniques and Methods of Analyses

Experimental Basins --

The golf-ball-over-the-shoulder technique was also used in locating the sites for coring. The point of impact became the center of a 12.8 cm (5 in) diameter core taken through the gravel to the plastic liner. The coring device was a two pound coffee can with both ends removed. The fibrous plant rhizomes were severed by slicing carefully around the perimeter of the device with a round-ended kitchen knife with a serrated edge. The coffee can was inserted until it came firmly in contact with the plastic liner, then the contents were carefully removed by hand. Before removing the sampler, a volume of fresh pea gravel equal to that

removed was placed in the core hole. Three cores were taken from each basin on each sampling date.

Pilot Plant --

Several coring devices were designed and tested, but all either failed to recover the large sized gravel, or were so designed as to cause concern that the plastic liner in the basin would be ruptured. After all the experiments were terminated (December 5, 1975), three 15 cm deep cores were taken with the coffee can from each section in the same fashion as for the experimental basins. These samples included all of the unharvested shoots and the rhizomes and about 75% of the roots. Then, a single excavation was made in the center of each section and a 15 cm core taken in the middle of the 30-60 cm depth. The latter sample was used to represent both the 30-45 and the 45-60 cm interval.

Natural (Brillion) Marsh --

Cores were taken of the organic sediments available to the plants (uppermost 45 cm) with a 3.5 cm piston type coring device on August 5, 1975.

Cores were taken in midstream at each of the water sampling stations (Figure 8), in each of the plant harvesting quadrats, in midstream just above the entrance of the channel from the Brillion sewage treatment plant, from the channel immediately below the 50 m below the sewage treatment plant, and from a secondary influent stream (Deer Creek Run) which receives runoff from a golf course and farmlands. The cores were stored frozen in their plastic core-liner tubes until analysis. Where

the cores were more than 15 cm long, aliquots from the top, middle, and bottom, of each core were analyzed. If less than 15 cm of organic sediments were present, replicate cores were taken and a single aliquot analyzed from each. Only available phosphorus was determined. The extraction method was that of Olsen and Dean (1965); and the analytical methods for orthophosphate was the stannous chloride method of APHA (1971).

For the cores from both the experimental basins and the pilot plant, the plant roots and rhizomes were carefully separated from the gravel and other materials by hand. The gravel was then repeatedly washed with small volumes of distilled water until a total of 2 liters of eluate were collected. Eluate was then analyzed for dry matter, organic content and total phosphorus according to APHA (1971).

SECTION V

PHYSICAL FACILITIES

Field work was conducted on private property adjacent to the municipal treatment plant at Seymour, Wisconsin (population 2,257, Outagamie County). This site is 41 miles from the University of Wisconsin-Oshkosh campus where laboratory analyses were performed. Major features of the field facility were plastic-lined excavations in which vegetation was planted. These excavations were regarded as artificial marshes though no attempt was made to simulate natural marsh conditions. Design of the facility was done by a professional engineering firm. Construction was done in 1972 by a contractor after competitive bidding.

During operation, wastewater (primary or secondary effluent) was with-drawn at the municipal treatment plant and fed to the experimental basins after passing through a screen (in a 55 gallon drum) and an additional settling basin (modified stock tank, Figure 4). The screen and settling basin were not part of the original design, but were added later in 1973 in an attempt to alleviate some clogging problems associated with obtaining uniform flow. In 1973, the constant head tank was used to provide gravity flow to the basins. Uniform flow could not be sustained so, in 1974, the constant head tank was bypassed and a pump provided pressure. Meters were installed on feedlines and effluent pipes in all basins. Wastewater from basins was collected in

a common wet-well and pumped back to the municipal treatment plant to complete the pathway.

Basins were lined with black 20 mil PVC plastic to prevent water loss to the soil. They had flat, square bottoms (9.29 m²), sides with 2 to 1 slope, and vertical ends. A separate inlet pipe from the feedline entered each basin. Flow rates into the basins were adjusted by manually opening and closing the valves. Adjustments of flow rates and depth were the means of altering retention time. Effluent pipes were placed at depths of 15 cm, 45 cm, and 60 cm so that lower ones could be capped in order to increase water depth (Figure 5). When water depth of 0.5 m was desired it was achieved by using the bottom (6 cm) effluent pipe with a vertical standpipe attached to it. Thus, withdrawal was from the gravel instead of the surface.

In the first phase of experimental work (1973) ten basins were used. The arrangement in two rows of five basins each (Figure 4) was not experimentally significant but was a functional convenience. Pea gravel was placed to a depth of 15 cm in the bottom of each offnumbered basin to serve as a medium for roots to penetrate. Frames covered with hardware cloth were placed in even-numbered basins for the attachment of plants.

At the beginning of 1974, the use of wire-covered frames was abandoned and the even numbered basins (Figure 4) were consolidated into a single, long trench by removing partitions between them and installing a new

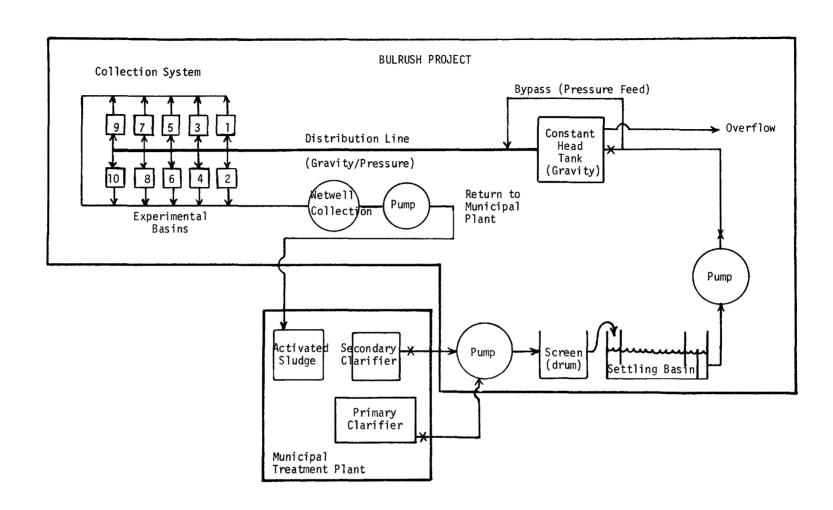


Figure 4. Schematic View of Municipal Treatment Plant and Experimental Facilities. Experimental Basins are Numbered 1 - 10.

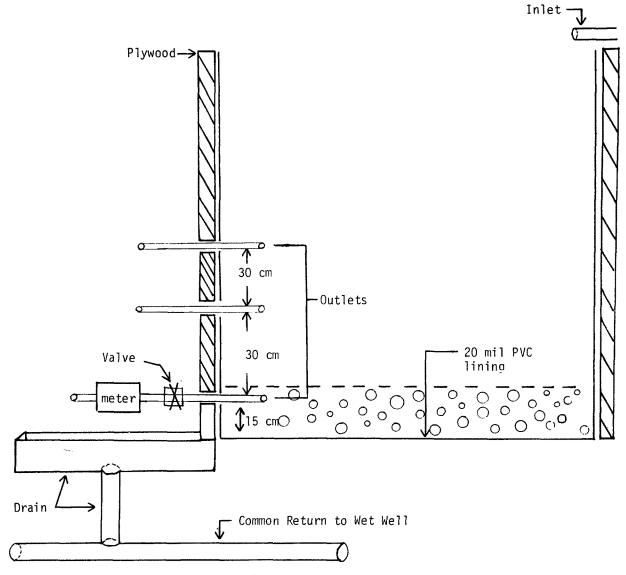


Figure 5. Diagramatic Cross Section of Experimental Basin (not to scale) Showing Relative Positions of Inlet and Outlets.

lining. This trench, the pilot plant, was 19.3 m long, 3.05 m wide at the bottom and 5.8 m wide at the top. Sides had a 2 to 1 slope as in experimental basins. A 15 cm layer of sand was put down on the bottom followed by 30 cm of coarse gravel (1.9 - 2.5 cm) then by 30 cm of pea gravel. Two baffles divided the pilot plant into three sections of equal length (Figure 6).

Influent entered the pilot plant via three pipes spaced uniformly across one end. Each pipe was a branch of a metered feedline and had a valve for flow regulation. In order to sample water at 15, 30, 45, and 60 cm, pipes of appropriate length were driven into the gravel (Figure 6) in each of the three longitudinal sections. Effluent was sampled at metered outlets in the wall at the downstream end. A four-inch drain tile line was placed across the downstream end at the bottom layer of coarse gravel. A one-inch pipe passed from the drain tile out of the pilot plant, through a flow meter to the drain. Outlets at other depths did not carry water from drain tiles. A metered overflow was provided to take off the extra water during rainstorms.

Three Rhodamine-B dye studies were conducted (July 13, July 21, and August 12, 1975) to observe hydrologic characteristics of the pilot plant. In each case, dye solution was added at the influent and samples for fluorometric readings were taken in horizontal and vertical series. In the first study, dye appeared in the effluent after 10 hours indicating surface flow. Low readings were observed at depths

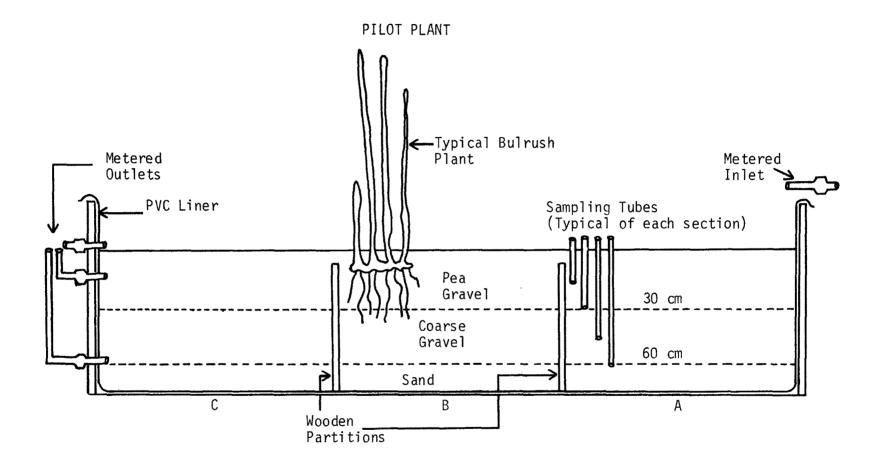


Figure 6. Diagramatic Cross-section of Pilot Plant (not to scale) Showing Relative Positions of Inlet, Outlets, and Sampling Tubes.

of 30 and 45 cm (Appendix A, Figure A-1). Prior to the second study the water level was lowered by about 2.5 cm so that no surface flow occurred. The second study indicated low flow rate in the pea gravel (zero reading at 15 cm). Dispersion into the coarse gravel was quite uniform horizontally and variable but without a distinct pattern vertically (Appendix A, Figure A-2). In spite of the uniform dispersion no dye was actually observed in the effluent. The uniform dye dispersion could not be reconciled with the distinct chemical stratification indicated by very high conductivity at 60 cm. Thus, the partitions (Figure 6) were removed to attempt to prevent stagnation. After removal of the partitions another Rhodamine-B study, this time with double the amount of dye, was done. Results were the same as in the second study. Failure to observe dye in the effluent (Appendix A, Figure A-3) could possibly be attributed to adsorption. Removal of the partitions produced a long-term improvement in the conductivity stratification but no change in dye distribution.

SECTION VI

SUBSTRATE SELECTION AND PROPAGATION EXPERIMENTS

A series of studies was undertaken at the beginning of the project in order to learn how to vegetatively propagate native plants which were likely to be useable for biological waste treatment. Criteria used in selection of species for experimental work included: 1. local availability; 2. ease of vegetative propagation; 3. rate of regrowth after harvesting; 4. tolerance to repeated harvesting. Literature reports on some native species were available. Importance of criteria 1 and 2 above is apparent. Response to harvesting was regarded as important because of the idea that nutrient removal could be achieved by repeated harvesting. Forty-two different experiments involving 1,254 individual propagules were conducted using the following species: Typha latifolia L. (cattail); Sparganium eurycarpum Engelm. (mace reed or burr reed); Phragmites australis (Cav.) Steudal (reed grass); Scirpus fluviatilis (Torr.) Gray, (river or three-square bulrush); Scirpus acutus Muhl., (hardstem bulrush); Scirpus validus Vahl. (softstem bulrush); Iris versicolor L. (wild iris). Since the most desirable kind of planting medium or substrate was not known, several kinds were tried (Table 2) in combination with varying moisture conditions. The goals of the experiments were to learn how to propagate the plants and to choose a substrate for use in the later phase of the work. Response to

harvesting was determined from literature or previous experience as well as by observation.

It was determined that greatest success in propagation was realized when propagules were planted with shoots exposed to the atmosphere. Complete immersion of propagules in water inhibits new shoot development. Special treatment such as gibberellin or scarification of tubers was not effective.

The conclusion regarding choice of substrate was that it should be porous. Pea gravel was chosen for use in later work. The basis for the choice was that pea gravel was easy to work with, would allow water to flow through it, and did not present the biological or chemical unknowns of some other substrates. Sparganium and Typha were considered not suitable for the project because they exhibited limited intercalary growth after harvesting. Intercalary growth is the kind shown by ordinary grass after mowing a lawn. The response of Sparganium and Typha to harvesting is to produce new shoots rather then lengthen the stubs. The time required to produce new shoots constituted an undesirably long delay. Phragmites may be acceptable but would have been impractical to obtain for the project since it grows in 1.5 - 2.0 m of water. The tough fibrous, tangled mats of rhizomes in deep water make hand digging very difficult.

<u>Scirpus validus</u> and <u>S. acutus</u> were judged the best candidates because they are easy to obtain, propagate easily, and regrow rapidly after harvesting. They also are morphologically similar to <u>S. lacustris</u> which has been used for similar studies in Europe.

Table 2. GREENHOUSE EXPERIMENTS ON PLANT PROPAGATION.

Exp.	Species	Substrate	Water depth above subs.	Type propagule	Success	Notes
1	River	Pea gravel	7 cm	3" root clump	(5/9) 55%	Several tops ex- posed
2	River	Pea gravel	7 cm	Tubers	(2/6) 33%	
3	Hardstem	Pea gravel	7 cm	Rhizomes	(0/12) 0%	
4	River	Pit run gravel	7 cm	3" root clump	(5/6) 83%	Several tops ex- posed
5	River	Pit run gravel	7 cm	Tubers	(3/8) 38%	
6	Hardstem	Pit run gravel	7 cm	Rhizomes	(0/12) 0%	
7	Hardstem	Pit run gravel	7 cm	3" root clump	(6/20) 30%	17 shoots/ 6 clumps
8	Phrag- mites	Pit run gravel	45 cm	3" root clump	(9/9) 33%	Cut above water line
9	Hardstem	Pit run gravel	45 cm	Rhizome	(0/8) 0%	
10	Hardstem	Pit run gravel	45 cm	3" root clump	(0/8) 0%	
11	River	Pit run gravel	45 cm	3" root clump	(2/6) 33%	Intact plants
12	River	Pit run gravel	4 5 cm	Tubers	(0/12) 0%	
13	River	Pea gravel	l cm below surf		(35/85) 41%	
14	River	Pea gravel	l cm below surf		(9/9) 100%	21 shoots/ clumps/ several tops in air

Table 2 (continued). PLANT PROPAGATION.

Exp.	Species	Substrate	Water depth above subs.		Success	Notes
15			l cm below surf.			3-5 cm sections
16	River	Pea gravel	l cm below surf.	Tubers	(5/23) 22%	
17	Phrag- mites	Pea gravel	1 cm below surf.	Rhizomes	(0/100) 0%	·
18	Softstem	Pea gravel	l cm below surf.	Rhizomes	(27/83 32%	63 shoots/ 27 Rhizomes
19	River	Organic soil	Field moisture	Tubers	(25/44) 57%	Shoots emerging at transplant
20	River		Field moisture	Tubers	(12/44) 27%	No shoots at transplant
21	Softstem	Organic soil	Field moisture	Rhizome	(14/45) 31%	Laid on surface
22	Spargan- ium	Organic soil	Field moisture	Root- shoot base	(0/18) 0% e	Laid on surface
23	Spargan- ium	Organic soil	Field moisture	Root- shoot base	(0/36) 0% e	Planted upright
24	<u>Typha</u> latifolia	Organic soil	Field moisture	Root- shoot base	(1/18) 6% e	Upright
25	<u>Typha</u> <u>latifolia</u>	Organic soil	Field moisture	Root- shoot base	(2/9) 22% e	Laid on surface
26	Softstem	Organic soil	Field moisture	Rhizome	(45/100) 45	tops above
27	Spargan- ium	Organic soil	Field moisture	Root- shoot base		soil
28	<u>Typha</u> <u>latifolia</u>	Crushed lime- stone	l cm below surf.	Root- shoot base	(2/30) 7% e	

Table 2 (continued). PLANT PROPAGATION.

Exp.	Species	Substrate	Water depth above subs.		Success	Notes
29	Spargan- ium	Crushed lime- stone	l cm below surf.		(1/50) 2%	
30	Spargan- ium	Crushed lime- stone	1 cm below surf.	Root- shoot base	(1/40) 2½% e	
31	Softstem	Crushed lime- stone	1 cm below surf.		(12/12) 1009	6
32	Softstem	Crushed lime- stone	l cm below surf.		(1/3) 33%	20-45 cm rhizomes
33	3 square	Pea gravel	l cm below surf.	Tubers	(8/21) 38%	10 ppm gibberellin
34	Softstem	Pea gravel	l cm below surf.	Rhizomes	(10/19) 52%	10 ppm gibberellin
35	Spargan- ium	Pea gravel	1 cm below surf.		(8/24) 33%	10 ppm gibberellin
36	<u>Iris</u>	Wire racks	at surface	Root- shoot base	(26/66) 40%	
37	River	Wire racks	at surface	Tubers	(16/20) 80%	Growth at transplant
38	River	Wire racks	at surface	Tubers	(41/94) 44%	No growth at trans- plant
39	Softstem	Wire racks	at surface	Rhizomes	(11/19) 58%	
40	Spargan- ium	Wire racks	at surface	Root- shoot base		
41	<u>Typha</u> <u>latifolia</u>	Wire racks	at surface	Root- shoot base	(6/6) 100% e	

Table 2 (continued). PLANT PROPAGATION.

Exp. Species	Substrate	Water depth above subs.		Success	Notes
42 <u>Iris</u>	Pea gravel	at surface	Root- shoot base	(44/54) 72% e	<u>(</u>
TOTAL				391/1254 31.2%	

<u>Iris</u> are easy to transplant but do not recover quickly from cutting and, as they are not usually found in large clones in nature, it is difficult to secure large quantities for transplanting. <u>Iris pseudacorus</u> has been used in Europe, however, to "remove" coliform bacteria, so the genus was included in this study.

Of the seven species examined in culture technique experiments, four were chosen for scrutiny under wastewater loading conditions. They were: Scirpus acutus, S. validus, S. fluviatilis, and Iris versicolor.

SECTION VII

GREENHOUSE STUDIES

Field studies did not begin as soon as anticipated at the beginning of the project (summer 1972), so greenhouse experiments were undertaken as a follow up to the propagation experiments discussed in Section VI. The major goal was to see which of the four species selected in the preliminary work was most effective at purification of wastewater. Other goals included obtaining additional information about responses of the plants to various treatments and determining what retention times should be attempted in the field.

A large tray in the greenhouse was divided into five compartments or beds and then lined with plastic. The beds were 80 cm x 90 cm x 12 cm. Pea gravel having a porosity of 40% was placed to a depth of seven cm in each bed. Each bed had a collection well to facilitate draining and sampling. The beds were numbered and planted as follows: Bed 1 - Iris; Bed 2 - control (no plants); Bed 3 - hardstem bulrush; Bed 4 - softstem bulrush; Bed 5 - softstem bulrush. The beds were batch fed chlorinated primary effluent collected once or more per week at the Oshkosh, Wisconsin (Winnebago County) municipal wastewater treatment plant. Effluent was stored temporarily in a refrigerated vault and measured amounts were added to the beds daily Monday through Friday. Prior to feeding each day an amount of water was drained from each bed so that

the water level after feeding should be flush with the gravel surface. Each bed had a capacity of 21 liters.

An experiment was conducted to see how the quality of effluent varied with retention time. Analyses were performed twice weekly on influent and effluent for retention times of 1.5, 3, and 5 days. Parameters included BOD, COD, pH, conductivity, orthophosphate, and total phosphorus. Evapotranspiration losses were estimated by determining the difference between the influent and effluent volumes. Since day length is short in winter, artificial light was employed (in addition to natural light) 24 hours per day. At night the light intensity was 200 - 300 foot-candles at the gravel surface. To begin the experiment, beds were allowed a colonization period (month of December 1972) during which time microcommunities could also become established. Sampling was initiated on January 2, 1973, and continued twice each week until March 26, 1973, with four weeks allocated to each of the three retention times.

Results of the retention time experiments were judged by comparing the total output (of phosphorus for example) from a bed with the total input during a time period as described in Section IV. Detailed results of analyses are presented in Appendix B. Table B-1 gives the number of liters of influent and effluent as a basis for computing total amounts of constituents. Table 3 shows a summary of the effects of the green-house beds on the wastewater treated by them. A significant fraction of water (about 60%) was lost by evapotranspiration (Appendix B, Table B-1). Effluent quality was obviously best with 5-day retention.

Table 3. EFFECTIVENESS OF SEVERAL SPECIES OF PLANTS AT TREATMENT OF PRIMARY EFFLUENT IN THE GREENHOUSE.

	Retentior time)		Percent	reduction	
Parameter	days	Iris	Control	Hardstem	Softstem	Softstem
BOD	5	98	97	98	98	97
	3	94	92	92	96	95
	1.5	89	90	90	88	81
COD	5	89	89	87	86	83
	3	73	75	89	85	82
	1.5	49	54	61	57	29
Orthophosphate	5	58	52	68	75	70
	3	8	2	41	63	49
	1.5	-3	-1	28	57	30
Total phosphorus	5	80	76	84	83	80
	3	34	-4	45	62	54
	1.5	30	32	59	60	51
Coliform	5 3 1.5	68 7	80 22	83 15	88 60	 89 51
Total solids	5 3 1.5	32 17	35 24	35 5	25 -5	25 -14
Suspended solids	5 3 1.5	61 39	 66 54	69 22	 59 16	57 -15
Dissolved solids	5	54	60	60	51	44
	3	21	28	25	13	13
	1.5	13	19	2	-9	-14

Increasing retention time from 3 to 5 days improved effluent quality much more than increasing from 1.5 to 3 days. Phosphorus removal, BOD reduction, and coliform die-off were regarded as the most important parameters upon which to judge results but coliform analyses were not

performed in all cases for this experiment. All beds, including the control, reduced BOD by a high percent (97-98%). The BOD of the effluent (15-22 mg 1⁻¹, 5-day retention) appears high until corrected for evapotranspiration. Phosphorus removal followed a similar pattern. Seventy-six to 83% of total phosphorus was removed with 5-day retention. No pronounced difference appeared in the treatment efficiency of the beds, including the control, if total phosphorus removal and BOD reduction are considered. This suggests that if the vegetation actually plays a role in waste treatment, the effect may not be observable until retention times are lengthened beyond 5 days.

