

# **EFFECTS OF PESTICIDES IN WATER**

**A Report to the States**



**U.S. ENVIRONMENTAL PROTECTION AGENCY**

**WASHINGTON D.C. 20460**

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## FOREWORD

The Water Quality Improvement Act of 1970 amended the Federal Water Pollution Control Act by adding a new subsection 5(1). The Federal Water Pollution Control Act Amendments of 1972 modified the language of Subsection 5(1) and renumbered it as Subsection 104(1) of the Federal Water Pollution Control Act. Subsection 104(1) reads as follows:

(1)(1) The Administrator shall, after consultation with appropriate local, State and Federal agencies, public and private organizations, and interested individuals, as soon as practicable but not later than January 1, 1973, develop and issue to the States for the purpose of carrying out this Act the latest scientific knowledge available in indicating the kind and extent of effects on health and welfare which may be expected from the presence of pesticides in the water in varying quantities. He shall revise and add to such information whenever necessary to reflect developing scientific knowledge.

(2) The President shall, in consultation with appropriate local, State and Federal agencies, public and private organizations, and interested individuals, conduct studies and investigations of methods to control the release of pesticides into the environment which study shall include examination of the persistency of pesticides in the water environment and alternatives thereto. The President shall submit reports, from time to time, on such investigations to Congress together with his recommendations for any necessary legislation.

This document is issued to fulfill the requirement of paragraph 104(1)(1). A document entitled Pesticides in the Aquatic Environment has been prepared for submission to the Congress by the President to fulfill the requirement now contained in paragraph 104(1)(2). Any person interested in the problems associated with pesticides in the aquatic environment may wish to read both of these documents.

The scientific information contained in this document consists of current knowledge of the effects on health and welfare of the presence of pesticides in water. It must be emphasized, however, that many other factors must be considered in reaching decisions as to whether to undertake particular control measures. Some of the more important considerations are:

- The nature of the environmental effect of the presence of pesticides in water (e.g., long or short term, temporary or permanent, localized or widespread, etc.).
- The economic and social impact of the control measure, including both impact associated with restricting use of pesticides, and the impact of the environmental damage to be alleviated.
- The practicality and enforceability of the control measure, including the availability of techniques and instrumentation for determining whether particular standards are being met.

-- The availability of other control measures for meeting the same objectives, and the relationship to, and consistency of the control measure with, policy and action under other programs. Especially to be kept in mind, in this regard, is the national program administered by the Environmental Protection Agency pursuant to the Federal Insecticide Fungicide and Rodenticide Act (FIFRA). Historically, this program has controlled the distribution and labeling of pesticides through a case by case registration process. The major extension and revision of the Environmental Protection Agency's authority in the pesticide field provided for in the recent amendments to FIFRA will vastly improve the means available to EPA for protecting the aquatic environment.

Thus, this document provides available information to the States for the purpose of carrying out the Federal Water Pollution Control Act, but it provides information on only one of several factors to be considered in determining whether to undertake a given control measure. It does not recommend the adoption of particular standards or other types of control measures.

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## INTRODUCTION

Because water is used by man for many different purposes, pesticides in water may affect health and welfare in a variety of ways. For convenience, we can discuss the effects of pesticides according to the various uses of water, whether the water is withdrawn for use or is used where it is. Water for municipal, industrial, and agricultural uses is drawn from surface or sub-surface sources; effects of pesticides on health, industrial processes, domestic animals and crops are of interest here.

Water in place is useful for recreation, aesthetic purposes, and propagation of fish and other aquatic life; the effects of pesticides on organisms living in, or dependent upon organisms living in, water are of concern. In addition to the obvious effects on fish or shellfish and the well-known transfer of certain pesticides to man or other mammals and birds through food chains, effects on other biological processes such as decomposition, energy transfer, mineral cycling, and photosynthesis are of interest.

The effects of pesticides in potable water and irrigation waters are not dealt with here because that subject is addressed specifically in setting tolerances for pesticides in potable water, fish, shellfish, meat, and poultry, and crops watered by irrigation water under authority provided by the Federal Food, Drugs and Cosmetic Act (see 408, 409, 68 Stat. 512; 21 USC 346). For this report it is sufficient to note that: (a) in regulating human intake of pesticides, all routes must be considered -- not just water. The amount taken in from drinking water is relatively small compared to other sources; (b) fish and other aquatic organisms are more influenced by pesticide levels in water than are other organisms, including man. (Aquatic organisms are continuously submersed in water and its associated contaminants, and thus are subjected to the contaminants through their gills and body surfaces as well as through their food.) Thus, levels of pesticides in water sufficiently low to protect fish and aquatic life are generally more than adequate to protect man. In the unusual event that drinking water supplies are drawn from waters that cannot support fish and aquatic life, special attention must be paid to assuring that drinking water standards for pesticides are met.

Pesticides are chemicals, natural and synthetic, used to control or destroy plant and animal life considered adverse to human society. Since the 1940's, a large number of new synthetic organic compounds have been developed for pesticide purposes. While there are approximately 33,000 registered formulations incorporating nearly 900 different chemicals, 25 substances account for 75 percent of U.S. production. Production and use of pesticides increased annually from 1957 through 1968; in 1969 and 1970 pesticide production declined.

The maximum pesticide levels found in 529 samples of surface water collected annually at approximately 100 stations during a five-year period are shown in Appendix Table 1. Coupling these figures with a 2-liter/day water consumption, the daily intake per person for each pesticide is computed. For comparison, the average daily intake of pesticides from food is shown, as is the acceptable daily intake as established by WHO-FAO expert committees.

The final column shows that intake from water would be only 1/6 that from food in the case of aldrin-dieldrin; 1/10 that from food in heptachlor-heptachlor epoxide; and 1/20 that of DDT+DDE+DDD.

Because average levels in water are far below these maximum figures, the actual contribution of water to pesticide intake is much less. Furthermore, drinking water treatment removes suspended matter (in which much pesticide residue is adsorbed) from the raw water and therefore reduces still further the actual amount reaching man.

Pesticides are used for a wide variety of purposes in a multitude of environmental situations. Often they are categorized according to their use or intended target (e.g., insecticide, herbicide, fungicide), but their release in the environment presents an inherent hazard to many non-target organisms. Some degree of contamination and risk is assumed with nearly all pesticide use. The risk to aquatic ecosystems is dependent upon the chemical and physical properties of the pesticide formulation; weather conditions, methods of application, and other factors influencing the amount reaching the system; and the nature of the receiving system.

The pesticides of greatest concern are those which are persistent for long periods and accumulate in living organisms and the environment; those which are highly toxic to man, fish and wildlife; and those which are used in large amounts over broad areas.

The majority of these compounds are either insecticides or herbicides which are used extensively in agriculture, in public health and for household or garden purposes. Generalization about such a diverse group of chemicals is subject to many contradictions, but some generalization is required to serve as a guide for managing pesticide residues in water. In any final consideration, however, each pesticidal formulation must be considered individually according to information on its behavior in the environment and its effects on man and other organisms. The benefits to be derived from its use also should be taken into account.

## EFFECTS OF PESTICIDES IN AQUATIC SYSTEMS

The biota of aquatic systems is the result of complex evolutionary processes in the course of which organisms tolerating many different conditions have evolved. Thus, under natural conditions, the biota of a given body of water is made up of an aggregation of organisms adapted to, and in dynamic balance with, the environmental conditions.

As conditions change, organisms unable to tolerate the new conditions are eliminated, and new organisms suited to these new conditions replace them. With evolutionary rates of change, many of the earlier organisms evolve to tolerate the new conditions; with rapid changes, such as many of those caused by man, most new organisms arrive by immigration rather than evolution. Bacteria are an exception and often accommodate to new conditions by very rapid evolution. When the rate of change is very rapid, or the change very severe, relatively few organisms survive. Depending upon the nature of the area and accessibility to replacements, immigration may not provide replacements immediately. When the change consists of the introduction of substances toxic to a wide variety of organisms, the biota may be diminished greatly, as to both numbers of species and number of individuals within species; if the toxic substance remains in the system (is "persistent" or is continuously replaced), the lowered populations and numbers of species may continue until the material is sufficiently diluted, sequestered, or detoxified to permit repopulation of the systems. If the area affected is large, return to the earlier condition may take a very long time, and some species may have been extirpated. If the toxic material degrades rapidly, depopulation will be temporary if the area affected is small.