SECTION VIII EXPERIMENTAL BASIN STUDIES

QUALITY OF EFFLUENT

Preliminary Study

The experimental basins described in Section V were used as biological systems to treat secondary effluent starting in summer, 1973. Goals included determining whether water quality was significantly improved, working out flaws in the physical facilities, allowing vegetation to become established, and observing responses of species to various treatments. Samples of influent and effluent were collected at one week intervals beginning in the late summer after elapse of a growth period. Results are presented in Table 4. For this time period conclusions are based on comparing concentrations of constituents in influent and effluent. One of the mechanical problems was with flow regulation. Thus accurate data on volumes of influent and effluent were not obtained. As a result, the mass balance flux could not be computed. Furthermore, retention time estimates are meaningless causing data to be of value mainly for deciding where the emphasis should be placed in later parts of the project.

For the ten basins collectively, BOD reduction did not show impressive differences. The control and one experimental basin gave less than

4

Table 4. SUMMARY OF PRELIMINARY DATA ON EFFECTIVENESS OF EXPERIMENTAL BASINS FOR TREATING SECONDARY EFFLUENT. AUGUST - OCTOBER 1973.

		$\overline{\mathbf{x}}$	BOD			\overline{x}	COD			x To	tal so	1 i ds	
<u>Substrate</u>	Species	mg 1-1	S.D.	n	% red.a	mg 1-1	S.D.	n	% red.	mg 1-1	S.D.	n	% red.
	Influent	18	5.8	8		33	6.2	5		770	40	3	
Gravel	Control	iĭ	7.6	8	39	47	17.3	4	-42	750	10	3 2	3
Wire	Control	9	5.8	8	50	36	6.6	4	9	730	50	4	5
Grave1	Hardstem	9	4.5	8	50 50	27	13.3	5	18	730	70	4	5
Wire	Hardstem	8	5.1	8	56	27	10.7	5	18	790	50	4	-3
Grave1	Softstem	9	4.8	8	50 50	29	11.5	4	12	730	20	3	5
Wire	Softstem	6	4.1	8	67	29	6.5	3	12	740	50	3	1
Gravel	River	7	3.0	8	61	33	14.0	5	0	7 7 0	40	4	0
Wire	River	9	3.4	8	50	35,	18.9	4	6	760	70	4	1
Gravel	Iris	9	3.6	6	50 50	31	9.9	3	6	690	20	2	10
Wire	Iris	12	5.9	7	33	33	1.3	2	0	750	50 50	4	3
wire	11.12	12	5.9	,	33	33	1.3	2	U	/50	50	4	3
	**************************************	X	Conduct	ivit		x Ort	hophosp	hate		x Tota		p hor i	ıs
Substrate	Species	umhos	S.D.	n	% red.	mg 1-1	S.D.	n	% red.	mg 1-1	S.D.	n	% red.
	Influent	1040	140	10		16.4	6.9	8		14.2	3.1	8	
Grave1	Control	943	168	9	9	12.1	4.6	8	26	13.1	4.1	8	8
Wire	Control	943 925	129	10	11	14.3		8	13	13.1	3.8	8	5
		1014	85	10	3	15.6	5.1	8	5	14.1	3.7	8	1
									i.i	14.1	J./	-	,
	Hardstem										20	0	7
Wire	Hardstem	927	165	10	11	12.4	4.2	8	24	13.2	3.8	8	7
Gravel Wire Gravel	Hardstem Softstem	927 973	165 148	10	11 6	12.4 13.8	4.2 3.2	8 7	24 16	13.2 14.2	3.2	7	7 0
Wire Gravel Wire	Hardstem Softstem Softstem	927 973 978	165 148 143	10 9 8	11 6 6	12.4 13.8 12.6	4.2 3.2 3.2	8 7 6	24 16 23	13.2 14.2 13.4	3.2 3.9	7 6	6
Wire Gravel Wire Gravel	Hardstem Softstem Softstem River	927 973 978 1054	165 148 143 90	10 9 8 9	11 6 6 -1	12.4 13.8 12.6 15.0	4.2 3.2 3.2 2.7	8 7 6 8	24 16 23 9	13.2 14.2 13.4 15.1	3.2 3.9 2.6	7 6 8	6 -6
Wire Gravel Wire Gravel Wire	Hardstem Softstem Softstem River River	927 973 978 1054 988	165 148 143 90 110	10 9 8 9	11 6 6 -1 6	12.4 13.8 12.6 15.0 13.6	4.2 3.2 3.2 2.7 3.7	8 7 6	24 16 23 9 17	13.2 14.2 13.4 15.1 15.4	3.2 3.9 2.6 5.3	7 6	6 -6 -8
Wire Gravel Wire Gravel	Hardstem Softstem Softstem River	927 973 978 1054	165 148 143 90	10 9 8 9	11 6 6 -1	12.4 13.8 12.6 15.0	4.2 3.2 3.2 2.7	8 7 6 8	24 16 23 9	13.2 14.2 13.4 15.1	3.2 3.9 2.6	7 6 8	6 -6

apercent reductions are not adjusted for water-loss by evapotranspiration. They are based on comparison of concentrations because accurate flow data could not be obtained.

Table 4 (continued). EFFECTIVENESS OF EXPERIMENTAL BASINS FOR TREATING SECONDARY EFFLUENT.

		x	Turb	dity		log col	Colifo	rm			рН		
Substrate	Species	JTU	<u>S.D.</u>	n	% red.ª	100m1-1	S.D.	n	% redb	рН	S.D.	<u>n</u>	
	Influent	7.57	3.9	11		5.24	5.48	6	w	7.55	.07	11	
Gravel	Control	6.93	4.1	ii	8	4.78	4.91	6	66	8.09	.39	ii	
Wire	Control	6.85	5.2	11	10	4.86	4.94	6	59	8.01	.62	11	
Grave1	Hardstem	4.84	3.1	11	36	4.62	4.79	6	76	7.45	.08	11	
Wire	Hardstem	4.98	3.1	1.1	34	5.07	5.15	6	36	7.85	.46	11	
Gravel	Softstem	4.73	2.5	10	38	4.81	4.87	5	63	7.34	.09	10	
Wire	Softstem	6.47	4.1	9	1 5	5.02	5.33	5	40	7.48	.14	9	
Grave1	River	3.76	1.4	10	50	4.29	4.48	6	89	7.62	.16	10	
Wire	River	5.12	3.6	10	32	5.14	5.29	6	21	7.61	.11	10	
Gravel	Iris	9.33	9.1	8	-23	4.80	4.62	5	64	8.22	.58	8	
Wire	Iris	5.55	2.2	10	27	4.80	4.95	6	65	7.47	.15	10	

aPercent reductions are not adjusted for water-loss by evapotranspiration. They are based on comparison of concentrations because accurate flow data could not be obtained.

bIn this case the percent reduction is computed by comparing number of colonies per 100 ml, not by

comparing logarithms.

40% reduction while all the others gave 50% or more and the high was 67% by softstem (Table 4). Total phosphorus removal was very low ranging from 11% to an apparent increase. Such increases are referred to as negative removals (Table 4). Three basins showed zero or negative total phosphorus removal. A control basin ranked second best at phosphorus removal. Other parameters showed similarly unimpressive results, but working out solutions to mechanical problems dominated the summer's work and it was felt that progress in that area would allow greatly improved control of conditions in the following growing season. Correction of data for water loss by evapotranspiration would give more realistic results.

Subjective observations coupled with the observed good BOD reduction in the field and phosphorus removal in the greenhouse led to the choice of softstem bulrush as the most promising species. Softstem displayed greatest ease of propagation and most rapid and vigorous regrowth after harvesting. Tissues of softstem did not contain a distinctly higher percent phosphorus than other species, but it appeared that in future work softstem would produce the most biomass per unit time. Iris and river bulrush showed extremely poor recovery after cutting and would not have been possible to use in a project requiring repeated harvesting. Fastening plants to wire-covered frames proved to offer no advantage and, in fact, to be unnecessary. Although on some occasions, frames with vegetation in place were moved from one basin to another; the need to do so was not related to the waste treatment processes of the biota.

Solids did not accumulate on roots as anticipated so the need to back-wash did not arise. A distinct drawback to the frames was that they did not provide adequate support. Tall vegetation lodged easily leading to self-shading and decomposition. For these reasons and for the sake of convenience, future work did not employ frames.

Literature review and the low level of water quality improvement achieved initially pointed to the desirability of a major design change. Five of the basins were modified by removing partitions to form a single trench (Section V) which was expected to offer a hydrologically more suitable situation. The trench (pilot plant) was planted with softstem.

TREATMENT OF SECONDARY EFFLUENT

At the beginning of the growing season in 1974, flow meters were installed on the inlet and outlet pipes of five experimental basins so that careful monitoring was possible. By this time, operation of physical facilities had become smooth and dependable. Studies were conducted on some or all of these basins during 1974 and 1975, simultaneously, with work on the pilot plant. As a routine matter, samples were collected weekly during the growing season in order to evaluate long term effectiveness of treatment of secondary effluent. Weekly grab samples, however, can fail to detect important events of diurnal or other short duration. Thus, intensive studies were carried out on several occasions, particularly, to examine the effect of harvesting on water quality.

Long Term Variation in Effluent Quality

In this work two variables were examined for gross influences on quality of effluent produced. Although softstem was quite clearly the choice of species, one basin of river bulrush was studied in addition to a control basin and three softstem basins. Retention time was the second factor. Data are presented in Appendix C, Figures C-1 to C-8.

Variation With Species --

Between June 19 and August 31, 1974, weekly samples showed that the river bulrush basin was distinctly less effective than the other basins at reducing the concentration of BOD, coliforms, and total phosphorus (Table 5). Retention time was five hours. Total phosphorus concentration was reduced 23% by the river bulrush and an average of 31% by three softstem basins (maximum 39%). BOD was reduced 84-91% by softstem but only 62% by river bulrush and 75% by the control. Results obtained by weekly sampling showed a wide range of variability (standard deviations (Table 4). Although improvement in effluent quality was not as great as anticipated it was suspected that perhaps treatment efficiency was considerably better at some times than others and that more rigorous sampling would reveal subtle changes occurring over short periods.

Variation With Retention Time --

Lack of ability to obtain retention time of more than 5 hours was a serious handicap. Success was obtained, however, below 5 hours with no

difficulty and up to 16-17 hours with considerable difficulty. An attempt was made to achieve 10-day retention time. In order to lengthen retention time in this kind of system, either the input rate must be lessened or the volume contained by the basin increased. The first method is the necessary one if wastewater is to be prevented from accumulating and rising above the gravel surface. For a 5-hour retention time an input rate of 1.91 min-1(0.5 gpm) was required. That was about the lower limit because the gate valves plugged with solids at low flow rates. Flow rates down to 0.41 min-1 (0.1 gpm) could be maintained for short periods. The 5-hour and 16-hour retention times were studied in 1974 and 1975, respectively, and the 10-day retention was attempted at the end of 1975.

BOD reduction did not change as retention time increased (Table 6).

Removal was in the range of 71-79% for both control and bulrush basins at each of the three retention times except for 91% removed in the softstem bulrush basin at 5-hour retention. Phosphorus removal was about the same for 5-hour (no-free-standing water) and 10-day (free-standing water) and higher for the 16-hour (no-free-standing water) retention time. The bulrush basin removed 44% of the load it received compared to 81% for the control. But in absolute terms, the bulrush basin removed 0.3 kg compared to 0.4 kg for the control (Appendix C, Table C-3). Thus, the difference between 44% and 81% is misleading. The explanation is in the fact that the load of total phosphorus applied to the bulrush basin was 1.4 times as great as that applied to the

Table 5. EFFECTIVENESS OF EXPERIMENTAL BASINS IN TREATMENT OF SECONDARY EFFLUENT. SUMMER 1974. FIVE-HOUR RETENTION.

	Influent load			Efflu	ent loa	d	% reduc-
	mg 1-1a	1	kg	mg]-1a	1	kg	<u>tion</u> b
BOD							
Control	49.5	63948	3.2	14.3	56514	0.8	75
Softstem	49.5	51045	2.5	10.5	38883	0.4	84
Softstem	49.5	67449	3.3	6.7	60594	0.4	88
River	49.5	75166	3.7	18.4	73909	1.4	62
Softstem	49.5	85911	4.3	4.8	77237	0.4	91
COD							
Control	41.5	63948	2.7	33.0	56514	1.9	30
Softstem	41.5	51045	2.1	29.9	38883	1.2	43
Softstem	41.5	67449	2.8	28.1	60594	1.7	39
River	41.5	75166	3.1	27.5	73909	2.0	35
Softstem	41.5	85911	3.6	17.6	77237	1.4	61
Orthophosphate							
Control	13.4	63948	0.9	10.4	56514	0.6	34
Softstem	13.4	51045	0.7	10.6	38883	0.4	43
Softstem	13.4	67449	0.9	10.4	60594	0.7	23
River	13.4	75166	1.0	11.4	73909	0.9	10
Softstem	13.4	85911	1.2	10.6	77237	0.8	33
Total phosphorus							
Control	13.8	63948	0.9	10.8	56514	0.6	33
Softstem	13.8	51045	0.7	11.1	38883	0.4	39
Softstem	13.8	67449	0.9	12.1	60594	0.7	22
River	13.8	75166	1.1	11.5	73909	0.9	23
Softstem	13.8	85911	1.2	10.4	77237	0.8	33

aConcentrations are averages of values obtained by sampling each week during the growing season.

bWater balance data are given in Appendix C, Table C-5.

control. The application of a greater load to the bulrush basin means that the retention was lessened. In terms of concentration, the influent and effluent did not differ much in most cases. The removal of phosphorus only becomes apparent upon taking water loss by evapotranspiration into account.

Short Term Variation in Effluent Quality

Three studies were conducted in 1974 to examine short term variation in influent and effluent quality. Samples were collected each four hours

Table 6. PERCENT REDUCTION IN BOD, TOTAL SOLIDS AND TOTAL PHOSPHORUS. SUMMARIZED FROM TABLES 5, 12 AND APPENDIX C, TABLES C-3 AND C-4.

Effluent	Retention time		OD Bulr.	Total Cont.	solids Bulr.	Total Cont.	
Secondary	5-hour	75	91	-17	25	33	31
Secondary	16-hour	77	72	57	49	81	44
Secondary	10-day	71	79	43	29	36	32
Primary	10-day	83	87	37	18	36	25

for 72 hours (Appendix C, Figures C-9 to C-15 and Tables C-1 and C-2) for the purpose of determining whether important fluctuations in influent and effluent quality were occurring and to tell, if possible, how the experimental basins were coping with a variable influent. Bulrushes were harvested between the first two studies to see whether harvesting altered the apparent effectiveness of the experimental basins. The third study served as a follow-up designed to detect improvement in treatment efficiency which might result from recovery of the vegetation which had been harvested. Results of the three studies are shown in Tables 7 through 9. Retention time was 5 hours. Excellent control over volume loading rates was possible. Since all sampling was done by hand, the person on the site had only to make a check on flow rates each 4 hours to insure accuracy. Careful control of volume loading rate apparently did not insure uniform outflow rate as would have been

Table 7. EFFECTIVENESS OF EXPERIMENTAL BASINS IN TREATMENT OF SECONDARY EFFLUENT DURING WEEK PRIOR TO HARVESTING. FIVE-HOUR RETENTION.

Parameter	Basin	Inf mg 1-	luent lo l liters		Efflu mg 1-1	Effluent load ^a mg 1-1 liters kg			
BOD	Control	38	7843	0.30	5	7865	0.04	87	
	Bulrush	38	7835	0.30	5	7067	0.04	87	
COD	Control	42	7843	0.33	31	7865	0.24	27	
	Bulrush	42	7835	0.33	36	7067	0.25	24	
Total	Control	23	7843	0.18	18	7865	0.14	22	
phosphorus	Bulrush	23	7835	0.18	21	7067	0.15	17	
Dissolved	Control	916	7843	7.20	930	7865	7.30	-1	
solids	Bulrush	916	7835	7.20	910	7067	6.40	11	

^aConcentrations are averages of values obtained sampling every four hours for four days.

Table 8. EFFECTIVENESS OF EXPERIMENTAL BASINS IN TREATMENT OF SECONDARY EFFLUENT DURING FIRST WEEK AFTER HARVESTING. FIVE-HOUR RETENTION.

Parameter	Inf mg 1-	luent lo l liters		Eff]u mg 1-1	ent loa liters		% reduc- tion	
BOD	Control	65	5352	0.35	9	7059	0.06	83
	Bulrush	65	6022	0.39	5	5840	0.03	92
Total	Control	21	5352	0.11	18	7059	0.13	-18
phosphorus	Bulrush	21	6022	0.13	16	5840	0.09	31
Total	Control	988	5352	5.30	873	7059	6.20	-17
solids	Bulrush	988	6022	5.90	761	5840	4.40	25
Dissolved solids	Control	758	5352	4.10	744	7059	5.30	-29
	Bulrush	758	6022	4.60	687	5840	4.00	13
Suspended	Control	232	5352	1.20	168	7059	1.20	0
solids	Bulrush	232	6022	1.40	75	5840	0.44	69

Table 9. EFFECTIVENESS OF EXPERIMENTAL BASINS IN TREATMENT OF SECONDARY EFFLUENT DURING THIRD WEEK AFTER HARVESTING.
FIVE-HOUR RETENTION.

D	D	-	uent l		_ 1	ent loa		% reduc-
<u>Parameter</u>	Basin	mg 1-	liter	s kg	mg 1-1	liters	kg	tion
BOD	Control	40	6109	0.24	4	4705	0.02	92
	Bulrush	40	640 8	0.26	3	7517	0.02	92
Total	Control	22	6109	0.13	18	4705	0.08	38
phosphorus	Bulrush	22	6408	0.14	18	7517	0.16	-14
Total	Control	860	6109	5.30	829	4705	3.90	26
solids	Bulrush	860	6408	5.50	909	7517	6.80	-24
Dissolved solids	Control	762	6109	4.70	740	4705	3.50	26
	Bulrush	762	6408	4.90	749	7517	5.60	-14
Suspended solids	Control	93	6109	0.57	89	4705	0.42	26
	Bulrush	93	6408	0.60	165	7517	1.20	-100

expected. In both the pre-harvest and the first post-harvest studies, the control basin yielded more water than it received. The bulrush basin yielded more water than it received in the second post-harvest study (Tables 7-9). This finding revealed a greater than anticipated ability of the basins to buffer flow rates which were not carefully checked prior to the beginnings of the intensive studies.

Results of the tests using four-hour sampling intervals showed that the variability of the influent was considerably greater than that of the effluent although the variability was high in the effluent for some parameters. Distinct events changing with time of day could not be identified. A summary of data for the three studies (Table 10) also shows that harvesting the bulrushes did not affect changes in quality

of effluent. In fact, average values of percent reduction scatter widely for all parameters except BOD (83-92%). The negative percent reductions result from the excess of effluent volume over influent volume in some cases (Appendix C, Table C-2). For example, in the second post-harvest study, the total phosphorus concentration was lowered by both the control and the bulrush basin from 22 to $18 \text{ mg } 1^{-1}$. The bulrush basin had 17% more effluent than influent however, so in spite of a reduction of 18% in concentration (22 mg 1^{-1} to $18 \text{ mg } 1^{-1}$), there was more phosphorus leaving the basin than going in during the three-day period.

Table 10. EFFECT OF HARVESTING BULRUSHES ON TREATMENT EFFICIENCY. PERCENT REDUCTIONS ARE SUMMARIZED FROM TABLES 7, 8, and 9.

***************************************	Pre-harve			st-harvest	Second post-harv.		
<u>Parameter</u>	Control	Bulrush	Control	Bulrush	Control	<u>Bulrush</u>	
BOD	87	87	83	92	92	92	
Total phosphorus	22	17	-18	31	38	-14	
Total solids			-17	25	26	-24	
Dissolved solids	-1	11	-29	13	26	-14	
Suspended solids			Q	69	26	-100	

TREATMENT OF PRIMARY EFFLUENT

Primary effluent was treated in one control and in one bulrush experimental basin from August 21 to November 4, 1975, and in the pilot plant for the

same period. Major emphasis was put on the pilot plant.work. The flow regulation difficulty mentioned earlier kept the desired 10-day retention time from being achieved with accuracy. The goal of treating primary effluent in experimental basins was to see whether the system behaved differently than it did with secondary effluent. Unfortunately, it was not possible to achieve 10-day retention time without having standing water and surface flow.

On the mass balance basis, treatment of primary effluent was not as good as treatment of secondary effluent in terms of percent reduction except in the case of BOD (Table 6). BOD reduction was nearly 10% greater in

Table 11. WATER BALANCE DATA FOR EXPERIMENTAL BASINS. 1975. (liters)

Influent	Rainfall	Total effluent	<pre>% evapotrans- piration</pre>
n			
19784	1308	11533	45.3
26866	1308	18887	32.9
21397 34337	2422 2422	13013 20590	45.3 43.9
— 14576 35734	5962 5962	11979 32808	41.6 21.3
	21397 34337	21397 2422 34337 2422	Influent Rainfall effluent On

primary (83-87%) than in secondary effluent (71-79%). Interpretation of this result is complicated by the fact that the load on the bulrush basin

was about 2.5 times as great as on the control basin (Table 11) due to very poor flow rate regulation in this peripheral experiment. Examination of the number of kilograms of BOD removed shows that the bulrush basin removed 2.5 times as much BOD as the control but the percent reductions are nearly identical (Table 12). In the case of secondary effluent, there was a loading differential of 1.6 times as much BOD on the bulrush basin as on the control. In that case the bulrush basin removed 1.8 times as much BOD. For this parameter the bulrush basin distinctly did the best job because it removed the same fraction of the load from the primary effluent as the control in spite of a much heavier load. Roughly, the same number of liters was lost by evapotranspiration from each of the two basins even though one was loaded at a higher rate (Table II). This accounts for the difference between evapotranspiration loss of 42% in the control and 21% in the more heavily loaded bulrush basin.

Phosphorus (total) was not removed to nearly such a great extent as BOD. The control and bulrush basins removed 36% and 25%, respectively. However, the bulrush basin removed 1.7 times as much phosphorus as the control while being loaded with 2.5 times as much (Table 12). Obviously, both basins did not have the same retention time. It seems safe to assume that the bulrush basin would have, at worst, removed phosphorus at the same rate as the control under the same loading rate.

Removal of total solids from primary effluent was less efficient than from secondary effluent. Best total solids removal was achieved when

Table 12. EFFECTIVENESS OF EXPERIMENTAL BASINS FOR TREATMENT OF PRIMARY EFFLUENT. TEN-DAY RETENTION.