### Behavior of Pesticides in the Environment

Many pesticides have a very low water solubility, and often are rapidly sorbed on suspended or sedimented materials; those with high fat solubility often accumulate in plant and animal lipids. Soluble or dispersed fractions in the water rapidly diminish after initial contamination resulting in increased concentrations in the sediments (Yule and Tomlin, 1971). In streams, much of the residue is in continuous transport on suspended particulate material or in sediments (Zabik, 1969). The distribution within the stream flow is non-uniform because of unequal flow velocity and distribution of suspended materials within the stream bed (Feltz, 1971). Seasonal fluctuations in run-off and use patterns cause major changes in concentration during the year, but the continuous downstream transport tends to reduce levels in the upper reaches of streams while increases may be observed in the downstream areas and eventually in major receiving basins (lake, reservoir, estuary, and ocean). If applications in a watershed cease entirely, residues in the stream gradually and continuously decline (Sprague, et al., 1971), and a similar decline would be expected in the receiving basins but at a slower rate and a later time.

In lakes, sediments apparently act as a reservoir from which the pesticide is partitioned into the water phase according to the solubility of the compounds, the concentration in the sediment, the type of sediment, and the degree of absorption (Hamelink, et al., 1971). Many herbicides applied to aquatic systems to control aquatic plant growth pass from water to organic sediments where they may persist for long periods, although water concentrations remain low (Frank and Comes, 1967). Dissolved natural organic materials in the water



may greatly enhance the amounts of some pesticides carried in water (Wershaw, et al., 1969). Some investigations indicate pesticides may be less available to the water in highly fertile systems where the higher organic content in the sediments has a greater capacity to bind pesticide residues. This may in part explain the difference in times required for different waters to "detoxify" as observed in lakes treated with toxaphene to eradicate undesirable fish species (Terriere, 1966).

### Kinds of Effects

Pesticides may be harmful because they eliminate or reduce populations of desirable organisms directly, or because they indirectly alter conditions required by these organisms. Direct effects include mortality, birth defects, induction of tumors and genetic changes, altered behavior patterns, or physiological changes including alterations in reproduction; the effects may take place during any stage of the organism's life history. Indirect effects may include reduction of species used as food sources by other species; reduction in rates of photosynthesis, decomposition or mineralization, with attendant unsatisfactory conditions for certain species; and temporary increased BOD (biological oxygen demand) in a body of water resulting from decomposition following death of plankton or other organisms caused by a pesticide. Additionally, many species can accumulate pesticide residues directly from the water, or from sediments, and these residues may in turn be accumulated in organisms that feed on the lower forms of life. This latter phenomenon has resulted in pesticide residues entering human food supplies and in effects on other mammals and fish-eating birds.

Except in the case of man and domestic animals, effects on individuals are less important than effects on populations, communities of organisms, and whole ecosystems. Most organisms in the wild have short life spans, and the turnover rates of the populations are very high. Furthermore, because most species have high reproductive rates, most individuals in a population are biologically excess to the continued existence of the population and unless the fraction of the population killed is high, the level of the population will be only temporarily suppressed. Birth defects, tumors, and even genetic changes may be relatively unimportant to survival in wild populations with rapid turnover rates because individuals rendered unfit will be eliminated without detriment to the continuation of the population; i.e., mortality is transferred to the unfit. Where turnover rates are low and life spans relatively long as in osprey, eagle, brown pelican and certain other species, birth defects, genetic changes, and tumors may be significant.