Parameter	Basin	Inf] mg 1-1	uent loa liters	ıd kg	Efflue	ent load liters	kg	% reduc- tion
					mg .			
BOD	Control	320	14576	4.70	65	11979	0.78	83
	Bulrush	320	35734	11.40	47	32808	1.50	87
COD	Control	400	14576	5.80	130	11979	1.60	72
	Bulrush	400	35734	14.30	110	328 0 8	3.60	75
Orthophos-	Control	26	14576	0.38	20	11979	0.24	37
phate	Bulrush	26	35734	0.93	20	32808	0.66	29
Total	Control	27	14576	0.39	21	11979	0.25	36
phosphorus	Bulrush	27	35734	0.96	22	32808	0.72	25
Total	Control	1066	14576	15.50	814	11979	9.80	37
solids	Bulrush	1066	35734	38.10	952	32808	31.20	18
Suspended	Control	97	14576	1.40	37	11979	0.44	69
solids	Bulrush	97	35734	3.50	19	32808	0.62	82
Dissolved	Control	969	14576	14.10	778	11979	9.30	34
solids	Bulrush	969	35734	34.60	93 2	32808	30.60	12

wastewater had to pass through gravel (Table 6) even though retention time was shorter. Primary effluent was not treated with a short enough retention time, however, to study solids removal under the no-free-standing-water condition.

SECTION IX PILOT PLANT STUDIES

Wastewater was treated in the pilot plant in 1974 and 1975. The 1974 growing season was allowed to pass mainly as a growth period for the softstem bulrushes. Limited sampling of effluent was conducted in 1974. In 1975, samples were collected weekly at each of four depths at each of three sites along the length of the pilot plant (Figure 6) as well as of the influent. One intensive three-day study was conducted. The partitions dividing the pilot plant into three sections were removed August 6, 1975. Sampling in 1975 was according to the schedule shown in Table 13.

Table 13. 1975 SAMPLING SCHEDULE FOR PILOT PLANT.

Nature of effluent	Depth of discharge
Secondary	60 cm
Secondary (Partitions removed)	15 cm
	60 cm
Primary	60 cm
	Secondary Secondary (Partitions removed) Secondary

Retention time was calculated to have been 10 days (Section V) but the realized time was probably somewhat less. The variables altered in this work were the nature of the wastewater (secondary, primary) and depth at which effluent was withdrawn at the downstream end.

Early results on secondary effluent indicated promising BOD reduction ability (92%) but less favorable total phosphorus removal (35%, Table 14 and Appendix D, Table D-1). Since these figures were taken soon after construction of the pilot plant they were questioned. The following summer, BOD reduction dropped to the 30-38% range for treatment of secondary effluent (Tables 15-17). Removal of the partitions did not change the rate of BOD reduction. Occasional checks at different locations and depths in the pilot plant showed that no dissolved oxygen was present at any time.

Table 14. EFFECTIVENESS OF THE PILOT PLANT IN TREATING SECONDARY EFFLUENT. SUMMER 1974.

Б .		uent loa		Eff]	% reduc-		
Parameter	mg 1-1	liters	kg	mg 1-	liters	kg	tion
BOD	50.0	607227	30.0	4.0	604468	2.4	92
COD	42.0	607227	26.0	24.0	604468	14.5	44
Orthophosphate	13.4	607227	8.1	8.5	604468	5.1	37
Total phosphorus	13.9	607227	8.4	9.0	604468	5.4	35

Phosphorus removal was better than in the experimental basins but not as good as it was in the greenhouse. During July 17 - August 21, 1975, 60-64% of total phosphorus was removed from secondary effluent in two experiments (Tables 16, 17). Distribution of the phosphorus into components of the pilot plant ecosystem is discussed below.

Primary effluent was treated in the pilot plant late in the growing season (beginning August 21). BOD reduction of 77% and total phosphorus removal of 37% were observed (Table 18 and Appendix D, Table D-2). This level of treatment was roughly comparable to secondary treatment given

Table 15. EFFECTIVENESS OF PILOT PLANT IN TREATING SECONDARY EFFLUENT. EFFLUENT DRAWN FROM DEPTH OF 0.60 m. JUNE 5 - JULY 17, 1975.

	4	ent load		. 1	uent load		% reduc-
Parameter	mg]-1	liters	kg	mg]-1	liters	kg	tion
BOD	19	119200	2.3	15	108255	1.6	30
COD	38	119200	4.5	41	108255	4.4	2
Orthophosphate	23	119200	2.7	14	108255	1.5	44
Total phosphorus	24	119200	2.9	15	108255	1.6	45
Total solids	827	119200	98.6	1253	108255	135.6	-38
Suspended solids	70	119200	8.3	147	108255	15.9	-92
Dissolved solids	757	119200	90.2	1107	108255	119.8	-33

by the municipal treatment plant. Secondary effluent from the municipal plant had BOD of 19-50 mg 1^{-1} (Tables 15-17) and total phosphorus of 23-24 mg 1^{-1} . The effluent from the pilot plant had BOD of 72 mg 1^{-1} and total phosphorus of 17 mg 1^{-1} . Based on these concentrations, the pilot plant effluent is higher in BOD and slightly lower in total phosphorus than the secondary effluent. The lower total phosphorus (17 mg 1^{-1}) was not as low as the concentration found in secondary

effluent after passing through the pilot plant (12-15 mg l^{-1}). Numerous data are presented in Appendix D, Tables D-3 to D-7.

Table 16. EFFECTIVENESS OF PILOT PLANT IN TREATING SECONDARY EFFLUENT. EFFLUENT DRAWN FROM DEPTH OF 0.15 m. JULY 17 - AUGUST 6, 1975.

Parameter	Influmg 1-1	ent load liters	l kg	Eff]uo mg 1-	ent load liters	kg	% reduc- tion
BOD	28	57036	1.6	25	39376	1.0	38
COD	115	57036	6.6	95	39376	3.7	44
Orthophosphate	25	57036	1.4	12	39376	0.5	64
Total phosphorus	24	57036	1.4	12	39376	0.5	64
Total solids	1028	57036	586.6	1117	39376	439.8	25
Suspended solids	54	57036	3.1	18	39376	0.7	77
Dissolved solids	974	57036	55.6	1099	39376	43.3	22

Table 17. EFFECTIVENESS OF PILOT PLANT IN TREATING SECONDARY EFFLUENT AFTER REMOVAL OF PARTITIONS. EFFLUENT DRAWN FROM DEPTH OF 0.60 m. AUGUST 6 - AUGUST 21, 1975

Parameter	Influ mg l-l	ent load liters	kg	Eff]ue	ent load liters	kg	% reduc- tion
BOD	50	44924	2.2	43	35919	1.5	33
COD	72	44924	3.2	58	35919	2.1	34
Orthophosphate	20	44924	0.9	14	35919	0.5	44
Total phosphorus	23	44924	1.0	12	35919	0.4	60
Total solids	1066	44924	47.9	845	35919	30.4	37
Dissolved solids	970	44924	43.6	825	35919	29.6	32
Suspended solids	97	44924	4.4	19	35919	0.7	84

Table 18. EFFECTIVENESS OF PILOT PLANT IN TREATING PRIMARY EFFLUENT.

AUGUST 21 - NOVEMBER 4, 1975.

_	Influent load Effluent load				% reduc-		
<u>Parameter</u>	mg 1-1	liters	kg	mg]-1	liters	kg	tion
BOD	317	188273	59.7	72	187376	13.5	77
COD	401	188273	75.5	117	187376	21.9	71
Orthophosphate	26	188273	4.9	17	187376	3.2	35
Total phosphorus	27	188273	5.1	17	187376	3.2	37
Total solids	973	188273	183.2	1156	187376	216.6	-18
Suspended solids	42	188273	7.9	32	187376	6.0	24
Dissolved solids	932	188273	175.5	1124	187376	210.6	-20

SECTION X

PHOSPHORUS DISTRIBUTION AND REMOVAL

EXPERIMENTAL BASINS

Theoretically, large quantities of phosphorus and other nutrients which are taken up by emergent aquatic plant shoots could be removed by harvesting in a manner similar to cutting hay in agricultural practice. Studies of natural stands have shown that highest concentrations of phosphorus occur in young tissue and that concentrations decrease with increasing age of tissue. This suggests that the quantity of phosphorus removed can be maximized by frequent harvesting so that plant tissues are not permitted to mature. Studies of the effect of harvesting on a natural marsh near Seymour, Wisconsin (Hanseter, 1975), have confirmed that considerable quantities of phosphorus can be removed by harvesting. Repeated harvest during the growing season can increase the removal of phosphorus by a factor of three.

On August 3, October 1, and October 23 (after frost), 1973, plant shoots were harvested (by cutting above the intercalary meristem) from quadrats in the eight experimental basins. Generally, it was found that the most rapid recovery after harvesting and, thus, the greatest potential for phosphorus removal was by the softstem bulrush (Table 19). The Iris produced very little biomass, and only negligible amounts of

phosphorus could be removed. Two to seven grams P m⁻² were removed from the six basins containing bulrushes. Examination of the plants on October 23 showed almost all of the growth of the river bulrush was from old shoots, (intercalary meristimatic growth from previously harvested shoots). In the hardstem and softstem old shoots and new shoots (arising from the rhizomes) contributed about equally.

During 1974, the plants in the four experimental basins containing gravel substrate were harvested on four occasions between June 17 and September 23. In the two basins containing softstem bulrush, 3.51 and 3.83 g P m-2 were removed (Table 20). In the river bulrush basin only 1.13 g P m-2 were removed and only 0.99 g P m-2 were removed from the Iris basin.

A removal of 3.5 g P m⁻² is equivalent to 35 Kg P ha⁻¹ (31.25 lb acre⁻¹). This amount of phosphorus is equal to that contributed by 33 persons in four months (the growing season) if a <u>per capita load of 3 lb per annum</u> is assumed (Steward, 1970). This also represents the annual removal capacity since it encompasses the entire growing season. Thus, removal capacity of only 10 persons per acre could be assumed on a year-round basis.

The frequency of harvesting had a similar effect on the total amount of plant tissue generated in a single year as Hanseter reported for a natural marsh (1975). In experimental basin number 3 (softstem bulrush), only about three-fourths as much tissue was removed during four harvests in 1974 as in two late season harvests in 1975 (Table 21). However, 5, 6 times as much phosphorus was removed by the more frequent harvesting in 1974.

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Table 19. PHOSPHORUS REMOVAL BY HARVESTING OF PLANT SHOOTS FROM EXPERIMENTAL BASINS, 1973. $(g \ P \ m^{-2})$

Basin number ^a	Plant Species	Substrate	Aug. 3	0ct. 1	Oct. 23 New shoots	Oct. 23 Old shoots	Total
2	Hardstem bulrush	Wire rack	0.28	1.72	0.31	0.33	2.64
3	Hardstem bulrush	Gravel	1.11	1.07	0.26	0.59	3.03
4	Softstem bulrush	Wire rack	2.00	4.87	0.46	0.25	7.58
5	Softstem bulrush	Gravel	1.78	1.75	0.38	0.46	4.37
6	River bulrush	Wire rack	3.74	0.38	0.18	0.43	4.73
7	River bulrush	Gravel	0.92	0.75	0.10	0.19	1.96
9	Iris	Grave1	0.002	0.0002	0.001	0.16	0.16
10	<u>Iris</u>	Wire rack	0.007	0.005	no growth	0.52	0.52

aBasin numbers 1 and 8 were control basins having gravel and wire racks, but no plants.

Table 20. PHOSPHORUS REMOVAL BY MULTIPLE HARVESTING OF EXPERIMENTAL BASINS IN 1974.

(g P m-2)

Date	Softstem bulrush	Softstem bulrush	River bulrush	Softstem bulrush
June 17	1.53	1.86	0.58	
July 19	1.32	1.06	0.33	0.37
August 8	0.30	0.61	0.16	0.39
September 23	0.36	0.30	0.06	0.23
Total	3.51	3.83	1.13	0.99

Water quality data indicated that more phosphorus was being removed by the system than was present in the harvestable plant tissue. To determine where the phosphorus was located, triplicate cores were taken from each basin at the time of harvesting in June, September, and December 1974 and in June and November of 1975. Only 0 to 14% of the total phosphorus was present in the harvestable plant tissue of any basin at any one time (Table 22). From 34 to 80% of the total phosphorus in the system was usually incorporated in the total harvestable and unharvestable plant biomass during the growing season. The remainder was retained as organic and inorganic precipitates and microflora in the gravel substrate. During the growing season of 1974, the phosphorus in the microflora-substrate complex ranged from 1-11 g m⁻² and averaged about 7 g m⁻². Between September and December of 1974, the concentration of phosphorus in the basins dropped to much lower levels, indicating a loss from the system after the autumn frosts. The only exception to

Table 21. EFFECT OF HARVESTING FREQUENCY ON STANDING CROP AND PHOSPHORUS REMOVAL FROM AN EXPERIMENTAL BASIN.

Year	Number harvests	Standing crop harvested (g dw m ⁻²)	Phosphorus removal (g P m ⁻²)
1974	4	1689.0	3.51
1975	2 (Aug; Oct)	2433.7	0.63

this pattern was Basin 9 in which the phosphorus was retained in the substrate. This basin received more sediments from erosion of the clay rich embankments than the other basins. The phosphorus retention capacity of the clay may explain the unexpected phosphorus retention of the basin after frost.

Presuming that the loss of phosphorus from the substrate after freezing is normal and predictable behavior, this loss could be a feature which would permit the removal of 15 or more g P m-2 each year. Removal could be accomplished by discontinuing the flow through the system at the time of the first heavy frost. The system could then be flushed so the concentrated washings could be collected and treated by conventional means. The maximum loading that such a system could retain is undetermined, but after receiving primary effluent for about $2\frac{1}{2}$ months, more than 20 g P m-2 was present in each of the two experimental basins. While the last data are from cores taken about the same time as in 1974, no heavy frost had yet occurred in 1975, and it is reasonable to believe that most of the phosphorus accumulated since June was still present.

Table 22. DISTRIBUTION OF PHOSPHORUS IN PLANTS AND SUBSTRATE IN EXPERIMENTAL BASINS.

			19	74				19	75	
	June 1		Sept	. 23	Deg	. 4	Jun	e 2	Nov	
Matter and the second s	g P m-2	<u>%</u>	g P m ⁻²	%	g P m-2	%	g P m ⁻²	<u>%</u>	g P m-2	2 %
Basin 3 Softstem Bulrush										
Harvested shoots	1.53	8.11	0.36	1.83	0.00	0.00	a		0.63	2.31
Unharvested shoots	1.83	9.70	2.46	12.53	0.75	12.30	1.56	17.70	0.84	3.08
Rhizomes	3.04	16.12	5.45	27.76	2.32	38.03	1.44	16.34	0.67	2.46
Roots	1.25	6.63	3.92	19.97	1.86	30.49	0.86	9.76	1.69	6.20
Total biomass	7.65	40.56	12.19	62.10	4.93	80.82	3.86	43.82	3.83	14.08
Gravel	11.21	59.94	7.44	37.90	1.17	19.18	4.95	56.18	23.39	85.92
	18.86	100.00	19.63	100.00	6.10	100.00		100.00	27.22	100.00
Basin 5 Softstem Bulrush Harvested shoots	1.86	14.09	0.30	0.75	0.00	0.00	2		ь	
Unharvested shoots	0.82	6.21	1.74	10.16	0.58	7.82	0.98	7.46	U	ļ
Rhizomes	1.42	10.76	3.12	18.21	2.30	28.86	2.18	16.59		
Roots	0.35	2.65	3.41	19.91	2.68	33.63	1.53	11.64		
Total biomass	4.45	$\frac{2.00}{33.71}$	8.57	50.03	5.56	$\frac{69.76}{69.76}$	4.69	35.69		
Gravel	8.75	66.29	8.56	49.97	2.41	30.24	8.45	64.31		
Total		100.00	17.13	100.00	7.97	100.00		100.00		
Basin 7 River Bulrush										
Harvested shoots	0.58	1.54	0.06	0.44	0.00	0.00	a		b	
Unharvested shoots	3.00	7.97	2.08	15.42	0.79	9.86	0.54	9.12		
Rhizomes	23.78	63.18	3.04	22.54	1.61	20.10	0.48	8.11		
Roots	$\frac{2.58}{29.94}$	$\frac{6.85}{79.54}$	$\frac{5.56}{10.74}$	41.22 79.61	$\frac{4.53}{6.93}$	$\frac{56.55}{86.52}$	$\frac{0.21}{1.23}$	$\frac{3.55}{20.78}$		
Total biomass	7.70	20.46	2.75	20.39	1.08	13.48	4.69	79.22		
Gravel Total		100.00	$\frac{2.75}{13.49}$	100.00		100.00		00.00		
iotai	37.07		10.73	100.00	0.01	100.00	J. 9 % 1	00.00		

aHarvested values combined with unharvested values. bExperiment terminated.

Table 22 (continued). PHOSPHORUS IN EXPERIMENTAL BASINS.

				74			19	75		
	June	_17	Sept	. 23	Deg	. 4		e 2		18
	g P m-	2 %	g P m-2	%	g P m-2	%	g P m-2	%	g P m-2	<u>%</u>
Basin 9 Softstem Bulrush										
Harvested shoots		a	0.23	1.80	0.00	0.00	ā	l	b	
Unharvested shoots	3.31	36.74	0.61	4.77	0.57	4.83	0.41	7.72		
Rhizomes	1.63	18.09	1.89	14.78	1.07	9.08	0.17	3.20		
Roots	0.91	10.10	1.84	14.39	3.19	27.06	0.17	3.20		
To tal bi omass	5.85	64.93	4.57	35.73	4.83	40.97	0.75	14.12		
Gr avel	3.16	35.07	8.22	64.27	6.96	59.03	4.56	85.88		
Total	9.01	100.00	12.79	100.00	11.79	100.00	5.31	100.00		
Basin 1 Control										
Gravel	7.91	100.00	5.18	100.00	0.38	100.00	4.35	100.00	21.6 10	0.00

dEntire plants were removed with the cores and are considered to be unharvested shoots. bExperiment terminated.

PILOT PLANT

A study similar to the one in the experimental basins was done on the distribution of phosphorus in the pilot plant. Difficulty was encountered in securing acceptable cores, however, (Section IV) so no samples were taken until after all experiments had been terminated. A record was made of tissue harvested, then cores were taken on December 5, 1975. At that time, there was an average of about 18 g P m^{-2} in the substrate (Table 23, 24). Considerable variation occurred, however, and there was a large concentration of phosphorus in the uppermost 15 cm of gravel near the influent. This high concentration was not.

Table 23. PHOSPHORUS DISTRIBUTION IN GRAVEL OF PILOT PLANT AFTER TWO SUMMERS.

(g P m-2)

Depth	Basin A	Basin B	Basin C	Mean
0-15 cm ^a	17.58	3.58	5.52	8.89
15-30 cm ^b	5.14	1.81	7.27	4.74
30-45 cm ^C	4.17	0.71	1.97	2.28
45-60 cm ^C	4.17	0.71	1.97	2.28
Total	30.72	6.81	16.74	18.09

aEach value is the mean of three.

however, associated with an accumulation of solids or dry matter at that end of the basin, as none were present (Appendix E, Tables E-1 and E-2).

bEach value is the result of analysis of a single sample.

CEach value represents a single sample taken at 45 cm and assumed to be representative of both depth intervals.

In spite of a very large standing crop, especially in Basin A, the total amount of phosphorus removed by harvesting was only about 2.09 g m-2 because the two harvests were made late in the growing season. The high yield in Basin A (2.67 g P m-2) compared to the yield in Basin C (0.69 g P m-2, Table 24) is indicative of a much more dense stand of plants at the influent end (Appendix E, Table E-3). There was an average of 420 shoots m^{-2} (1.20 m tall and 0.73 cm diameter) at the

Table 24. DISTRIBUTION OF PHOSPHORUS IN PILOT PLANT DURING 1975. $(g \ P \ m^{-2})$

Sample	Date	Basin A	Basin B	Basin C	Mean	%
Harvested shoots ^a	Aug. 1	2.12	2.76	0.48	1.80	7.14
Harvested shootsa	Sept. 24	0.45	0.20	0.21	0.29	1.15
Unharvested shoots $^{\mbox{\scriptsize b}}$	Dec. 5	0.82	0.79	0.49	0.70	2.78
Rhizomes	Dec. 5	2.91	1.61	0.64	1.72	6.82
Roots	Dec. 5	2.64	4.07	1.12	2.61	10.41
Gravel substrate	Dec. 5	30.72	6.81	16.74	18.09	71.76
Total		39.66	17.92	19.68	25.21	100.00

aEach value is the mean of three 1 m 2 quadrat samples. bEach value is the mean of three 12.6 cm diameter cores.

influent, while there were only 240 shoots m^{-2} (0.79 m tall and 0.57 cm diameter) at the effluent end. The reason for the apparent difference in vogor at the ends of the pilot plant is probably that much of the vegetation at the downstream end was killed accidentally by desiccation in 1974 and was replaced by new plants in 1975. The overall stand was quite substantial, however, with 4 - 9 g P m^{-2} in the total biomass.

This is comparable to total biomass in the experimental basins (Table 22). The total phosphorus retention capacity of the pilot plant system is probably in excess of 40 g P m^{-2} , primarily because of the great volume of the gravel substrate. None of the phosphorus in the substrate below 30 cm is likely to be available to the plants since 30 cm is as deep as the roots penetrated.

SECTION XI

BRILLION MARSH STUDIES

DESCRIPTION OF BRILLION MARSH

Physical and Biological

Brillion Marsh is located in T19N and T2ON, R2OE, Calumet County, Wisconsin. It is in the drainage basin of the North Branch of the Manitowoc River, which flows into Lake Michigan. While the total marsh area is 15 - 18 km², only a portion of the northeastern area was used in this study. The study area is located in SW½, Sec. 26; SE½, Sec. 27; NE½, Sec. 34; and NW½, Sec. 35; T2ON, R2OE (Figure 7). It has an area of 156 ha and lies at an elevation of 805 to 807 feet (245 m) above mean sea level.

Spring Creek is the major stream flowing into the study area. At its point of entrance to the marsh, it has a drainage area of 31.3 km^2 . Spring Creek has an open channel about 400 m long extending into the marsh. The channel is about 10 m wide and 1 - 1.5 m deep at the marsh entrance, and gradually diminishes in size as small distributaries lead into the marsh. The creek does not have a channel through the marsh for about 1400 m, until the marsh becomes narrow, at which point a channel about 10 - 15 m wide and 1 - 1.3 m deep is present. At the narrow part, the marsh is 200 m wide. Above this channel the marsh has a total drainage area of 49.7 km².

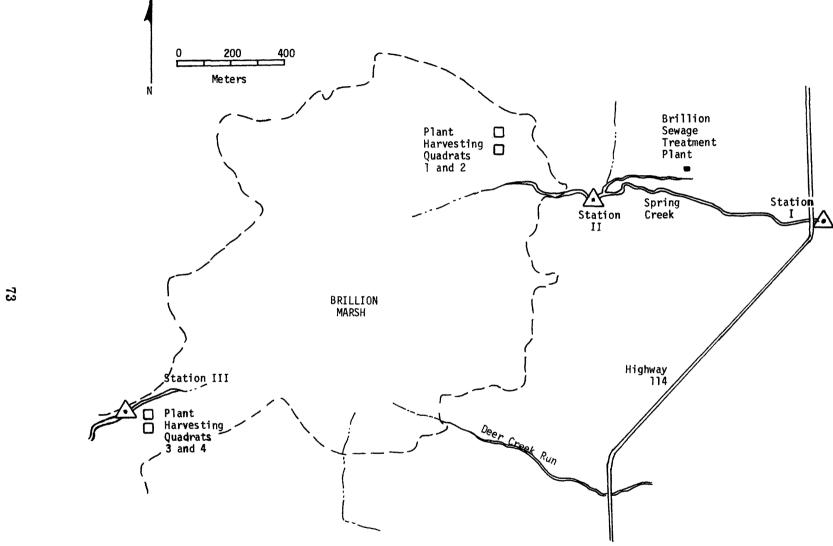


Figure 7. Location of Spring Creek and Brillion Marsh Drainage Basin.

The City of Brillion is located on Spring Creek just upstream from the marsh. Spring Creek receives effluent from the city sewage treatment plant. In addition, local industries such as a foundry and a cannery may contribute effluent at times. Agricultural runoff and urban runoff from storm sewers go into Spring Creek. There are several other streams flowing into the marsh study area. The drainage basin lies in a glaciated area with glacial tills and clays forming the predominant surficial material. The topography is flat to gently rolling.

The upper marsh has a stand of Typha latifolia-augustifolia complex. Sparganium eurycarpum is growing in a 1 - 30 m band along the stream channels at both ends of the marsh and in isolated low pockets of the marsh proper. Where Spring Creek enters the marsh (Figure 8, Station II), there are no submergent aquatic plants in the stream channel. This is presumably due to the high concentration of effluent from the Brillion sewage treatment plant which enters the stream from a channel 15 m upstream. Deer Creek Run which enters the marsh via a golf course south of Brillion is intermittent. There is a dredged channel from Highway 114 into the marsh. This channel contains water all year and has a diversified flora. Sagittaria latifolia Willd. lines the channel, along with Typha and Sparganium. Submergent vegetation includes Vallisneria americana Michx., Elodea canadensis Michx., Potomogeton pectinatus L., Ceratophyllum demersum L., and the filamentous alga Oedogonium sp. In the dredge channel at the downstream end of the marsh (Figure 8, Station III), there is a sparse stand of Ceratophyllum

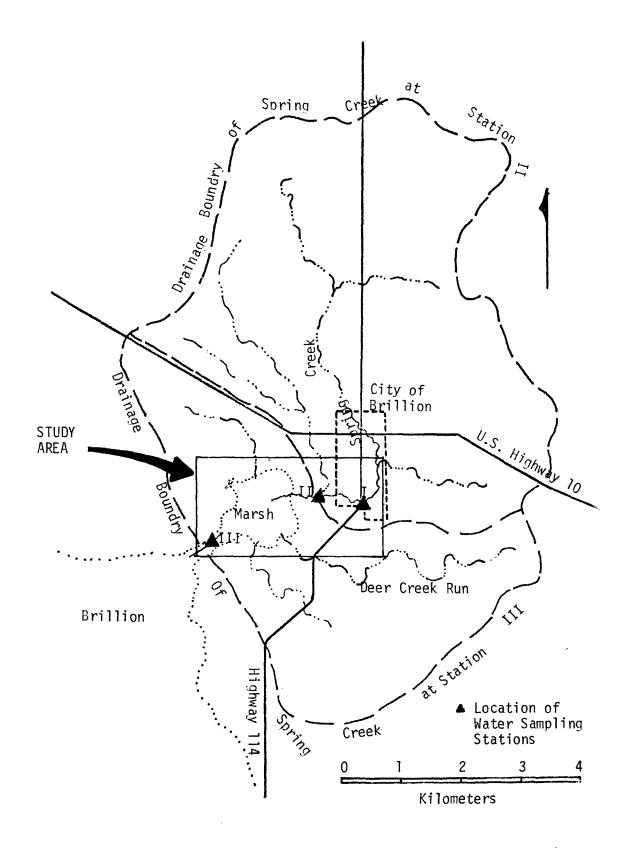


Figure 8. Map of Brillion Marsh Study Area Showing Water Sampling ♠ and Plant Harvesting ♠ Sites.

<u>demersum</u>, with isolated shoots of <u>Potomogeton pectinatus</u>. <u>Lemna covers</u> much of the surface in late summer.

Hydrology

Runoff --

The Brillion area receives a yearly average of 73 cm of precipitation. About 54 cm of this is lost through evapotranspiration, with an average runoff of 19 cm. The U.S. Geological Survey has made an estimation of the low-flow of Spring Creek at Brillion of 9.6 l sec⁻¹ for the sevenday low-flow with a two-year recurrence interval, $7Q_2$, and 4.8 l sec⁻¹ for the sevenday low-flow with a 10-year recurrence interval, $7Q_{10}$. The monthly precipitation at Brillion for June 1974 to July 1975 is given in Table 25.

With an average runoff of 19 cm year $^{-1}$ and a drainage basin area of 31.3 km 2 , the mean runoff is 0.19 m 3 sec. $^{-1}$. However, the usual runoff pattern is to have a high spring runoff and low flows the remaining months. Table 26 shows the Manitowoc River runoff for water years 1974 and 1975.

Sewage Flow --

Records obtained from the City of Brillion (population 2,588) indicate that the monthly discharge of the Brillion wastewater treatment plant ranged from 2.2×10^4 m³ to 4.6×10^4 m³ during the study period. Monthly flow of Spring Creek at Station II, has been estimated, where it merges with the wastewater discharge channel. The estimated values

were obtained by multiplying the monthly runoff from the Manitowoc River at Manitowoc by the drainage area of the sub-basin above Brillion Marsh. The monthly sewage discharge, estimated runoff of Spring Creek at Station II, and the ratio of sewage flow to runoff are given in Table 27.

Table 25. MONTHLY RAINFALL AT BRILLION, WISCONSIN. (cm)

Month	Year	Rainfall
June	1974	7.21
July	1974	4.52
August	1974	5.92
September	1974	3.30
October	1974	4.19
November	1974	4.39
December	1974	3.53
January	1975	3.12
February	1975	2.62
March	1975	8.31
April	1975	5.36
May	1975	8.08
June	1975	6.99
July	1975	6.55
August	1975	21.50
-		

During months of low runoff, the monthly sewage flow may be as much as 50 percent of the estimated monthly runoff. However, during periods of high runoff, the monthly sewage flow may be as little as 2 percent of the estimated runoff value. During low-flow months the daily sewage treatment plant flow is about 757 m 3 (200,000 gal.). The 7Q $_2$ is 812 m 3 da $^{-1}$, and the 7Q $_{10}$ is 420 m 3 da $^{-1}$. Thus, during low-flow periods, the sewage discharge may equal or exceed the natural stream flow.

Table 26. MANITOWOC RIVER RUNOFF FOR WATER YEARS 1974 AND 1975. (cm)

Month	Year	Runoff
October	1973	0.61
November	1973	0.99
December	1973	1.52
January	1974	1.14
February	1974	0.94
March	1974	7.47
April	1974	7.01
May	1974	2.39
June	1974	2.54
July	1974	0.66
August	1974	0.30
September	1974	0.15
October	1974	0.20
November	1974	0.33
December	1974	0.47
January	1975	0.30
February	1975	0.20
March	1975	4.46
April	1975	6.32
May	1975	2.19
June	1975	0.93
July	1975	0.32
August	1975	0.32
September	1975	1.26
2eh cemper	1970	1.20

WATER QUALITY

Station Locations

Three stations were selected on Spring Creek for the collection of grab samples. Samples were collected in a plastic bucket from the surface, and then transferred to plastic bottles for transport.

Station I was at the Main Street bridge in Brillion (Figure 8). The stream is 10 - 25 cm deep and 6 - 7 m wide, with a sandy bottom. The

Table 27. BRILLION SEWAGE DISCHARGE AND ESTIMATED SPRING CREEK RUNOFF. JUNE 1974 - JUNE 1975.

Month		Discharge X 104 m ³	Estimated runoff X 104 m ³	Ratio of sewage to runoff
June July August September October November December January February March April May June	1974	4.597 3.074 2.710 2.341 2.575 2.358 2.431 2.715 2.185 4.241 4.142 3.825 3.243	79.5 20.7 9.4 4.7 6.3 10.3 14.7 9.4 6.3 139.6 197.8 68.5 29.1	.06 .15 .29 .50 .41 .23 .17 .29 .35 .03 .02

location is upstream from the sewage treatment plant, but receives urban and agricultural runoff.

Station II was in Spring Creek at Wolfschmidt Road. The stream enters the marsh immediately downstream from the station. The stream is about 10 m wide and about 1 - 1.5 m deep with a sludge deposit on the bottom. A small channel 2 - 3 m wide and 1 m deep enters Spring Creek 20 m above this station. The channel carries the effluent from the Brillion sewage treatment plant, located some 300 m away.

Station III was located on the open channel about 1900 m below the upper part of Brillion Marsh. The channel is 10 - 15 m wide and 1 - 1.3 m deep. Station III has an organic bottom along the banks but, in midstream, the bottom is mineral matter.

Samples of effluent from the Brillion sewage treatment plant were collected from the effluent pipe

Long-Term Studies

Samples were collected from June 1974 through August 1975. The results of the analyses are given for each station and also for the Brillion sewage treatment plant effluent in Appendix F, Tables F-1 thru F-4. The mean values of the determinations for each parameter at Stations I, II and III are given in Table 28. Also shown on this table is the percent reduction as the water passes through the marsh from Station II to Station III.

The impact of the treatment plant effluent on the water quality of Spring Creek can be seen by comparing the water quality of Station I with Station II. BOD, COD, orthophosphate, total phosphorus, and coliform bacteria all show significant increases. Conductivity and total solids are lower, indicating dilution of dissolved solids in the stream by wastewater.

The impact of the marsh on water quality is shown by the percent reduction column. The most dramatic decrease was in BOD (80.1%) and total coliforms (86.2%). Significant decreases also occurred in turbidity (43.5%), suspended solids (29.1%) and nitrate (51.3%). Reductions in total phosphorus (13.4%), orthophosphate (6.4%), and conductivity (7.7%) also occurred. There was a 34% increase in dissolved solids which caused an 8.5% increase in total solids.

Table 28. MEAN VALUES FROM LONG TERM STUDIES AT STATIONS I, II, III.

Parameter	Station I	Station II	Station III	% Reduction
BOD	5.9	26.9	5.35	80.1
COD	30.5	106	59.7	43.7
Orthophosphate	1.27	3.13	2.93	6.4
Total phosphorus	1.28	3.43	2.97	13.4
Conductivity (umho)	1540	1065	983	7.7
Turbidity (JTU)	26	20	11.3	43.5
Nitrate	1.78	1.17	0.57	51.3
Coliform (log col 100 ml-1)	4.76	5.54	4.68	86.2
рН	7.91	7.81	7.42	
Total solids	955	707	767	-8.5
Suspended solids	154	127	90	29.1
Dissolved solids	801	580	677	-16.7

aAll values are mg 1-1 unless otherwise indicated in parentheses.

<u>Intensive Study</u>

A three-day intensive study of water quality in Brillion Marsh was made on August 19 - 22, 1974. The purpose of the investigation was to determine the nature and extent of daily variations in parameter concentrations. Starting at noon on August 19th, the samples were collected every four hours until noon, August 22. Samples were collected at Stations I, II, and III. The results of the samples are graphed in Appendix F, Figures F-1 to F-12.

The flow of the Brillion sewage treatment plant and the flow of Spring Creek at Station I during this period are given in Table 29.

Table 29. BRILLION SEWAGE DISCHARGE AND SPRING CREEK STREAMFLOW DURING AUGUST 19 - AUGUST 22, 1974.

Date	Brillion sewage treatment plant discharge	Stream flow in Spring Creek at Station I
August 19, 1974	1070 m ³ da-1	3350 m ³ da-1
August 20, 1974	1000 m ³ da-1	2860 m ³ da-1
August 21, 1974	1000 m ³ da-1	2450 m ³ da-1
August 22, 1974	945 m ³ da-1	2350 m ³ da-1

PRIMARY PRODUCTION AND PHOSPHORUS

Primary Production

Shoots were harvested from the experimental quadrats (Table 30) in Brillion Marsh on July 1, and October 8, 1974, and again on June 19, August 5, and October 7, 1975. Control quadrats were harvested only in October each year. There was generally less growth in both the control and experimental quadrats in 1975 than in 1974 (Table 30). Less growth in 1975 may be a result of climatological variation. There was also generally less total tissue removed from the experimental Typha and Sparganium plots than from the controls. In the artificial marsh basins at Seymour, the total harvest of Scirpus tissue was not diminished by frequent harvesting.

Decreased productivity with harvesting in a natural marsh was also recorded by Hanseter (1975) and is probably typical of natural marsh systems. It can also be noted (Table 30) that the experimental quadrats (multiple harvests) from the effluent end of the marsh produced only about 40% as much standing crop as their control plots. Experimental quadrats on the receiving or upstream end of the marsh produced about 80% as much tissue as their control. There was only a slight difference between control plots. This seems to indicate some sort of limiting factor or lack of reserves in the plants at the downstream end of the marsh.

Table 30. PLANT TISSUE HARVESTED FROM BRILLION MARSH. (g dw m-2)

Receiving end of marsh								
Quadrat	Treatment	July 1	1974 Oct. 8	Total	June 19	197 Aug. 5	5 Oct. 7	Total
								10001
1	Experi-	561	314	875	263	429	11	703
2	mental	950	483	1433	243	423	31	697
1c	Control		1072	1072			924	924
2c	CONCIOI		1382	1382			1071	1071
			1002	1002				1071
Effluent	Effluent end of marsh							
			1974			197	-	
Quadrat	Treatment	July 1	Oct. 8	Total	June 19	Aug. 5	Oct. 7	<u>Total</u>
2	Evnovi	306	373	679	218	324	6	458
3 4	Experi-			558	137	174	2	313
4	mental	291	267	220	137	1/4	۷	313
3c	Control		1168	1168			763	763
4c			1665	1665			1481	1481

Phosphorus

Content in Plant Tissue --

On June 19, 1975, undisturbed control plots were harvested. The old tissue remaining standing from 1974 was separated from the new shoots and the tissue was analyzed for phosphorus content. In the <u>Typha</u>, so little phosphorus remained in the old stalks that it could not be detected by the method employed (Table 31). The old <u>Sparganium</u> shoots in quadrat 3 contained a small amount of phosphorus. The new 1975 shoots, however, contained 0.05 - 0.09% phosphorus, so that this single, early season harvest removed 0.20 - 0.30 g P m-2. Thus, old plant tissue contains no phosphorus and in spite of a large standing crop, a winter or early spring harvest of tissue would remove little or no phosphorus.

Removal by Harvesting --

By multiple harvesting, the average phosphorus content of the plant tissue was maintained at about 0.125%, while that of the control plants harvested once at the end of the season was only about 0.058% phosphorus (Table 31). There was no apparent difference between the phosphorus content of plants from the upstream and the downstream ends of the marsh.

The amount of phosphorus removed from the control quadrats averaged about $0.6~g~m^{-2}$ (Table 33), and because there was little difference in standing crops, there was little or no difference between

Table 31. CONTRIBUTION OF OLD AND NEW SHOOTS TO HARVESTABLE BIOMASS AND PHOSPHORUS REMOVAL FROM BRILLION MARSH. (g m-2)

Quadrat	Species	Tissue	Dry Weight	% Phosphorus	Phosphorus
1	Typha	01d New	839 287	0.00 0.09	0.00 0.26
2	<u>Typha</u>	01d	801	0.00	0.00
2		New	288	0.07	0.20
3	<u>Sparganium</u>	01d	189	0.04	0.06
3		New	243	0.09	0.22
4	Typha	01d	212	0.00	0.00
4		New	558	0.05	0.30

Table 32. CONCENTRATION OF PHOSPHORUS IN HARVESTED SHOOTS FROM BRILLION MARSH.

(% P)

Receiving end of marsh 1974 1975							
Quadrat	Treatment	July 1	/4 Oct. 8	June 19	1975 Aug. 5	Oct. 7	
1 2	Experimental	0.254 0.094	0.064 0.084	0.095 0.091	0.158 0.137	0.100 0.093	
1c 2c	Control		0.054 0.048			0.052 0.067	
Effluent Quadrat	end of marsh Treatment	July 1	0ct. 8	June 19	Aug. 5	Oct. 7	
3 4	Experimental	0.304 0.122	0.110 0.080	0.119 0.078	0.169 0.114	0.093 0.114	
3c 4c	Control		0.058 0.050			0.074 0.067	

phosphorus removed from the receiving and effluent ends of the marsh. In the multi-harvested experimental plots, at the receiving end of the marsh, there was about twice as much phosphorus removed $(0.83 - 162 \text{ g P m}^{-2})$ as from the control plots (Table 33). At the effluent end of the marsh, however, there was about 40% more phosphorus removed from the experimental plots in 1974 but 30% less in 1975. This, again, reflects the apparent lack of reserve of some unknown limiting factor at the effluent end of the marsh.

In spite of previous findings, it was thought that it might be possible to remove a significant portion of the phosphorus from the marsh system by harvesting the plants. If an average of 1 g P m⁻² could be removed from the system by two or more harvests during the growing season, only 1.56×10^3 kg of phosphorus would be removed from Brillion Marsh. That is only 6.5% of the estimated influx. This minor reduction would be at a very high cost to wildlife using the system. After the first early summer harvest, only 10-30% as much standing crop was present at any time in the harvested quadrats as in the unharvested control plots. Thus, valuable wildlife cover would be removed and machinery used for harvesting would disturb nesting birds. In addition, it was noted that in one experimental quadrat (number 4) the Typha had been largely replaced by Spraganium in 1975. Similar changes in species composition were recorded by Hanseter (1975).

Mass Balance --

If a marsh is to be regarded as an alternative to traditional wastewater treatment processes, it must be capable not only of reducing BOD and killing pathogens, but it must also provide a mechanism for removing mineral nutrients ($\underline{i}.\underline{e}.$, phosphorus). Water quality data indicated that there was usually a significant reduction in phosphorus concentration between Stations II and III. Exceptions were recorded, however, and it was known that the marsh was receiving additional phosphorus from

Table 33. REMOVAL OF PHOSPHORUS FROM BRILLION MARSH BY HARVESTING OF PLANT SHOOTS.

(g P m-2)

Receiving end of marsh								
Qua- drat	Treatment	July 1	1974 Oct. 8	Total	June 19	1975 Aug. 5	0ct., 7	Total
1 2	Experimental	1.42 0.89	0.20 0.41	1.62 1.30	0.25 0.22	0.67 0.58	0.01 0.03	0.93 0.83
1c 2c	Control		0.58 0.66	0.58 0.66			0.48 0.56	0.48 0.56
Efflu Qua-	ent end of mar	sh	1974			1975		
•								
<u>drat</u>	Treatment	July 1		Total.	June 19		Oct. 7	Total
<u>drat</u> 3 4	Treatment Experimental	July 1 0.93 0.36		Total 1.34 0.57	June 19 0.15 0.11		0.01 0.002	Total 0.77 0.31

non-point agricultural and recreational sources via surface run-off and other tributaries. Thus, an attempt was made to estimate the mass balance of phosphorus in and out of the Brillion Marsh. Values obtained in this study have been expressed as concentrations. Physical constraints

of the channels prevented stream flow measurements anywhere but Station

I. It was not possible to measure to the mass flux of water through
the marsh system.

An estimate of the mass flux can be made using the average monthly discharge of the Manitowoc River at Manitowoc, Wisconsin. For the period June 1974, through May 1975, this was 18.1 cm for the drainage basin; about an average year. The average phosphorus concentration during this period at each station was: Station I: $0.88 \text{ mg } 1^{-1}$; Station II: $3.65 \text{ mg } 1^{-1}$; Station III: $1.78 \text{ mg } 1^{-1}$.

Station II is assumed to be representative of the $31.2~\rm km^2$ drainage basin of Spring Creek entering the marsh and Station I representative of the remaining $18.7~\rm m^2$ drainage basin of the marsh study area (Figure 7). With an annual runoff of $5.67~\rm x~10^6~\rm m^3$ into the marsh from Spring Creek, a total of $20.7~\rm x~10^3~\rm kg$ of phosphorus entered the marsh through Spring Creek. An additional $3.40~\rm x~10^6~\rm m^3$ of runoff entered from the rest of the drainage basin, carrying with it $2.99~\rm x~10^3~\rm kg$ of phosphorus.

Drainage from the marsh will be presumed to be equal to the runoff into it. Precipitation over the marsh is probably cancelled by evapotrans-piration from the marsh. Total runoff would be 9.07 x 10^6 m³ with 16.1 x 10^3 kg P.

Total input of phosphorus to the marsh was 23.7×10^3 kg and output was 16.1×10^3 kg. A reduction of 32 percent is noted. However, the method

of approximation is fairly crude in terms of estimating the volumes of water. The data should, perhaps, be interpreted as meaning only that the output is on the same order of magnitude as the input. This was also the conclusion reached by Lee, Bently and Amundson (1969) in their study of natural Wisconsin marshes. The marsh does tend to store phosphorus during the summer. Phosphorus is slowly released from the plants and sediments during the fall and winter when flows are low but concentrations are high. A large slug of phosphorus passes from the marsh during spring runoff.

Precipitation Into Sediments --

On August 5, 1975, a series of sediments cores was taken from Brillion Marsh to determine if there was any precipitation of phosphorus into the sediments. The phosphorus concentration in the organic marsh sediments was very uniform and ranged from 1.1 - 2.4 mg P (g dw sediments)⁻¹ (Figure 9). There was no difference between the upstream and downstream ends of the marsh. Cores taken from Spring Creek channel in the City of Brillion and especially from the effluent channel of the sewage treatment plant contained up to 20.6 mg P (g dw sediments)⁻¹. Over 9 mg per gram were also found in the bottom of Deer Creek Run, a small tributary receiving runoff from dairy farms and a golf course. It must be concluded, then, that some, perhaps a major, portion of the phosphorus which had been assumed to be entering the marsh was, in fact, being removed by some nonbiological precipitation process in the stream channel. Some of the phosphorus may have been

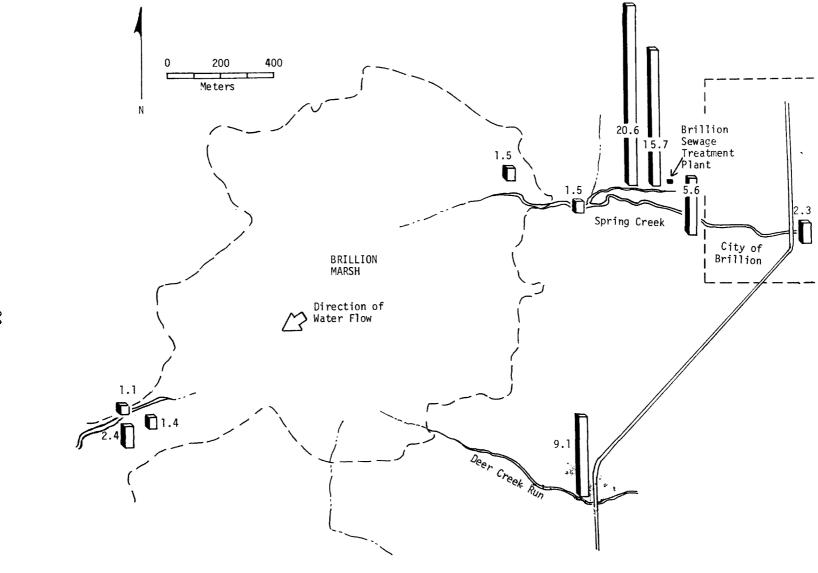


Figure 9. Available Phosphorus in Surface Sediments in Brillion Marsh. (mg P $\rm g^{-1}$ dry weight sediment)

removed by adsorption onto clay and silt from non-point runoff while another portion may have precipitated with carbonates. This is not to suggest that the aquatic plants played no role in the precipitation process. Wetzel (1969) demonstrated that aquatic macrophytes play a very significant role in precipitation of phosphorus in hard water situations such as in Spring Creek at Brillion. A large stand of submerged plants was present in Deer Creek Run and almost the entire length of the receiving stream and 200 m of Spring Creek above Station II were lined with Sparganium and Typha.