### Lethality

Concentrations of pesticides that are lethal to aquatic life have occurred in local areas where applications overlap streams or lakes, in streams receiving runoff from recently treated areas, and where misuse, spillage, or improper waste disposal have occurred. Applications of pesticides to water to control noxious plants, fish, or insects have also killed desirable species. Past experience with local fish mortalities from pesticide contamination has shown that some fish populations recover within a few months to a year after pesticide contamination is stopped (Elson, 1967). The recovery of aquatic invertebrates in areas that have been heavily contaminated may require a longer period with

some species requiring several years to regain precontamination numbers (Cope, 1961; Ide, 1967). Recovery in Arctic areas also is very slow (Reed, 1966). Less desirable species of insects may be the first to repopulate the area (Hynes, 1961) and in some instances the species composition has been completely changed (Hopkins, 1966). Areas that are contaminated by pesticide application are subject to loss of fish populations and/or reduced food available for fish growth (Schoenthal, 1963; Kerswill, et al., 1967). Where residues are persistent in bottom sediments for long periods, benthic organisms may be damaged even though water concentrations remain low (Wilson and Bond, 1969).

Great differences in susceptibility exist among species and within species for different compounds. As an example, Pickering, et al. (1962) reported 96-hour LC<sub>50</sub> values of 5 to 610,000 ug/l (ppb) for various fish species exposed to organophosphate pesticides. In addition to species differences, the toxicity may be modified by differences in formulation, environmental conditions such as temperature and water hardness, animal size and age, previous exposure, and physiological condition. The effects of combinations of pesticides on aquatic organisms are not well understood. Macek (unpublished) reported that some combinations of various common pesticides were synergistic in their action on bluegill and rainbow trout, while others had only additive effects.

Most data on the effects of a given pesticide on aquatic life are limited to concentrations that are lethal in short-term tests and for only a few species. The relatively few chronic tests conducted with aquatic species indicate that effects usually occur at concentrations much lower than lethal concentration levels, and that continued exposure to relatively low concentrations may often result in detectable effects. Mount and Stephan (1967) found the 96-hour LC<sub>50</sub> for fathead minnows to malathion was 9000 ug/l, but spinal deformities in adult fish occurred during a 10-month exposure to 580 ug/l. Eaton (1970) found that bluegills with a 96-hour LC<sub>50</sub> of 108 ug/l suffered the same spinal deformities as the fathead minnows after chronic exposure to only 7.4 ug/l malathion.

Mount and Stephan (1967) have raised the possibility of estimating no detectable effect levels for species of fish for which such levels have not been experimentally determined through the use of "application factor" values. The "application factor" for a pesticide or other chemical is the ratio obtained by dividing laboratory determined maximum concentrations of the pesticide or other chemical that have no detectable chronic exposure effect in a species of fish by the 96-hour LC<sub>50</sub> for that species. The hypothesis is that the ratio or "application factor" does not vary substantially for the same compound among species of fish, whereas both sensitivity of a fish species to different toxicants and sensitivity to the same toxicant by different species vary widely. The experimental data to support the hypothesis are as yet few, and for pesticides even fewer. Table 1 presents available data on application factors for pesticides and, in addition, presents data on other substances where more than a single species of fish has been tested.

Where data for more than a single species or a compound exist, the greatest disparity between the high and low application factor is a factor of 10 for Chromium<sup>+6</sup>. Copper is next with a factor of 7. For all others, the difference is 3 or less. On the other hand, the application factors for different compounds for the same species differ as much as 1000-fold (fathead minnows in relation to diazinon at 0.0005 and lindane at 0.5).

Table 1. "Application Factors" Determined Experimentally.

Chemical	Fish Species	96- Hour LC <sub>50</sub> (mg/l)	Application Factor <u>c/</u>
<u>More than 1 species tested</u>			
Malathion	Fathead Minnow	10.5	.02
	Bluegill	.08	.04
	Brook Trout	.2	.02
Lindane	Fathead Minnow	50	.5
	Brook Trout	26	.38
Chromium <sup>+6</sup>	Fathead Minnow	33	.03
	Brook Trout	50	.01
	Rainbow Trout	69	.003
Copper	Fathead Minnow <u>a/</u>	.47	.03
	Fathead Minnow <u>b/</u>	.075	.14
	Bluegill	1.1	.02
	Brook Trout	.1	.09
Cadmium	Fathead Minnow	31	.001
	Bluegill	20	.0015
	Green Sunfish	20	.0025
Methyl Mercury	Fathead Minnow	.04	.006
	Brook Trout	.096	.003
Lead	Brook Trout	4.5	.013
	Rainbow Trout	.14 (18 day)	.043
<u>Pesticides where only a single species has been tested</u>			
Diazinon	Fathead Minnow	6	.0005
Captan	Fathead Minnow	.065	.10
2,4-D Butoxy- ethanol ester	Fathead Minnow	5.6	.05
Carbaryl	Fathead Minnow	9	.023
Methoxychlor	Fathead Minnow	.0075	.017

a/ Hardwater.b/ Softwater.

c/ The "application factor" is the ratio obtained by dividing experimentally determined maximum concentrations of a pesticide that have no detectable effect during chronic exposures by the 96-hour LC<sub>50</sub> for that species.

Thus, further investigation may provide additional support for estimating no detectable chronic effect levels when only acute toxicity data for the fish species of interest is available, but an application factor has been determined experimentally for some other fish species for the pesticide of interest.

Some reported acute toxicity values and sub-acute effects of pesticides for freshwater aquatic life are listed in Appendix Table 2 and for estuarine and marine life in Appendix Table 3.

Work done in this area for invertebrates has shown acute and chronic toxicity levels to be much closer together than those for fish. As a result of this phenomenon, the possibility of determining application factors for use in establishing no detectable effect levels for invertebrates is not promising.

Even when concentration levels which are acutely toxic and those which produce no detectable effect are both known, their use in the regulation of pesticides is complex. Maximum "acceptable" levels for particular bodies of water or portions thereof must reflect full consideration of both the benefits of pesticide use and the environmental costs of such use. The two levels discussed thus far may be considered as merely two points among the full range of dose and effect; the entire range may be of interest as regulatory decisions are made. Concentrations equal to or greater than the 96-hour LC<sub>50</sub> may well be "acceptable" depending on the area affected, the time involved, the importance of pesticide use, water use classification, and other factors.

#### Persistence and Biological Accumulation

All organic pesticides are subject to metabolic and non-metabolic degradation in the environment. Different compounds vary tremendously in their rate of degradation and some form degradation products which may be both persistent and toxic. Many pesticides are readily degraded to non-toxic or elemental materials within a few days to a few weeks. These "non-persistent" compounds may be absorbed by aquatic organisms, and may affect the organism, but the residues do not necessarily accumulate or persist for long periods. Concentrations in the organism may be higher than ambient water levels, but sublethal amounts decline rapidly as water concentrations decrease. Examples of such dynamic exchange have been demonstrated with malathion (Bender, 1969), methoxychlor (Burdick, 1968), various herbicides (Mullison, 1970, and others). If degradation in water is sufficiently rapid that adverse physiological effects do not occur, these non-persistent compounds do not pose a long-term hazard to aquatic life. Degradation rates are often a function of environmental conditions, however, and great variation may be observed. The organophosphate insecticides, for example, are rapidly hydrolysed in alkaline waters and at higher temperatures, whereas at lower pH and temperature they may persist for several months (Gakstatter and Weiss, 1965). Repeated applications and slow degradation rates may maintain elevated environmental concentrations and hence bring about undesired changes in the biota, but there is no indication that these compounds can be accumulated through the food chain.

Effects of pesticides may be persistent or even cumulative, even though the pesticide may not be persistent or continuously present in the environment. Thus fish exposed to sublethal levels of malathion have shown depressed acetylcholinesterase levels which are slow to recover. Subsequent sublethal exposures

have resulted in additional cholinesterase depression sufficient to result in death (Coppage and Duke, in press).