SECTION XII

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SECTION XIII

GLOSSARY

<u>Artificial</u> <u>Marsh</u> - A constructed area containing herbaceous emergent vegetation through which wastewater is permitted to flow. In this case, plastic-lined basins were partially filled with gravel which was used as a substrate.

<u>Effluent</u> - The water flowing out of the experimental basins and the pilot plant. Not to be confused with primary effluent and secondary effluent which are produced by the municipal wastewater treatment plant.

<u>Emergent Vegetation</u> - Grasses, sedges and other herbaceous plants which typically grow in wetlands or natural marshes (see below) and which have a substantial portion of erect tissue extending above the water surface.

<u>Evapotranspiration</u> - Water loss resulting from transpiration via plant leaf surfaces and direct evaporation from other exposed surfaces.

Experimental Basin - The square, plastic-lined basins used in this project.

<u>Influent</u> - The water entering the experimental basins and the pilot plant. In some cases it is wastewater which has received secondary (activated sludge) treatment. In other cases it was wastewater which has received primary treatment only.

<u>Intercalary Meristem</u> - A meristem, removed from the apical meristem, which produces primary tissue; <u>e.g.</u>, the zone of cell division at the base of emergent leaves of some marsh plants.

Natural Marsh - An area having free standing water at least part of the year and having a substantial cover of grasses and other herbaceous plants but little or no substantial cover of perennial woody plants. Herein used to refer to the entire ecosystem, including plants, surface and near-surface water and sediments (primarily organic) to the depth of the plant roots.

<u>Pilot Plant</u> - The long, plastic-lined trench used in this project.

<u>Primary Effluent</u> - Wastewater which received primary treatment (only) in the municipal treatment plant.

<u>Propagule</u> - A vegetative propagation unit which may consist of any viable part.

<u>Secondary Effluent</u> - Wastewater which received secondary treatment in the municipal treatment plant.

<u>Shoot</u> - The emergent portion of the plant. In all cases used, this refers to sheathes of leaves only and does not include submergent rhizomes or other woody tissues.

Substrate - The medium into which the roots of the vegetation penetrate.

SECTION XIV

APPENDICES

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APPENDIX A

DYE STUDIES

Dye Study I

One gram Rhodamine-B dissolved in 1.2 1 water and introduced with influent water on June 14, 1975. At that time, there was approximately 2.5 cm (1 in.) of free-standing water on top of the gravel substrate due to recent rainfalls.

As might be expected, the water moved across the surface along the path of least resistance (Figure A-1). Within ten hours, dye was detected in the effluent. Some dye did penetrate into the gravel as low readings were recorded at 30 and 45 cm in the center basin after only 23 hours. The main flow of water, however, was horizontally over the surface to the area immediately above the outlet, then vertically down to the outlet at 60 cm depth.

Dye Study II

One gram of Rhodamine-B was introduced with influent on July 21, 1975. All flow at that time was below the surface (no surface water). Water collected from small depressions in the gravel indicated that there was little or no flow directly at the surface (Figure A-2). Subsurface flow, however, was nearly uniform indicating that the design was proper. Unfortunately, no dye was ever detected in the effluent. This

apparently indicates that the dye was absorbed onto organic materials in the inter-gravel spaces.

Dye Study III

This study was conducted in order to determine the effect of removal of the plywood partitions on the flow patterns in the pilot plant.

Two grams of Rhodamine-B in one 1 of water were introduced on August 13 and sampling was continued until August 25, 1975. Results were virtually identical to those in Study II in that dye reached downstream end after four days, but never was found in the effluent. The flow was apparently uniform throughout the portion of the basin in which dye was detected (Figure A-3).

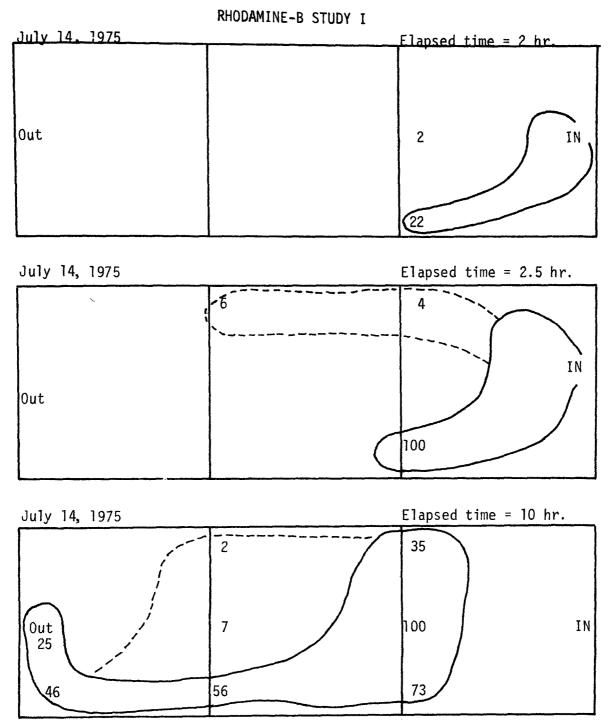


Figure A-1. Flow of Water in the Bulrush Pilot Plant When Water Level was 2.5 cm Above Gravel Substrate. Numbers are Fluorometer Readings. Vertical Series of Numbers Represent Readings at 0, 15 and 60 cm, Respectively.

RHODAMINE-B STUDY I

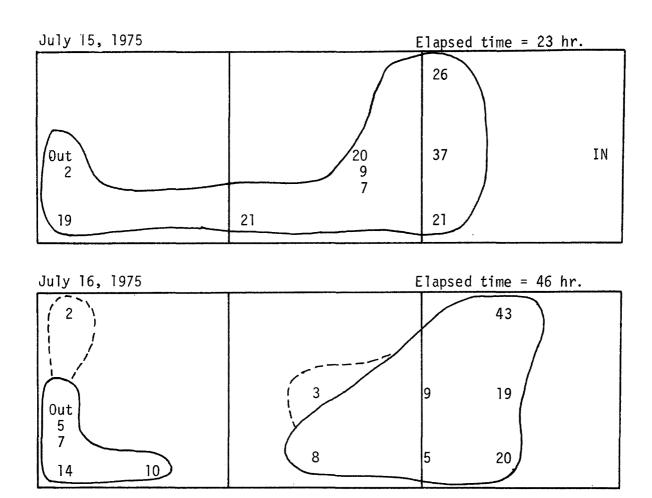


Figure A-1 (continued). Rhodamine-B Study I.

RHODAMINE-B STUDY II

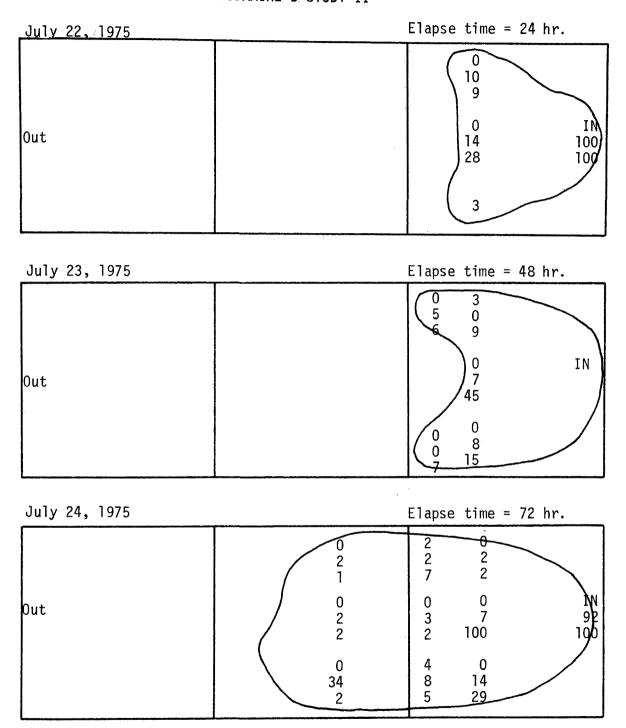


Figure A-2. Flow of Water in Bulrush Pilot Plant When Water Level Was at Gravel Substrate Surface. Numbers are Fluorometer Readings. Vertical Series of Numbers Represent Readings at 0, 15 and 60 cm, Respectively.

RHODAMINE-B STUDY II

July 25, 1975		Elapse time = 100 hr.						
	0 5 24	0 15 6	0 10 14					
Out) 0 11 21	0 5 4	4 46 49	IN 56 100				
	0 15 4	0 7 19	0 10 27					

July 28, 19	975		E1.	apse time	= 172 hr.
0 22 17	0 19 15	0 24 16 1	0 7 2	0 0 9 13 9 35	
	0 10 28	5 2 1 3 2	0 1 1 14 1 :	1 0 4 3 3 43	TN 4B
0 2 27	0 10 20		0 4 2	0 0 5 10 0 22	

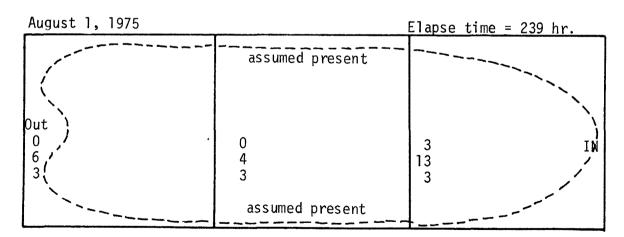
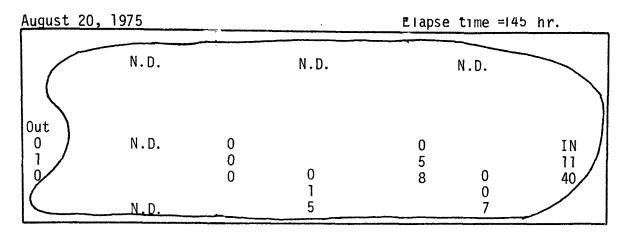


Figure A-2 (continued). Rhodamine-B Study II.

RHODAMINE-B STUDY III



August 21, 1975				Elapse time =169 hr.				
	. —————— N.D.		N.D.		T.D			
Out \ 0 1 0 0	N.D.	0 2 1	0 0 2	0 1	N.D.	IN 6 /		
1	N.D.		3 0 	,	_ <u>N</u> . D			

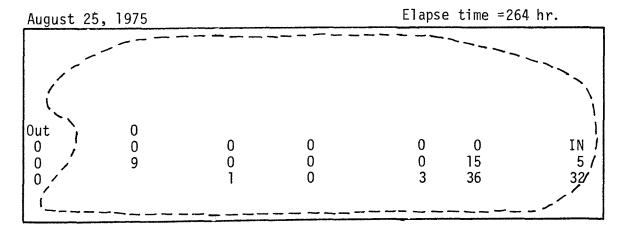


Figure A-3. Flow of Water Through Bulrush Pilot Plant When Outlet was at 60 cm and the Wooden Partitions Had Been Removed. Numbers are Fluorometer Readings a Vertical Series of Numbers Represent Readings at 0, 15, and 60 cm, Respectively.

RHODAMINE-B STUDY III

August 14, 1975	Elapse time = 24 hr.
Out O	0 0 IN 2 31 5 54

August 15, 1975			Ela	pse time	= 48 hr.
		N.D.		N.D.	
Out	0	0 1 1	0· 4 6	0 34 57	IN 100 100
	5	0 2 2		0 2 4	

August 18	3, 1975				Elapse tim	e = 96 hr.
	N.D.		N.D.		N.D.	
Out	0 4 3	0 2 0	0 2 2	0 2 13	0 17 60	IN 22 100
3 9	0 1 ND		0 2 3		0 3 10	

Figure A-3 (continued). Rhodamine-B Study III.

APPENDIX B

GREENHOUSE DATA

Table B-1. WATER BALANCE DATA FOR GREENHOUSE STUDIES. (liters)

	Influent	Total effluent	% evapotranspiration
Five day retention	0.4	20.7	C 2
Iris	84	30.7	63
Control	84	31.7	62
Hardstem	84	31.7	62
Softstem	84	33.7	60
Softstem	82	34.2	58
Three day retention	1		
Iris	140	86	38
Control	140	87	37
Hardstem	140	84	40
Softstem	142	91	36
Softstem	142	95	33
One & one-half day	retention		
Iris	377	282	26
Control	378	274	28
Hardstem	373	329	12
	373 373	343	8
Softstem	373 378	368	9
Softstem	3/0	300	3

Table B-2. EFFECTIVENESS OF GREENHOUSE BEDS IN TREATING PRIMARY EFFLUENT. ONE AND ONE-HALF DAY RETENTION.

	Influent load Eff			Eff]ı	ent loa	%	
	mg 1-1	liters	g	mg 1-1	liters		reduction
BOD							
Iris	104	377	3 9	15	282	4	8 9
Control	104	378	39	19	274	5 4	86
Hardstem	104	373	39	11	329	4	90
Softstem	104	373	39	13	343	4	88
Softstem	104	378	39	20	368	7	81
COD							
Iris	125	377	47	84	282	24	49
Control	125	378	47	78	274	22	54
Hardstem	125	373	47	55	329	18	61
Softstem	125	373	47	57	343	20	57
Softstem	125	378	47	90	368	33	29
Orthophosphate							
Iris	1.21	377	0.46	1.68	282	0.47	-3
Control	1.21	378	0.46	1.69	274	0.46	-1
Hardstem	1.21	373	0.45	0.98	329	0.32	28
Softstem	1.21	373	0.45	0.56	343	0.19	57
Softstem	1.21	378	0.46	0.86	368	0.32	30
Total phosphorus							
Iris	2.25	377	0.85	2.09	282	0.59	30
Control	2.25	378	0.85	2.10	274	0.57	32
Hardstem	2.25	373	0.84	1.05	329	0.34	59
Softstem	2.25	373	0.84	0.96	343	0.33	60
Softstem	2.25	378	0.85	1.12	368	0.41	51
Coliform (log col.							
Iris	3.28	377	5.86	3.38	282	5.82	7
Control	3.28	378	5.86	3.31	274	5.74	22
Hardstem	3.28	373	5.85	3.26	329	5.77	15
Softstem	3.28	373	5.85	3.30	343	5.84	4
Softstem	3.28	378	5.86	3.27	368	5.83	5

Table B-2 (continued). EFFECTIVES OF GREENHOUSE BEDS. ONE AND ONE-HALF DAY RETENTION.

+ (i : i	luent lo	ad	Effluent load			%
g]-1	liters	g	g1-1	liters	g	reduction
0.75	377	283	0.83	282	234	17
						24
0.75	373	280	0.82	322	264	5
0.75	373	280	0.86	343	295	-5
0.75	378	284	0.88	368	324	-14
0.11	377	41	0.09	282	25	3 9
0.11	378	41	0.07	274	19	54
0.11	373	41	0.10	322	32	22
0.11	373	41	0.10	343	34	16
0.11	378	41	0.13	368	48	-15
0.64	377	241	0.74	282	209	13
0.64	378	242	0.71	274	195	19
0.64	373	239	0.72	322	232	2
0.64	373	239	0.76	343		-9
0.64	378	242	0.75	368	276	-14
	0.75 0.75 0.75 0.75 0.75 0.11 0.11 0.11 0.64 0.64 0.64	0.75 377 0.75 378 0.75 373 0.75 373 0.75 378 0.11 377 0.11 378 0.11 373 0.11 373 0.11 378 0.11 378 0.64 377 0.64 378 0.64 373 0.64 373	0.75 377 283 0.75 378 284 0.75 373 280 0.75 373 280 0.75 378 284 0.11 377 41 0.11 378 41 0.11 373 41 0.11 373 41 0.11 378 41 0.11 378 41 0.11 378 41 0.11 378 241 0.64 377 241 0.64 378 242 0.64 373 239 0.64 373 239	0.75 377 283 0.83 0.75 378 284 0.78 0.75 373 280 0.82 0.75 373 280 0.86 0.75 378 284 0.88 0.11 377 41 0.09 0.11 378 41 0.07 0.11 373 41 0.10 0.11 373 41 0.10 0.11 378 41 0.13 0.64 377 241 0.74 0.64 378 242 0.71 0.64 373 239 0.72 0.64 373 239 0.76	0.75 377 283 0.83 282 0.75 378 284 0.78 242 0.75 373 280 0.82 322 0.75 373 280 0.86 343 0.75 378 284 0.88 368 0.11 377 41 0.09 282 0.11 378 41 0.10 322 0.11 373 41 0.10 343 0.11 378 41 0.13 368 0.64 377 241 0.74 282 0.64 378 242 0.71 274 0.64 373 239 0.72 322 0.64 373 239 0.76 343	0.75 377 283 0.83 282 234 0.75 378 284 0.78 242 214 0.75 373 280 0.82 322 264 0.75 373 280 0.86 343 295 0.75 378 284 0.88 368 324 0.11 377 41 0.09 282 25 0.11 378 41 0.10 322 32 0.11 373 41 0.10 343 34 0.11 378 41 0.10 343 34 0.11 378 41 0.13 368 48 0.64 377 241 0.74 282 209 0.64 378 242 0.71 274 195 0.64 373 239 0.72 322 232 0.64 373 239 0.76 343 261

Table B-3. EFFECTIVENESS OF GREENHOUSE BEDS IN TREATING PRIMARY EFFLUENT. THREE-DAY RETENTION.

	Inf]uent load			Eff]uent load			%
- 7	mg 1-1	lite	rs g	mg 1-1	liters	g	reduction
BOD							
Iris	179	140	25	17.9	86	1.5	94
Control	179	140	25	40.9	87	3.6	86
Hardstem	179	140	25	23.2	84	1.9	92
Softstem	179	142	25	12.1	91	. 1.1	96
Softstem	179	142	25	13.6	95	1.3	95
COD							
Iris	409	140	57	179	86	15.4	73
Control	409	140	57	163	87	14.2	75
Hardstem	409	140	57	73	84	6.1	89
Softstem	409	142	58	96	91	8.8	85
Softstem	409	142	58	112	95	10.7	82
Orthophosphate							
Iris	4.08		0.57	6.1	86	0.53	8
Control	4.08		0.57	6.7	87	0.58	2
Hardstem	4.08		0.57	4.1	84	0.34	41
Softstem	4.08		0.58	2.4	91	0.21	63
Softstem	4.08	142	0.58	3.1	95	0.30	49
Total phosphorus							
Iris	5.51	140	0.77	5.9	86	0.51	34
Control	5.51	140	0.77	9.2	87	0.80	-4
Hardstem	5.51	140	0.77	5.1	84	0.42	45
Softstem	5.51	142	0.78	3.3	91	0.30	62
Softstem	5.51	142	0.78	3.8	95	0.36	54
Coliform (log col.	100 m1-1)						
Iris	3.29	140	5.43	3.00	86	4.93	68
Control	3.29	140	5.43	2.79	87	4.72	80
Hardstem	3.29	140	5.43	2.73	84	4.65	83
Softstem	3.29	142	5.44	2.56	91	4.52	88
Softstem	3.29	142	5.44	2.51	95	4.49	89
						···	

Table B-3 (continued). EFFECTIVENESS OF GREENHOUSE BEDS. THREE-DAY RETENTION.

	Influent load Ef			Eff]	uent loa	%	
	mg 1-1	liters	g	mg 1-1	liters	g	reduction
Total solids							
Iris	1046	140	146	1158	86	100	32
Control	1046	140	146	1085	87	95	35
Hardstem	1046	140	146	1128	84	94	35
Softstem	1046	142	148	1230	91	112	25
Softstem	1046	142	148	1168	95	111	25
Dissolved solids							
Iris	783	140	110	994	86	86	22
Control	783	140	110	944	87	82	22
Hardstem	783	140	110	981	84	82	25
Softstem	783	142	111	1032	91	94	15
Softstem	783	142	111	1003	95	95	14
Suspended solids							
Iris	258	140	36	165	86	14	61
Control	258	140	36	140	87	12	66
Hardstem	258	140	36	133	84	11	69
Softstem	258	142	37	167	91	15	59
Softstem	258	142	37	165	95	16	57

Table B-4. EFFECTIVENESS OF GREENHOUSE BEDS IN TREATING PRIMARY EFFLUENT. FIVE-DAY RETENTION.

	Influent load			Eff]ı	%		
	mg 1-1	liters	g	mg 1-1	liters	g	reduction
COD							
Iris	284	84	23.9	90	30.7	2.7	89
Control	284	84	23.9	90 84	31.7	2.6	89
Hardstem	284	84	23.9	98	31.7	3.1	87
Softstem	284	84	23.9	98 88	33.7	2.9	87 87
Softstem	284	82	23.3	135	34.2	4.6	80
00 / 00 0Cm	20.	0 L	20.0	100	0112	,	00
BOD							
Iris	250	84	21.0	15	31.7	0.5	98
Control	250	84	21.0	22	31.7	0.7	97
Hardstem	250	84	21.0	19	31.7	0.6	97
Softstem	250	84	21.0	16	33.7	0.5	97
Softstem	250	82	20.5	19	34.2	0.6	97
Orthophosphate							
Iris	6.30	84	0.5	7.35	30.7	0.2	57
Control	6.30	84	0.5	8.02	31.7	0.2	52
Hardstem	6.30	84	0.5	5.42	31.7	0.2	67
Softstem	6.30	84	0.5	3.74	33.7	0.1	76
Softstem	6.30	82	0.5	4.51	34.2	0.2	70 70
<u>Total phosphorus</u>							
Iris	13.19	84	1.1	7.37	30.7	0.2	80
Control	13.19	84	1.]	8.37	31.7	0.3	76
Hardstem	13.19	84	1.1	5.43	31.7	0.2	83
Softstem	13.19	84	1.1	5.51	33.7	0.2	83
Softstem	13.19	82	1.1	6.54	34.2	0.2	79
							

APPENDIX C

EXPERIMENTAL BASIN DATA

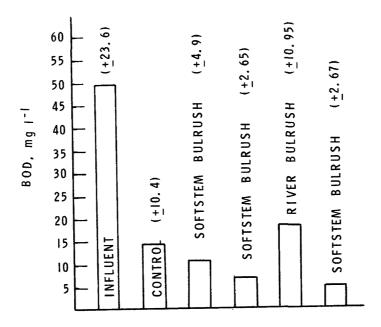


Figure C-1. BOD of Samples Collected Between June 19 and July 31, 1974.