Some pesticides, primarily the organochlorine and metallic compounds, are extremely stable, degrading only slowly or forming persistent degradation products. Residues may be detectable for weeks, months or years. Aquatic organisms may accumulate these compounds directly from water and from contaminated food. Some fish and some other aquatic organisms accumulate organochlorine compounds from remarkably low levels. Thus it has been shown experimentally that shrimp and some fish can take up polychlorinated biphenyls (PCB's) from concentrations of less than 1/10 part per billion (ppb) in water, and accumulate them by factors of as much as 75,000. (Nimmo, et al., 1971; Stallings and Mayer, 1972). In other cases, it appears that uptake is by algae and invertebrates with residues in fish resulting from feeding on contaminated foods. Either process, or a combination of the processes ultimately results in residues in the higher feeding levels that may be many thousand times higher than ambient water levels.

Food chain accumulation does not stop at the water's edge. In fact, fish-eating birds often contain the highest residue levels of DDT and its breakdown products (DDE and DDD) in food chains studied. DDT (including its breakdown products) is the best understood of the persistent pesticides. It is widely distributed in freshwater and marine environments in North America and throughout the world. Its most abundant breakdown product is DDE.

The discovery and clear demonstration of DDE-induced thinning of shells of the eggs of wild birds constituted a major research breakthrough of the late 1960's. In 1967, Derek Ratcliffe reported a synchronous, rapid, and widespread decline in weight and thickness of shells of eggs laid by British peregrine falcons and sparrowhawks that occurred in the mid-forties (Ratcliffe, 1967). Eggshell thinning proved not to be confined to Great Britain, for in 1968 Hickey and Anderson reported eggshell thinning of 18-26 percent in regional populations of three species of raptorial birds that had declined markedly in the United States (Hickey and Anderson, 1968). The period of decline coincided with the same occurrence in Great Britain and persisted through succeeding years. Museum studies were extended to include more than 23,000 eggs of 25 species (Anderson and Hickey, 1970). Some degree of shell thinning was found among 22 species representing seven Orders of birds. Nine of the species sustained shell thinning of 20 or more percent. Other workers have subsequently extended the list.

These findings produced the hypothesis of DDT involvement. The hypothesis was tested experimentally. The first clearcut experimental demonstration that DDE caused thin eggshells was provided in studies of mallard ducks (Heath, et al., 1969). In subsequent controlled experiments, dietary dosages of approximately 3 ppm wet weight of DDE thinned the shells of eggs laid by kestrels (Wiemeyer and Porter, 1970), black ducks (Longcore, et al., 1971), and screech owls (McLane and Hall, 1972), extending the experimental demonstration to four species of three Orders of birds. Quail and chickens, seed-eating galliform birds, were, at the most, only slightly susceptible (Smith, et al., 1970; Stickel and Rhodes, 1970; Cecil et al., 1971, 1972).

The third stage of investigation was to return again to the field and study the relationships between thinning of shells and residues in the eggs. The brown pelican provided the ideal test because of contrasting residues and reproductive success in different localities. The California colony was essentially failing, the Carolina population declining, and the Florida colonies remaining reasonably stable. Measurements of shell thickness and residues of dieldrin, PCB's, mercury, lead, DDT, DDD, and DDE were subjected to computerized statistical tests, which implicated DDE as the shell thinner (Blus, 1970; Blus, et al., 1971, 1972).

### Residues

Samples of fish have often contained pesticide residues in concentrations that give rise to concern. The highest concentrations are often in those species most highly prized as food or game species inasmuch as these species are usually at the top of a relatively long food chain. Sales in interstate commerce of coho salmon and several other species from Lake Michigan and of canned Jack mackerel in California, were prohibited in 1967 on the basis of DDT residues in excess of the 5 ppm interim guideline for DDT and its metabolites set for fish by the U.S. Food and Drug Administration. Pesticide residues in fish or fish products may enter the human food chain less directly; for example, through fish oil and meal used in domestic animal feeds, which result in turn in residues in meat and other animal products.