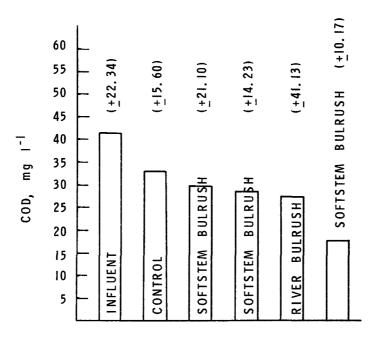


Figure C-2. COD of Samples Collected Between June 19 and July 31, 1974.

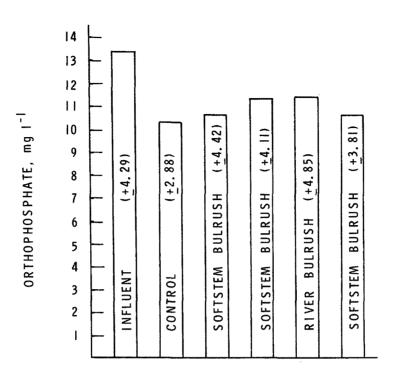


Figure C-3. Orthophosphate Concentrations in Samples Collected Between June 19 and August 31, 1974.

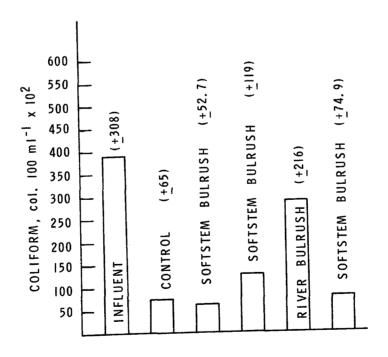


Figure C-4. Coliform Bacteria in Samples Collected Between June 19 and August 31, 1974.

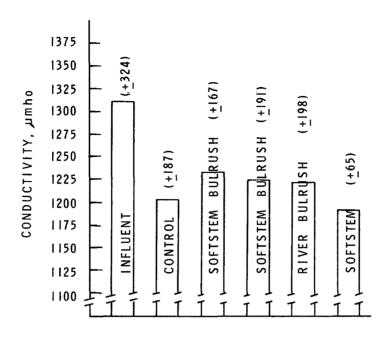


Figure C-5. Conductivity of Samples Collected Between June 19 and August 31, 1974.

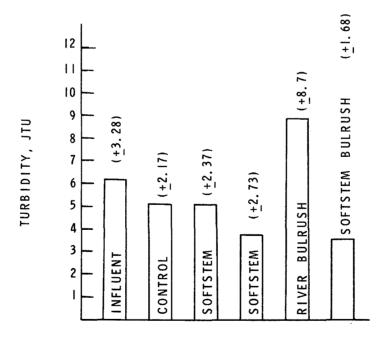


Figure C-6. Turbidity of Samples Collected Between June 19 and August 31, 1974.

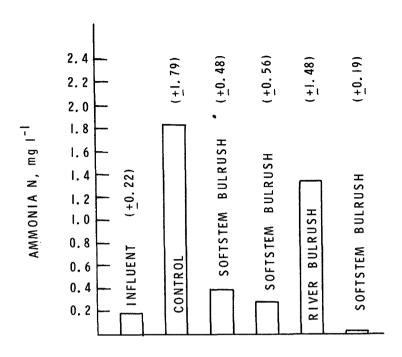


Figure C-7. Ammonia Concentrations in Samples Collected Between June 19 and July 31, 1974.

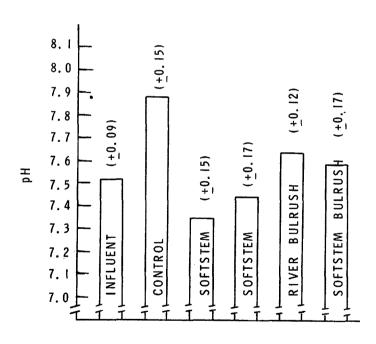


Figure C-8. pH of Samples Collected Between June 19 and August 31, 1974.

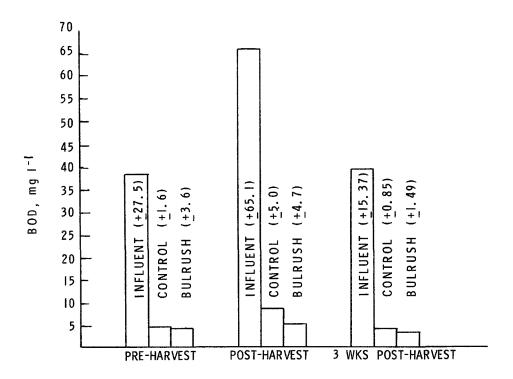
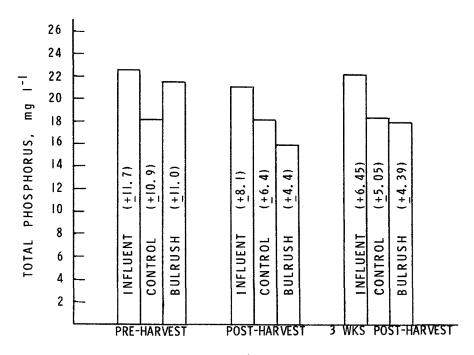


Figure C-9. BOD Reduction by Control and Experimental Bulrush Basin During Three Intensive Study Periods (five-hour retention).



Five C-10. Total Phosphorus Reduction by Control and Experimental Bulrush Basin During Three Intensive Study Periods (five-hour retention).

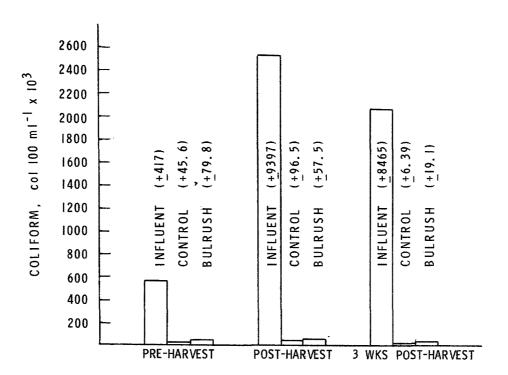


Figure C-11. Coliform Reduction by Control and Experimental Bulrush Basin During Three Intensive Study Periods (five-hour retention).

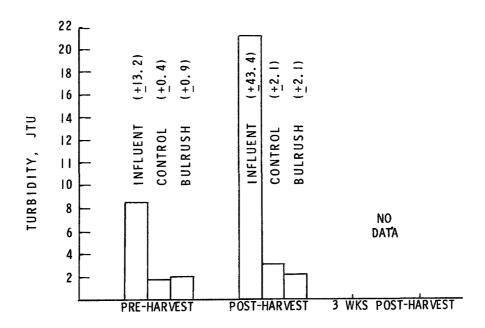


Figure C-12. Turbidity Reduction by Control and Experimental Bulrush Basin During Three Intensive Study Periods (five-hour retention).

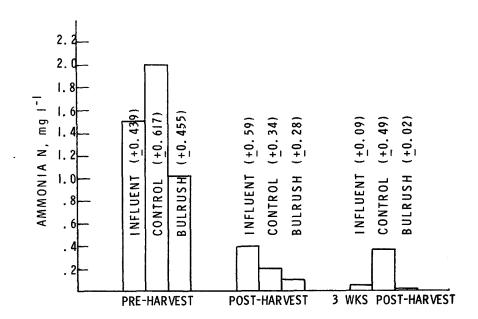


Figure C-13. Ammonia Removal by Control and Experimental Bulrush Basin During Three Intensive Study Periods (five-hour retention).

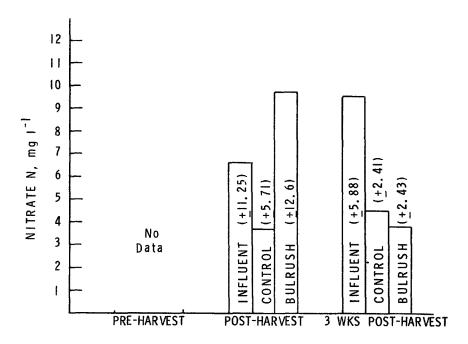


Figure C-14. Nitrate Reduction by Control and Experimental Bulrush Basin During Three Intensive Study Periods (five-hour retention).

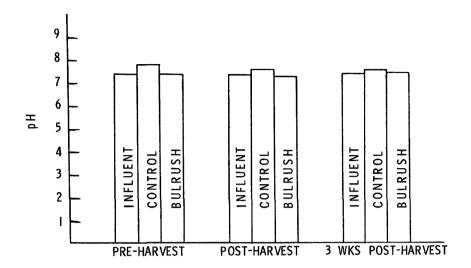


Figure C-15. pH Values of Secondary Effluent, Control Pond Effluent and Experimental Basin Effluent During Three Intensive Study Periods.

Table C-1. AVERAGE VALUES OF VARIOUS PARAMETERS DURING THREE INTENSIVE STUDY PERIODS. SUMMER 1974. FIVE-HOUR RETENTION.

	Pre-harvest ^a			First	post-har	vestb	Second post-harvest ^C			
Parameter	Influent	Control	Bulrush	Influent	Control	Bulrush	Influent	Control	Bulrush	
BOD COD	38 42	5 31	5 3 6	65	9	5	40	4	3	
Ammonia N Total organic N	1.5	2.0	1.2 0.7	0.4	0.2	0.1	0.05	0.4	0.004	
Nitrate N Turbidity (JTU) Total		1.8	2.0	6.6 23.2	3.7 3.1	9.7 2.2	9.5	4.5	3.9	
phosphorus Coliform (log	22.6	18.2	21.5	21.0	18.1	15.9	22.2	18.2	17.9	
col. 100 ml-1) Total solids Dissolved	5.69	4.47	4.69	6.41 988	4.70 873	4.77 761	6.32 860	3.76 829	4.08 909	
solids Suspended	916	930	910	758	744	687	762	740	749	
solids oH	7.44	7.85	7.45	232 7.42	168 7.67	75 7.37	93 7.50	89 7.66	165 7.52	
Conductivity (umho)	1436	1415	1440	1075	1005	950	1092	1076	1092	

aAll values not specified by parentheses are expressed as mg 1-1.

bIn most cases n = 24 samples taken at 4 hour intervals over a 4-day period.

cIn most cases n = 19 samples taken at 4 hour intervals over a 3-day period.

Table C-2. WATER BALANCE DATA FOR EXPERIMENTAL BASINS DURING TREATMENT OF SECONDARY EFFLUENT. INTENSIVE STUDIES. SUMMER 1974. (liters)

	Influent	Rainfall	Total effluent	% Evapotranspiration
Pre-harvest Control Bulrush	78 4 3 7835	0	7865 7067	0 9.8
First post harvest Control Bulrush	5352 6022	213 213	7059 5840	-28.6 6.3
Second post-harvest Control Bulrush	6109 6408	0	4705 7517	23.0 -17.0

Table C-3. EFFECTIVENESS OF EXPERIMENTAL BASINS FOR TREATMENT OF SECONDARY EFFLUENT. SIXTEEN HOUR RETENTION.

**************************************			Influent load Effluent load					%
Parameter	Basins	mg 1-1	liters	kg	mg 1-1	liters	kg	reduction
BOD	Control Bulrush	23 23	19784 26866	0.5 0.6	9 9	11533 18887	0.1	77 72
COD	Control	31	19784	0.6	42	11533	0.5	21
	Bulrush	31	26866	0.8	31	18887	0.6	29
Orthophos-	Control	25	19784	0.5	11	11533	0.1	80
phate	Bulrush	25	26866	0.7	20	18887	0.4	40
Total	Control	26	19784	0.5	12	11533	0.1	81
phosphorus	Bulrush	26	26866	0.7	22	18887	0.4	44
Total	Control	822	19784	16.3	603	11533	7.0	57
solids	Bulrush	822	26866	22.1	591	18887	11.2	49
Suspended solids	Control	75	19784	1.5	74	11533	0.8	43
	Bulrush	75	26866	2.0	79	18887	1.5	25
Dissolved solids	Control	746	19784	14.8	532	11533	6.1	62
	Bulrush	746	26866	20.0	512	18887	9.7	52

Table C-4. EFFECTIVENESS OF EXPERIMENTAL BASINS FOR TREATMENT OF SECONDARY EFFLUENT. TEN-DAY RETENTION.

Parameter	Basins	Influmg 1-1	ent loa		Efflumg 1-1	ent loa liters		% reduction
BOD	Control	31	21397	0.66	15	13013	0.19	71
	Bulrush	31	34337	1.06	11	20590	0.22	79
COD	Control	90	21397	1.92	52	13013	0.68	65
	Bulrush	90	34337	3.09	65	20590	1.33	57
Orthophos-	Control	22.6	21397	0.48	22.4	13013	0.29	40
phate	Bulrush	22.6	34337	0.78	21.9	20590	0.45	42
Total	Control	22.7	21397	0.49	23.9	13013	0.31	36
phosphorus	Bulrush	22.7	34337	0.78	26.2	20590	0.53	32
Total	Control	969	21397	20.70	910	13013	11.80	43
solids	Bulrush	969	34337	33.30	1153	20590	23.70	29
Suspended	Control	51	21397	1.09	20	13013	0.26	76
solids	Bulrush	51	34337	1.75	14	20590	0.28	84
Dissolved solids	Control Bulrush	918 918	21397 34337		890 1140	13013 20590	11.60 23.47	42 25

Table C-5. WATER BALANCE DATA FOR EXPERIMENTAL BASINS DURING TREATMENT OF SECONDARY EFFLUENT. SUMMER 1974. (liter)

	Control	Softstem	Softstem	River	Softstem
Influent	63948	51045	67449	75166	85911
Rainfall	3592	3592	3592	3592	3592
Effluent	56514	38883	60594	73909	77237
% Evapotranspiration	16.3	28.0	14.2	6.1	13.7

APPENDIX D

PILOT PLANT DATA

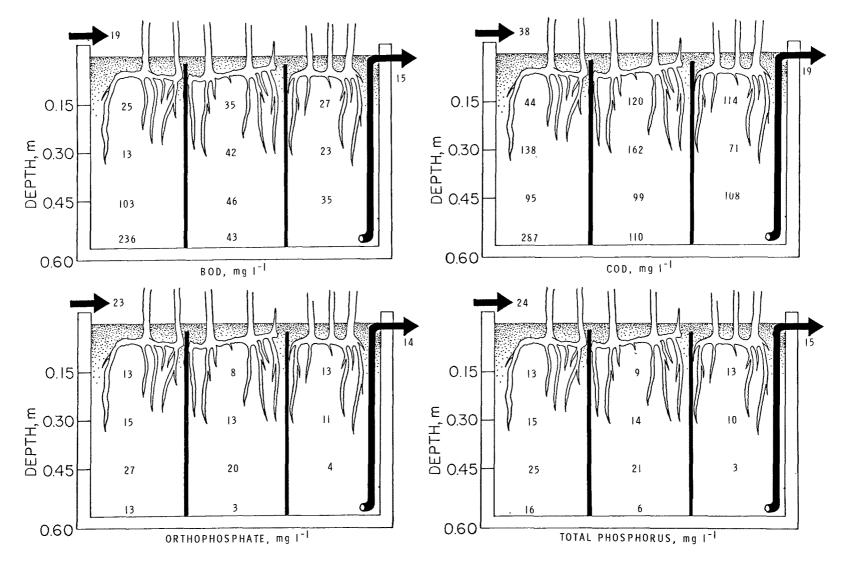


Figure D-1. Water Quality in Pilot Plant Receiving Secondary Effluent. June 5 - July 17, 1975. Each Value is the Mean of Six.

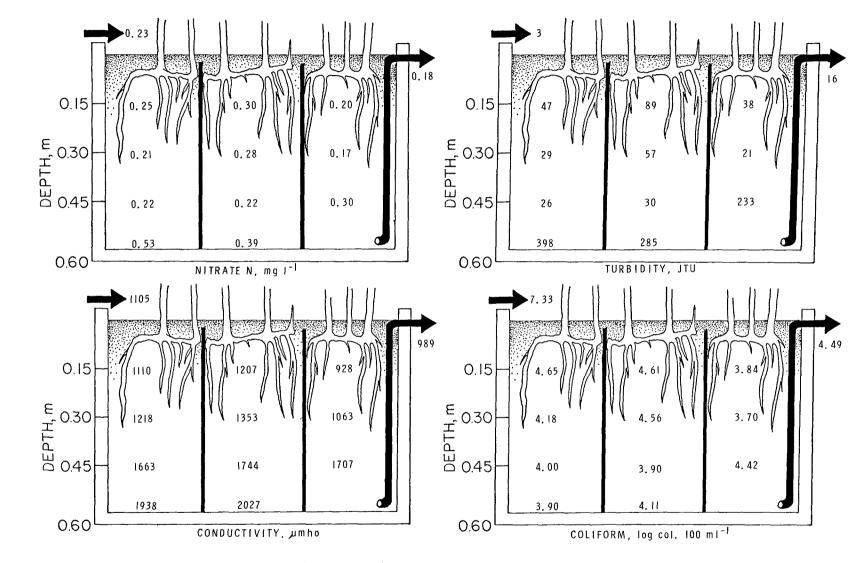


Figure D-1 (continued). June 5 - July 17, 1975.

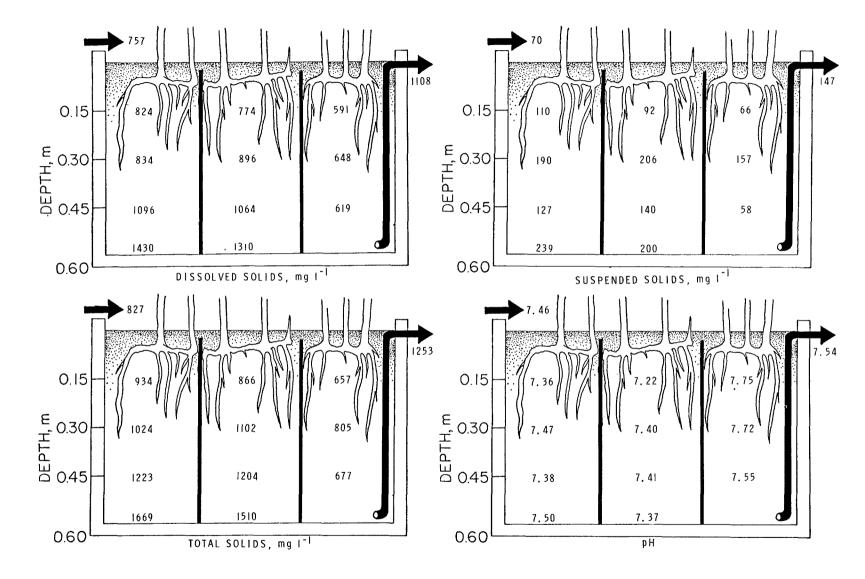


Figure D-1 (continued). June 5 - July 17, 1975.



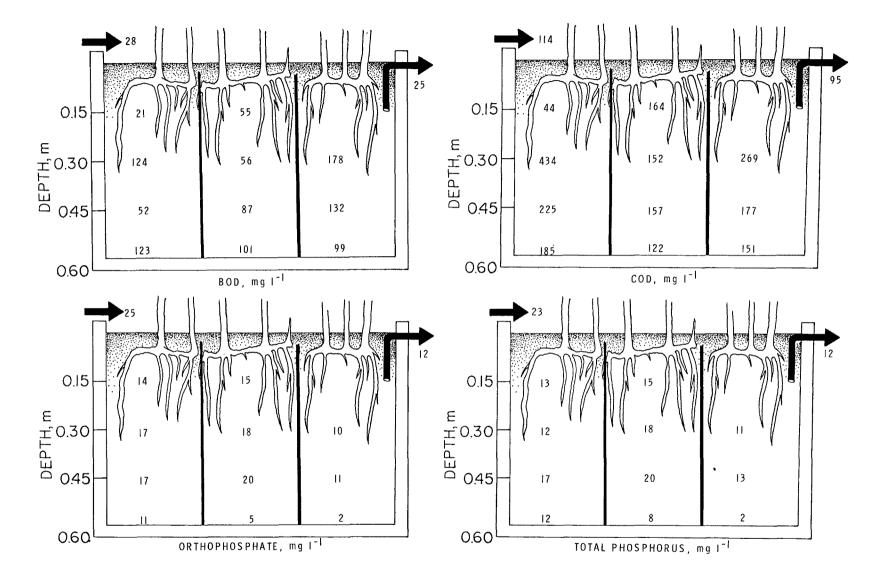


Figure D-2. Water Quality in Pilot Plant Receiving Secondary Effluent. July 17 - August 6, 1975. Each Value is the Mean of Four.

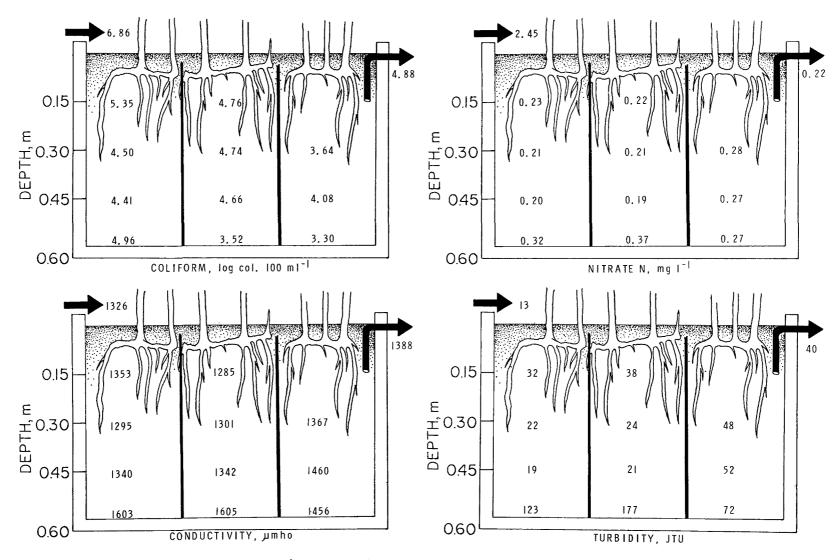


Figure D-2 (continued). July 17 - August 6, 1975.

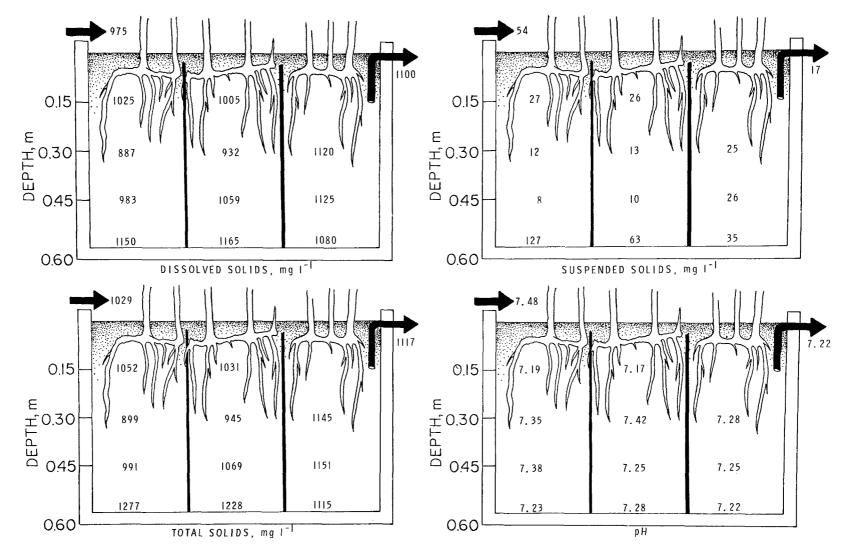


Figure D-2 (continued). July 17 - August 6, 1975.

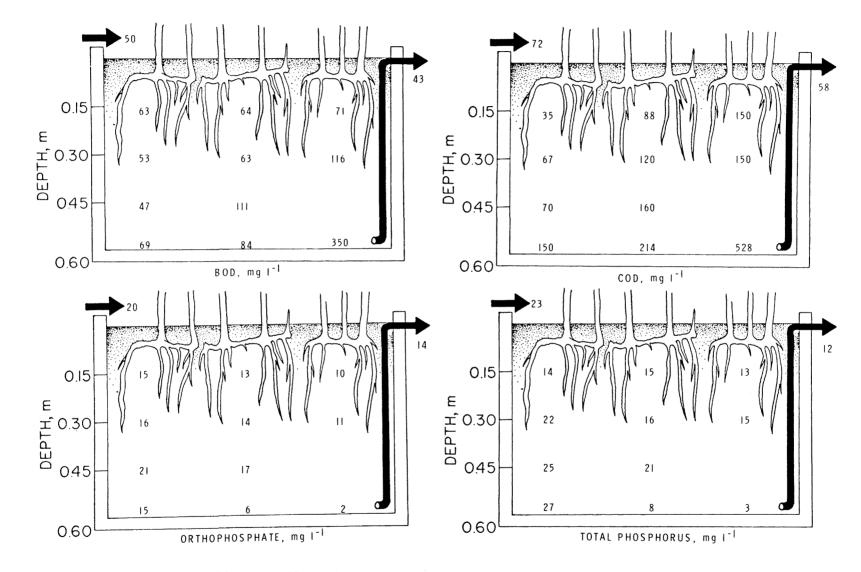


Figure D-3. Water Quality in Pilot Plant Receiving Secondary Effluent. August 6 - August 21, 1975. Each Value is the Mean of Three.

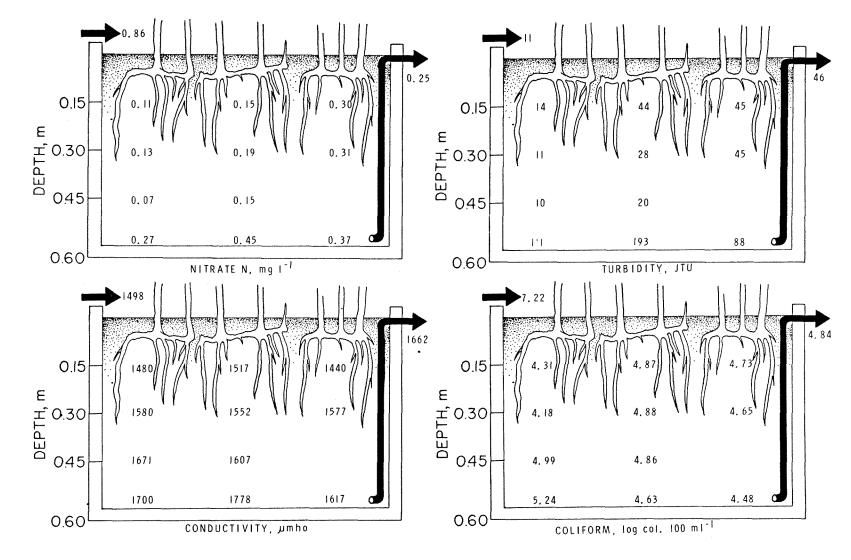


Figure D-3 (continued). August 6 - August 21, 1975.

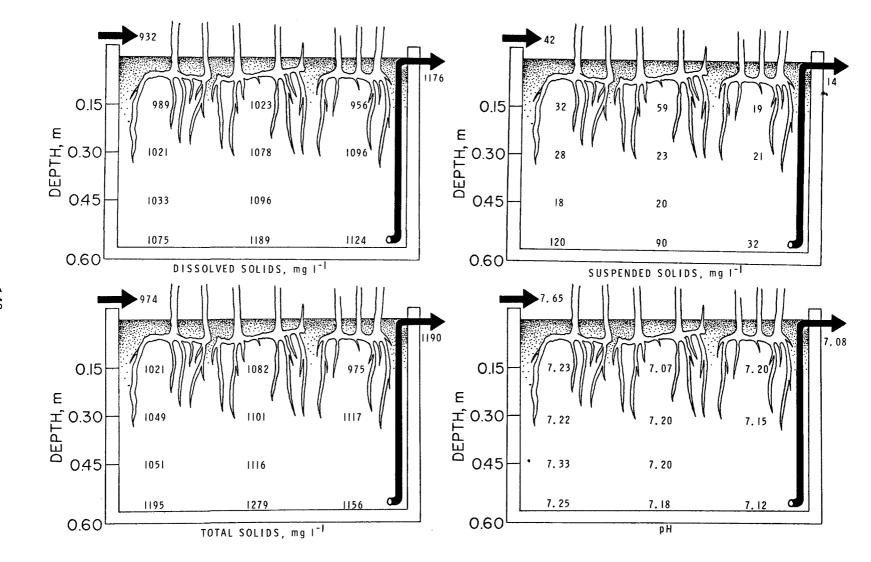


Figure D-3 (continued). August 6 - August 21, 1975.

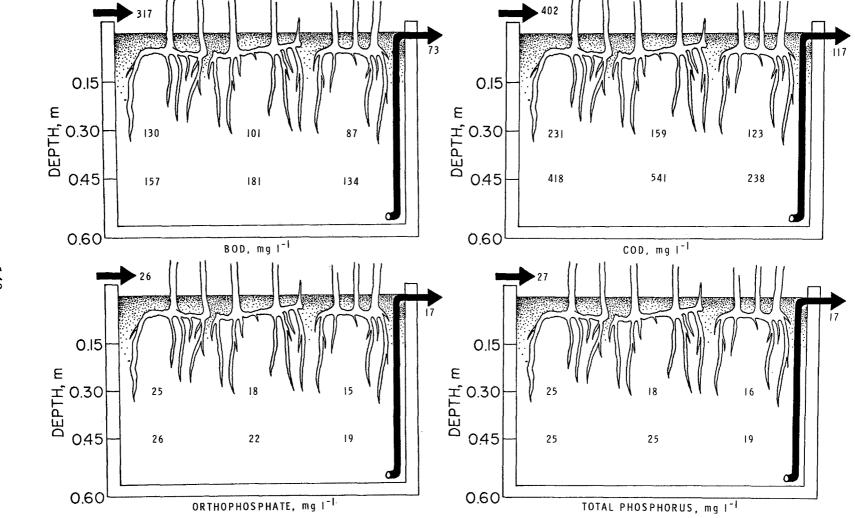


Figure D-4. Water Quality in Pilot Plant Receiving Primary Effluent. August 21 - November 4, 1975.

Each Value is the Mean of Twelve.

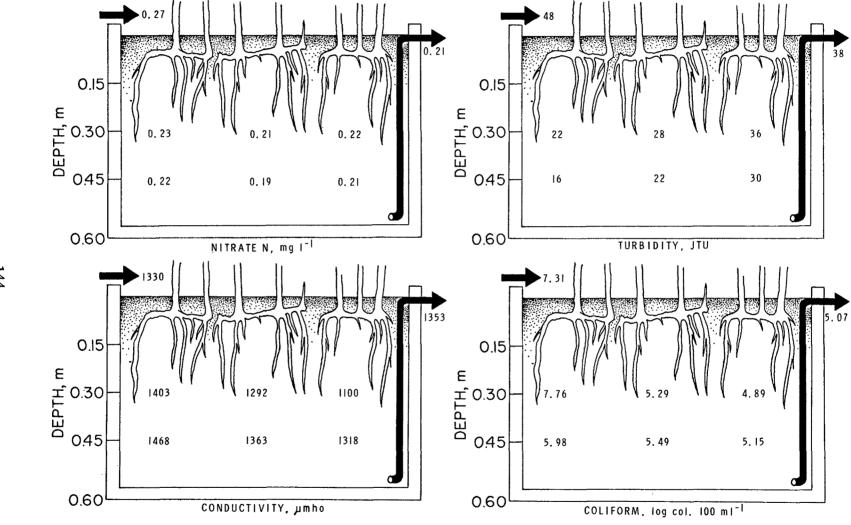


Figure D-4 (continued). August 21 - November 4, 1975.

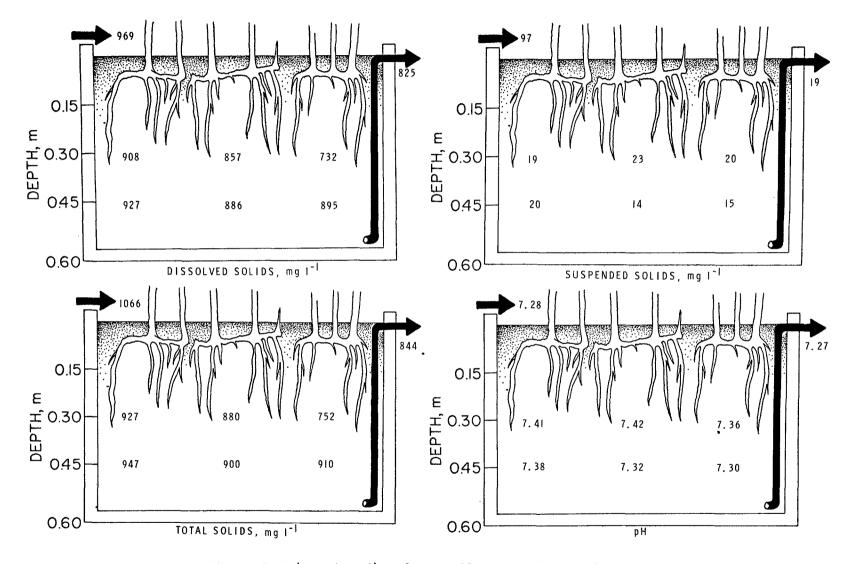


Figure D-4 (continued). August 21 - November 4, 1975.

Table D-1. EFFECTIVENESS OF PILOT PLANT IN TREATING SECONDARY EFFLUENTA. SUMMER 1974.

Parameter	Source	Meanb	Standard deviation	n	% reduction
BOD	Influent	50	24	8	
	Effluent	4	1	7	92
COD	Influent	42	22	8	
	Effluent	24	8	8	43
Orthophosphate	Influent	13.4	4.3	10	
	Effluent	8.5	3.5	9	37
Total	Influent	13.9	5.0	10	
phosphorus	Effluent	9.0	4.4	9	35
Conductivity	Influent	1313	324	7	8
(umhos)	Effluent	1210	83	7	
Turbidity	Influent	6.2	3.3	10	
(JTU)	Effluent	1.3	0.6	9	79
Ammonia	Influent Effluent	0.2 0.2	0.2 0.5		 0
Coliforms (log col. 100 ml-1)	Influent Effluent	5.58 5.63	5.49 5.94	6 5	 -9
pH ·	Influent	7.6	0.09	8	
	Effluent	7.9	0.09	7	-4

aMean values were not corrected for evapotranspiration. $^{\rm b}$ All values expressed as mg 1^{-1} unless otherwise indicated in parentheses.

Table D-2. EFFECTIVENESS OF PILOT PLANT IN TREATING PRIMARY EFFLUENTA. SUMMER 1975.

Parameter	Source	Meanb	Standard deviation	n	% reduction
BOD	Influent	317	133	10	
	Effluent	72	36	9	77
COD	Influent	401	158	12	
	Effluent	117	94	9	71
Orthophosphate	Influent	26	6	12	
	Effluent	17	3	9	35
Total	Influent	27	6	12	
phosphorus	Effluent	17	4	9	37
Conductivity (umhos)	Influent	1330	174	12	
	Effluent	1353	160	9	-2
Turbidity	Influent	48	16	12	
(JTU)	Effluent	38	5	9	21
Nitrate	Influent	0.27	0.27	10	
N	Effluent	0.21	0.09	9	22
Coliform (log col. 100 ml-1)	Influent Effluent	7.31 5.07		7 6	99.9
рН	Influent Effluent	7.28 7.27		12 9	
Total	Influent	973	113	3	
solids	Effluent	1156	19	2	-19
Dissolved	Influent	42	47	3	23
solids	Effluent	32	14	2	
Suspended	Influent	932	84	3	
solids	Effluent	1124	34	2	-21

aMean values were not corrected for evapotranspiration.
bAll values expressed as mg 1-1 unless otherwise indicated in parentheses.

Table D-3. AVERAGE VALUES FOR SEVERAL PARAMETERS AT THREE DEPTHS IN PILOT PLANT DURING AN INTENSIVE STUDY. JULY 7 - JULY 10, 1975.

		Standard		
Parameter	Meana	deviation	Low	High
T.: C1				
<u>Influent</u> BOD	12.22	5.06	7.71	27.34
COD	47.14	41.25	23.52	186.20
Turbidity (JTU)	7.35	7.34	2.00	23.00
pH	7.44	0.19	7.15	7.70
Conductivity (umho)	1200	125	900	1330
Nitrate N	2.84	1.88	0.00	6.40
Total phosphorus	24.52	4.28	17.03	32.78
Total solids Suspended solids	932.18 18.63	77.14 13.06	774.50 0.00	1096.00 38.50
Dissolved solids	913.55	80.03	746.50	1078.00
Coliform (log col. 100 ml ⁻¹)	5.96	6.06	3.32	6.60
,				
Downstream end; 0.15 m				
BOD Turbiditus (ITU)	64.67	19.20	41.34	85.33
Turbidity (JTU) pH	25.29 7.57	10.90 0.26	11.00 7.20	46.00 7.90
Conductivity (umho)	1056	119	800	1210
Nitrate N	0.26	0.07	0.17	0.37
Total phosphorus	9.46	1.50	6.92	12.75
Total solids_	874.13	113.04	510.00	1103.50
Suspended solids	28.05	19.75	0.00	82.00
Dissolved solids	846.08	137.35	469.50	1078.50
Coliform (log col. 100 ml-1)	3.85	3.88	2.47	4.34
Downstream end; 0.45 m				
BOD	14.25	6.92	8.67	33.33
COD	49.65	29.98	23.52	68.60
Turbidity (JTU)	46.89	15.80	26.00	76.00
pH	7.37	0.16	7.10	7.70
Conductivity (umho) Nitrate N	1278 0.29	122 0.07	1100 0.20	1500 0.42
Total phosphorus	8.43	1.51	5.31	10.12
Total solids	996.69	119.95	662.20	1181.00
Suspended solids	29.68	12.67	11.00	50.50
Dissolved solids	967.01	120.16	645.20	1133.50
Coliform (log col. 100 ml^{-1})	3.98	4.09	3.00	3.97

aAll values are mg l-1 unless otherwise indicated.

Table D-3 (continued). AVERAGE VALUES DURING INTENSIVE STUDY.
JULY 7 - JULY 10, 1975

Parameter	Meana	Standard deviation	1 000	Uich
TUTUTECET	nean-	deviation	Low	High
Downstream end; 0.60 m				
BOD	26.27	14.18	8.67	43.00
COD	Not tes		0.07	,0.00
Turbidity (JTU)	51.11	20.70	30.00	96.00
Н	7.32	0.19	7.10	7.55
Conductivity (umho)	1448	108	1250	1610
Nitrate N	0.32	0.07	0.21	0.48
Total phosphorus	24.52	4.28	17.03	32.78
Total solids	1181.89	107.97	1027.50	1442.50
Suspended solids	49.76	14.68	16.50	69.50
Dissolved solids	1132.13	111.71	973.50	1388.50
Coliforms (log col. 100 ml-1	3.30	3.23	2.78	3.48

aAll values are mg l-l unless otherwise indicated.

Table D-4. EFFECTIVENESS OF PILOT PLANT IN TREATING SECONDARY EFFLUENT (DRAIN AT 0.6 m).a SUMMER 1975.

			Standard		%
Parameter	Source	Meanb	deviation	n	reduction
BOD	Influent	19	15	6	
	Effluent	15	5	6	21
COD	Influent	38	21	6	
	Effluent	41	18	6	42
Orthophosphate	Influent	23	8	7	
	Effluent	14	5	7	39
Total	Influent	24	7	7	
phosphorus	Effluent	15	5	7	38
Conductivity (umhos)	Influent Effluent	1125 989	158 283	7 7	10
Turbidity	Influent	4.6	5	7	
(JTU)	Effluent	15.5	11	7	-474
Nitrate N	Influent	0.32	0.26	7	
	Effluent	0.18	0.06	6	22
Coliforms (log col. 100 ml-1)	Influent Effluent	6.2 3.90	3.4 3.2	7 7	 99.9
рН	Influent Effluent	7.42 7.54	0.18 0.15	7 7	
Total	Influent	827	48	7	
solids	Effluent	1253	162	7	-52
Dissolved	Influent	757	72	7	
solids	Effluent	1107	201	7	-46
Suspended	Influent	70	43	7	
solids	Effluent	147	151	7	-110

aMean values were not corrected for evapotranspiration.

bAll values expressed as mg l-l unless otherwise indicated in parentheses.

Table D-5. EFFECTIVENESS OF PILOT PLANT IN TREATING SECONDARY EFFLUENT (DRAIN AT 0.15 m)a. SUMMER 1975.

Parameter	Source	Meanb	Standard deviation	n	% reduction
BOD	Influent	28	19	4	
	Effluent	25	16	4	11
COD	Influent	115	98	5	
	Effluent	95	89	3	17
Orthophosphate	Influent	25	10	5	
	Effluent	12	1	3	52
Total	Influent	24	8	5	
phosphorus	Effluent	12	1	3	50
Conductivity	Influent	1326	240	5	
(umhos)	Effluent	1388	183	3	-5
Turbidity	Influent	13	19	5	
(JTU)	Effluent	40	14	3	-208
Nitrate N	Influent	2.45	4.70	5	
	Effluent	0.22	0.04	3	91
Total Coliforms (log col. 100 ml ⁻¹)	Influent Effluent	6.86 4.88	6.66 6.00	4 3	 99.9
рН	Influent Effluent	7.48 7.22	0.23 0.16	5 3	
Total	Influent	1029	180	4	
solids	Effluent	1117	90	4	-9
Suspended	Influent	54	87	4	
solids	Effluent	18	10	4	68
Dissolved	Influent	974	112	4	
solids	Effluent	1099	83	4	-13

 a Mean values were not corrected for evapotranspiration. b All values expressed as mg $^{1-1}$ unless otherwise indicated in parentheses.

Table D-6. EFFECTIVENESS OF PILOT PLANT IN TREATING SECONDARY EFFLUENT (PARTITIONS REMOVED)a. SUMMER.

Parameter	Source	 Mean ^b	Standard deviation	n	% reduction
Parameter	Source	Means	deviación	<u>n</u>	reduction
BOD	Influent	50	11	3	
	Effluent	4 3	34	3	14
COD	Influent	72	33	3	
	Effluent	58	9.3	3	19
Orthophosphate	Influent	20	0.58	3	
	Effluent	14	2.9	3	30
Total	Influent	23	5.9	3	
phosphorus	Effluent	12	0.91	3	48
Conductivity (umhos)	Influent Effluent	1498 1662	194 46	3 3	-11
Turbidity	Influent	11	11	3	-318
(JTU)	Effluent	46	8.1	3	
Nitrate N	Influent	0.86	0.08	2	
	Effluent	0.25	0.15	3	71
Coliforms (log col. 100 ml-1)	Influent Effluent	7.2 4.8	7.5 4.4	3 3	 99.9
рН	Influent Effluent	7.7 7.1	0.17 0.10	3 3	
Total	Influent	1066	147	12	21
solids	Effluent	845	309	10	
Dissolved	Influent	970	121	12	
solids	Effluent	8 2 5	303	10	15
Suspended solids	Influent Effluent	97 19	62 18	12 10	4.1

aMean values were not corrected for evapotranspiration. bAll values expressed as mg l-1 unless otherwise indicated in parentheses.

Table D-7. WATER BALANCE DATA FOR PILOT PLANT. (liters)

Sampling dates	Influent	Rainfall	Total effluent	% evapotranspiration
June 5 - Aug. 31 1974	607227	22301	604468	4.0
June 5 - July 17 1975	119200	22153	108255	23.4
July 17 - Aug. 6 1975	57036	1628	39376	32.8
Aug. 6 - Aug. 21 1975	44924	6154	35919	29.7
Aug. 21 - Nov. 4 1975	188273	37019	187376	16.8

APPENDIX E DATA RELATED TO PHOSPHORUS DISTRIBUTION

DISTRIBUTION OF DRY MATTER IN GRAVEL PILOT PLANT $(g \ dw \ m^{-2})$ Table E-1.

Depth	Basin A	Basin B	Basin C	Mean
0-15 cm ^a	533	1385	944	970
15-30 cm ^b	1761	1427	1722	1636
30-45 cm ^C	1037	2806	1449	1764
45-60 cm ^C	1037	2806	1449	1764
Total	4368	8424	5614	6135

aEach value is the mean of three.

bEach value is the result of analysis of a single sample. CEach value represents a single sample taken at 45 cm and assumed to be representative of both depth intervals.

Table E-2. DISTRIBUTION OF ORGANIC MATTER IN GRAVEL PILOT PLANT (g ash-free dw m-2)

Depth	Basin A	Basin B	Basin C	Mean
0-15 cm ^a	423	1072	697	731
15-30 cm ^b	1599	1295	1549	1481
30-45 cm ^c	888	2427	1149	1488
45-60 cm ^C	888	2427	1149	1488
Total	3798	7221	4544	5188

aEach value is the mean of three.

bEach value is the result of analysis of a single sample.

CEach value represents a single sample taken at 45 cm and assumed to be representative of both depth intervals.

Table E-3. NUMBER AND SIZE OF BULRUSH SHOOTS IN PILOT PLANT ON AUGUST 8, 1975.

	Length	Diameter	Number
	(m)	(cm)	Shoots (m-2)
East Basin			
Sample A	0.89	0.77	124
B	1.26	0.68	532
C	1.44	<u>0.74</u>	604
Mean	1.20	0.73	420
<u>Central Basin</u>			
Sample A	1.05	0.78	232
B	0.99	0.60	620
C	0.93	<u>0.89</u>	<u>76</u>
Mean	0.99	0.76	308
West Basin			
Sample A	0.81	0.50	512
B	0.73	0.56	140
C	0.83	<u>0.66</u>	<u>72</u>
Mean	0.79	0.57	240

APPENDIX F

BRILLION MARSH DATA

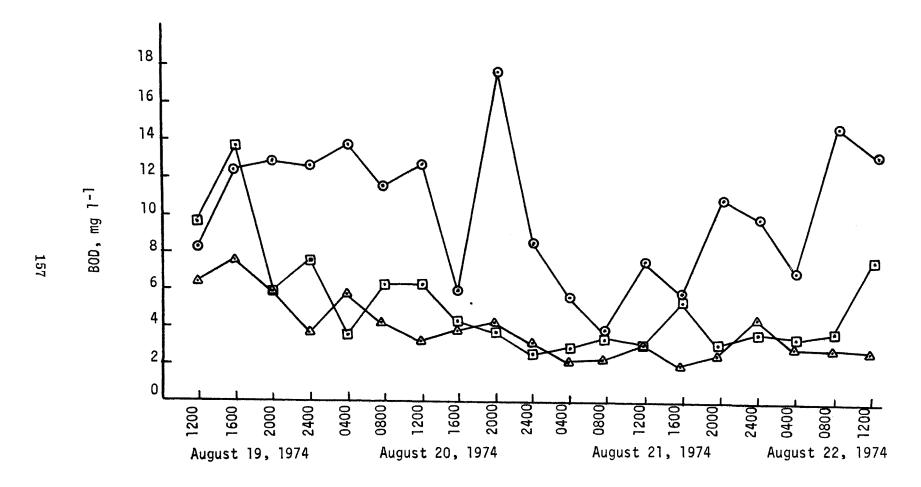


Figure F-1. Short Term Variation in BOD Concentration in the Brillion Marsh Study.

(△ - Station I; ⊙ - Station II; □ - Station III)

Figure F-2. Short Term Variation in Total Phosphorus Concentration in the Brillion Marsh Study. (\triangle - Station I; \bigcirc - Station II; \bigcirc - Station III)

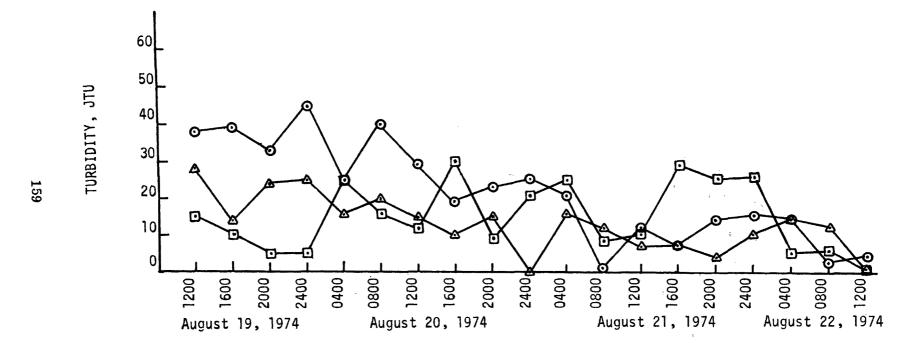


Figure F-3. Short Term Variation in Turbidity in the Brillion Marsh Study. (\triangle - Station I; \bigcirc - Station II; \bigcirc - Station III)

Figure F-4. Short Term Variation in Conductivity in the Brillion Marsh Study.
(△- Station I; ⊙ - Station II; ☐ - Station III)

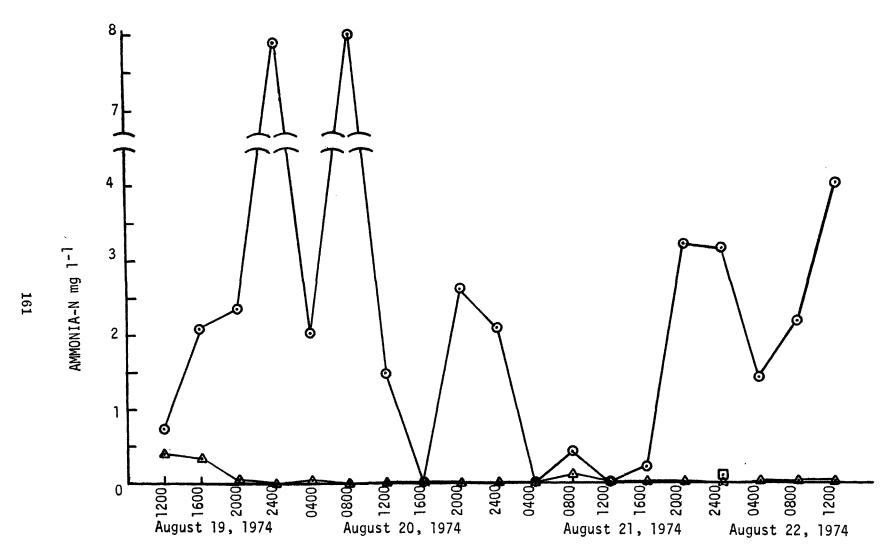


Figure F-5. Short Term Variation in Ammonia Concentration in Total Solids in the Brillion Marsh Study. (\triangle - Station I; \bigcirc - Station II; \bigcirc - Station III)

Figure F-6. Short Term Variation in Nitrate Concentration in Total Solids in the Brillion Marsh Study. (△- Station I; ⊙- Station II; ⊡- Station III)

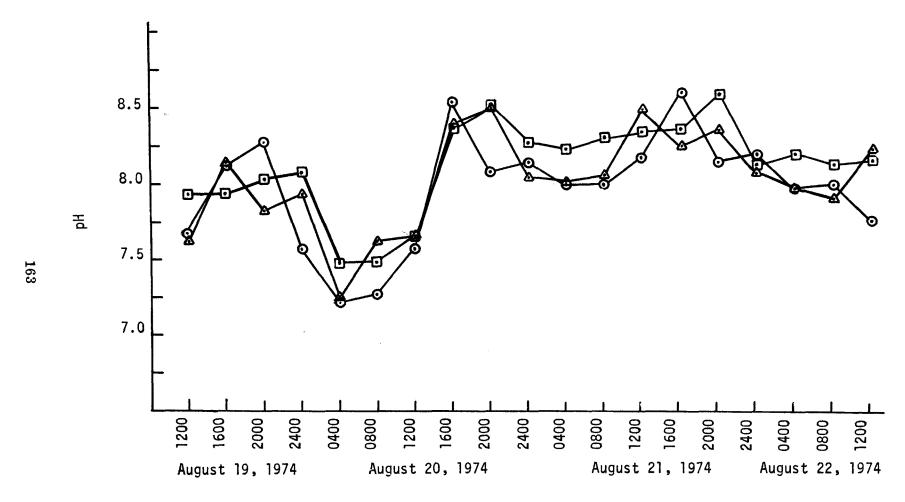


Figure F-7. Short Term Variation in pH in the Brillion Marsh Study. (△- Station I; ⊙ - Station II; ⊡ - Station III)

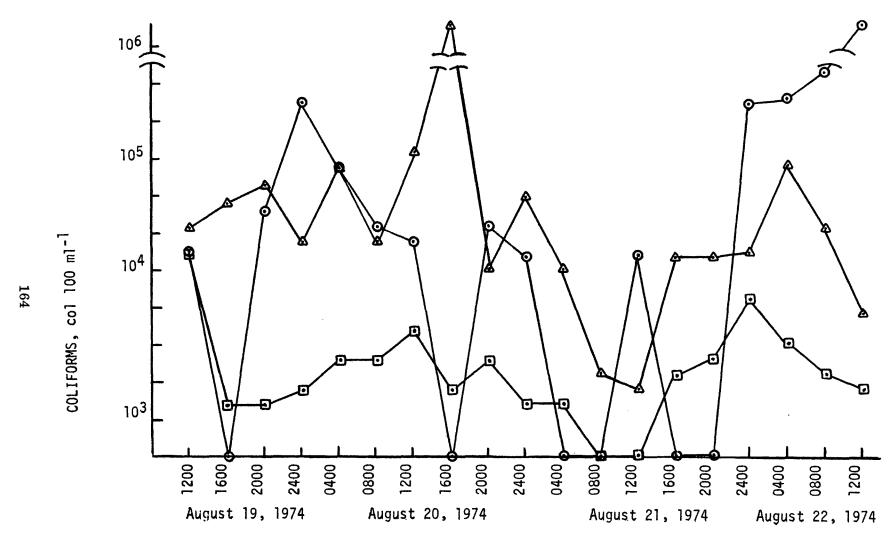


Figure F-8. Short Term Variation in Coliform Levels in the Brillion Marsh Study.

(△ - Station I; ⊙ - Station II; □ - Station III)

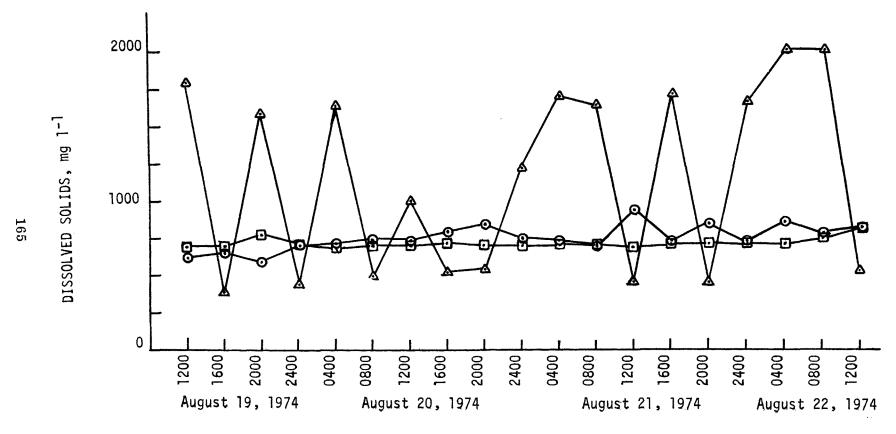


Figure F-9. Short Term Variation in Dissolved Solids Concentration in the Brillion Marsh Study. $(\Delta$ - Station I; \odot - Station II; \odot - Station III)

Figure F-10. Short Term Variation in Suspended Solids Concentration in the Brillion Marsh Study. $(\Delta$ - Station I; \odot - Station II; \odot - Station III)

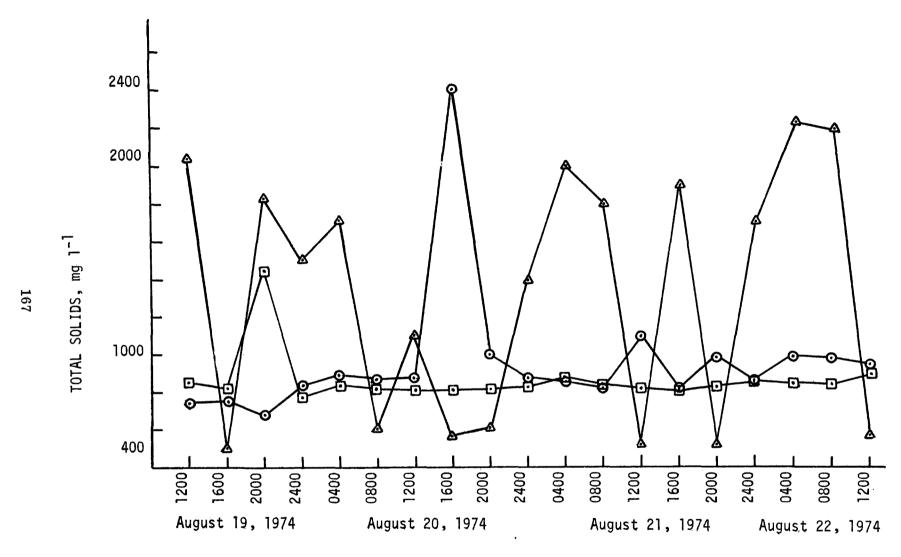


Figure F-11. Short term Variation in Total Solids Concentration in the Brillion Marsh Study. $(\Delta$ - Station I; \odot - Station II; \odot - Station III)

Table F-1. CHEMICAL ANALYSIS OF SAMPLES FROM STATION I LOCATED ON SPRING CREEK ABOVE THE BRILLION TREATMENT PLANT.

	B0D	COD	Total phosphorus	Ortho- phosphate	Conduc- tivity (umho)	Turbidity (JTU)	Nitrate	На	Total solids	Suspended solids	Dissolved solids	Coliform (log co].
June 19, 1974 July 1 July 9 Aug. 1 Sept. 11 Sept. 28 Oct. 8 Oct. 29 Nov. 20 Nov. 21 Dec. 17 Feb. 11, 1975	6.3 14.9 4.0 3.6 14.2 7.1 6.7 11.5 5.5 5.9 7.2	34 57 4 25 36 62 21 34 24 0 9	0.92 1.08 0.76 0.11 0.27 0.26 1.14 2.26 3.61 1.61 0.08 0.03	0.84 1.23 0.64 0.80 0.27 0.36 1.31 1.78 2.92 1.46 0.08	800 500 1750 3100 2700 850 1750 3000 760 2970 950	11 8 5 5 9 9 8 21 5 7 6	1.03 1.23 2.20 2.44	7.70 8.80 8.00 7.78 7.96 8.00 8.08 7.64 8.21 8.01 8.06 7.13				4.40 4.63 5.43 4.45 3.04 3.18 3.80 3.65 4.45
Mar. 4 Apr. 22 June 5 June 12 June 19 June 26 July 1 July 17 July 29 Aug. 5 Aug. 28	5.1 4.8 6.7 4.9 4.7 2.2 2.5 3.8 2.2 2.7	4 25 21 44 47 34 4 14 22 35 147	0.05 0.11 0.78 0.38 0.38 0.16 0.19 0.25 12.76 1.03	0.02 0.07 0.66 0.40 0.32 0.16 0.12 0.17 12.98 0.07	775 1000 800 1800 750 2900 750 1800 1725 2150 282	6 7 16 13 9 5 6 6 5 420	1.54 0.84 4.25 0.94 1.55 1.24 0.95 1.55 3.40	7.90 7.00 8.20 7.70 7.85 8.24 8.25 7.85 7.95 8.10 7.50	567 1080 782 570 515 1163 1209 1409 1303	93 214 80 87 20 14 1 12 869	474 866 702 483 495 1149 1208 1397 434	4.75 4.40 4.85 5.49 4.42 4.00 3.19

aAll values are expressed as mg 1-1 unless otherwise indicated in parentheses.

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Table F-2. CHEMICAL ANALYSIS OF SAMPLES FROM STATION II LOCATED ON SPRING CREEK BELOW THE BRILLION SEWAGE TREATMENT PLANT.

	вор	COD	Total phosphorus	Ortho- phosphate	Conduc- tivity (umho)	Turbidity (JTU)	Nitrate	Нd	Total solids	Suspended solids	Dissolved solids	Coliform (log col 100 ml-1)
June 19, 1974 July 1	7.2 9.8	30.4 52.5	0.79	0.65	850	15 15		8.00 8.75				5.20
July 9 Aug. 1 Sept. 11	2.8	15.7 49.3	1.56 3.68 3.71	0.82 3.39 3.71	570 900	6 16 15		7.93 7.73				5.99 4.80 5.80
Sept. 11 Sept. 28 Oct. 8	13.3 22.0 74.0	102.6 87.4 130.2	2.58 8.10	2.42 6.48	1450 1450 1000	16 30	0.48	8.01 7.76 7.70				4.80 3.48
Oct. 29 Nov. 20	62.5 100.0	115.6 64.6	5.62 8.20	5.62 6.95	1850 1100	37 11	0.41	7.46 8.00				3.62
Nov. 21 Dec. 17	51.5 23.0	137.3 79.2	6.27 2.54	5.51	1250 1200	17 16	0.91	7.85 7.90				3.70 3.84
Feb. 11, 1975 Mar. 4 Apr. 22	18.3 39.5 11.7	45.4 90.2 34.8	1.06	0.90 3.95 0.80	1100 1160 <i>7</i> 25	9 20 9	1.20	7.65 7.90 7.90				
June 5 June 12	13.0 25.0	23.0 64.0	2.90 3.45	2.73 3.24	775 825	20 37	1.19	7.85 7.55	815 601	249 119	566 402	6.07
June 19 June 26	9.2 15.8 15.3	48.2 41.8 17.3	0.81 3.51 3.23	0.60 3.41 2.98	700 1320	12 10 12	5.30 0.56 1.40	7.80 8.12 7.65	638 628 706	36 87	602 541	4.34 6.02
July 1 July 17 July 29	7.0	11.9	0.60	0.53 10.46	650 1700 1325	7 14	0.40	7.75 7.65	1022 805	138 90 76	568 932 730	5.90 4.36 5.81
Aug. 5 Aug. 28	38.5 20.5	64.0 71.3	2.13	2.04	1200 319	13 12	0.18	7.60 7.25	807 341	126 140	681 201	4.93 5.23

aAll values are expressed as mg l-1 unless otherwise indicated in parentheses.

Table F-3. CHEMICAL ANALYSIS OF SAMPLES^a FROM STATION III LOCATED ON THE OPEN CHANNEL BELOW THE UPPER PORTION OF BRILLION MARSH.

	800	000	Total phosphorus	Ortho- phosphate	Conduc- tivity (umho)	Turbidity (JTU)	Nitrate	赿	Total solids	Suspended solids	Dissolved solids	Coliform (10g col. 100 ml-1)
June 19, 1974	4.2	52.5	1.15	0.94		4		7.90				
July 1	2.7	52.5	2.20	1.93	500	4		8.50				4.08
July 9	4.3	66.6	2.95	2.42	600	3		7.49				4.04
Aug. 1	5.0	76.8	0.97	0.88	825	5		7.76				3.49
Sept. 11	4.9 2.9	34.0	0.48	0.47	1500	3 5 5 5		8.30				3.00
Sept. 28 Oct. 8	2.5	31.7 47.6	0.42 1.80	0.47 1.16	1310 1600	5 6	0.48	7.96				4.26 2.00
Oct. 29	3.7	40.4	1.25	1.15	1500	5	0.54	7.85				2.00
Nov. 20	2.3	32.1	1.83	1.53	1325	2	1.04	8.01				3.00
Nov. 21	2.6	48.6	1.02	1.04	1375	2	1.25	8.10				3.04
Dec. 17	4.0	93.4	1.67	1.98	1525	7	1.04	7.70				3.15
Feb. 11, 1975		61.5	3.88	3.60	1125	26		7.20				• • • • • • • • • • • • • • • • • • • •
	29.0	13.7	4.90	4.30	1600	12		7.00				
Apr. 22	8.7	40.4	0.42	0.32	640	5		8.00				
June 5	1.7	72.0	2.56	2.94	850	4		7.85	597	47	550	
June 12	3.8	61.4	1.95	2.34	800	40	0.46	7.50	550	90	459	4.10
June 19	1.3	70.3	2.01	1.91	825	14	0.03	7.25	759	87	672	3.84
June 26	5.6	92.2	7.33	9.10	890	23	0.41	7.88	518	46	472	4.74
July 1	7.1	53.4	11.85	10.27	775	30	1.70	7.60	957	121	836	5.35
July 17	6.7	118.8	2.43	2.33	825	4	0.04	7.25	1072	15	1056	4.68
July 29		102.8	10.52	10.85	825	16	0.15	8.00	972	7	964	4.70
Aug. 5	8.4	102.8	1.56	1.67	875	24	0.01	7.65	820	29	791	4.95
Aug. 28	4.3	47.5	0.43	0.79	610	7	0.45	7.45	659	365	294	5.44

aAll values are expressed as mg l-1 unless otherwise indicated in parentheses.

Table F-4. CHEMICAL ANALYSIS OF SAMPLES® FROM THE OUTFLOW PIPE OF THE BRILLION SEWAGE TREATMENT PLANT.

	вор	000	Total phosphorus	Ortho- phosphate	Conduc- tivity (umho)	Turbidity (JTU)	Nitrate	Ha	Total solids	Suspended solids	Dissolved solids	Coliform (log col. 100 ml-1)
July 9, 1974	55	133	9.82	9.31	950	39		7.54				
Sept. 11	82	173	7.23	7.23	1000	35		7.78				
Sept. 28	137	284	10.11	9.50	940	52		7.48				3.60
Oct. 8	170	309	12.08	9.91	1130	58	0.66	7.60				
Oct. 29	128	291	8.29	8.29	1100	53	0.61	7.60				5.20
Nov. 20	112	234	15.84	13.50	1100		0.48					3.90
Nov. 21	81	149	8.76	8.21	1100	21	0.44	8.01				3.47
Feb. 11, 1975	132	317	12.86	8.80	1100	62		7.60				
Mar. 4	112	293			1100	54		7.50				3.70
Apr. 22	61	175	6.19	5.35	1025	46	0.46	7.35				
June 5	36	97	13.04	10.78	800	24		7.50	641	75	566	
June 19	19	115	8.01	6.68	800	28	0.40	7.65	784	33	751	3.00
June 26	47	137	8.78	7.03	990	28	0.30	8.02	484	45	439	2.30
July 1	58	157	12.41	12.47	900	40	1.00	7.60	611	72	.539	2.00
July 17	70	184	10.82	9.29	1425	38	0.09	7.30	623	30	593	4.11
July 29		234	1.26	1.03	1075	41	0.10	7.60	710	69	642	3.43
Aug. 5		262	7.40	6.00	975	62	0.20	7.60	769	36	733	4.34
Aug. 28	27	57	1.23	0.96	242	76	1.90	7.15	469	12	457	6.14

aAll values are expressed as mg l-1 unless indicated otherwise in parentheses.

(F	TECHNICAL REPORT DATA lease read Instructions on the reverse before com	pleting)
1. REPORT NO. EPA-600/2-76-207	2.	3. RECIPIENT'S ACCESSION•NO.
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7. AUTHOR(S) Frederic L. Spang William E. Sloey C. W. Fetter, Jr.		8. PERFORMING ORGANIZATION REPORT NO.
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15. SUPPLEMENTARY NOTES

16. ABSTRACT

Investigations were conducted on the use of artificial and natural marshes as purifiers of effluent from municipal treatment plants. Observations were made on marsh influent and effluent quality. Phosphorus distribution in the ecosystem and removal by harvesting were studied. Responses of the vegetation to repeated harvesting were recorded.

Artificial marshes consisted of plastic-lined excavations containing emergent vegetation, especially <u>Scirpus validus</u>, growing in gravel. Various combinations of retention time, primary effluent, secondary effluent, basin shape, and depth of planting medium were studied. A polluted natural marsh was studied simultaneously.

The degree of improvement in water quality suggests that the process may be acceptable for certain treatment applications. Harvesting was not a practical phosphorus removal technique. Marshes remove phosphorus in the growing season, but release it at other times. Development of management techniques for successful use of marshes for wastewater treatment is thought possible.

17.	7. KEY WORDS AND DOCUMENT ANALYSIS								
a.	DESCRIPTORS	b.identifiers/open ended terms	c. COSATI Field/Group						
	Effluents sewage treatment Water pollution Primary biological productivity Pilot plant Marshes, phosphorus	Aquatic macrophytes Emergent vegetation Nutrient removal	13B						
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