Fish may survive relatively high residue concentrations in their body fats, but residues concentrated in the eggs of mature fish may be lethal to the developing fry. Burdick (1964) reported up to 100 percent loss of lake trout fry when residues of DDT-DDD in the eggs exceeded 4.75 mg/kg. (ppm). A similar mortality was reported in coho salmon fry from Lake Michigan when eggs contained significant quantities of DDT, dieldrin, and polychlorinated biphenyls (Johnson and Pecor, 1969; Johnson, unpublished). Johnson (1967) reported that adult fish which did not appear to be harmed by low concentrations of endrin in water, accumulated endrin levels in the eggs that were lethal to the developing fry.

Residues in fish may be directly harmful under stress conditions or at different temperature regimes. Brook trout fed DDT at 3.0 mg/kg. body weight per week for 26 weeks suffered 96.2 percent mortality during a later period of reduced feeding on clean food and declining water temperature; mortality of untreated control fish during the same period was 1.2 percent (Macek, 1968). Declining water temperature during the fall was believed to cause delayed mortality of salmon parr in streams contaminated with DDT (Elson, 1967).

Certain organochlorine pesticides (DDT, TDE, aldrin, dieldrin, endrin, chlordane, heptachlor, mirex, toxaphene, lindane, endosulfan and benzene hexachloride) are considered especially hazardous to aquatic life because of their accumulation in aquatic organisms. Some of these compounds, including some of their metabolites, are toxic to various aquatic species at concentrations of less than one ug/l, (See Appendix Tables 1 and 2). Their accumulation in aquatic systems presents a hazard, both real and potential, to animals in the higher part of the food chain, including man (Pimentel, 1971, Mrak, 1969; Kraybill, 1969; and Gillett, 1969).

On the basis of present knowledge, we cannot estimate with certainty levels of persistent pesticides in water that will not result in undesired effects. Water concentrations below the practical limits of detection have resulted in residues in fish sufficiently high to be unacceptable for human consumption, to prevent normal reproduction in some fish-eating bird species, and to affect reproduction and survival of aquatic life. In these circumstances, criteria may be based upon specific residue concentrations in the tissues of selected species. The ratio of residue concentrations in tissue to concentrations in the water can be determined experimentally. This accumulation factor might then be applied to acceptable tissue concentration levels to estimate acceptable water concentration levels.

Table 2 lists some "accumulation factors" determined experimentally. These accumulation factors represent direct uptake from water; they do not include food chain accumulations which often may be a factor of up to 8 to 10 from one feeding level to another (Buckley, 1969; Woodwell, et al., 1967). The combined accumulation factor of direct uptake from water and a single level of food chain accumulation could exceed 100,000 for DDT in certain fish and could be less than 500 for lindane in mussels. This is not to say that food chain accumulation and direct uptake from water are necessarily additive. The residue level existing at any time is a dynamic balance between intake and elimination, and the contribution to intake directly from water or from food will vary according to concentration in and assimilation rates from the two sources.

There are, of course, a number of difficulties in applying this system. The desirable levels in water will sometimes be below the practical limits of detection; accumulation factors will not be known with precision, especially where both direct uptake from the environment and food chain accumulation with several feeding levels are involved. Nonetheless, based on estimates of residue levels that will not adversely affect man or valuable organisms at the top of food chains, and estimates of accumulation factors (from both direct uptake and food chain transfers), corresponding levels in water can be computed.

Table 2 BIOLOGICAL ACCUMULATION OF PESTICIDE CHEMICALS

ORGANISM	CHEMICAL	EXPOSURE CONCENTRATION	CONCENTRATION FACTOR	TIME	SPECIAL DETAILS	REFERENCE
BACTERIA						
<u>Pseudomonas</u> spp.	Nonachlor	10 ppm	0.57	10 days	Mixed culture of four species	Bourquin, 1972
	Chlordane	10 ppm	0.83	↓		
	Heptachlor	10 ppm	0.1	↓		
CILIATES						
<u>Tetrahymena pyriformis</u> W	Mirex	0.9 ppb	193	1 week	Axenic cultures incubated at 26°C; concentration factor on dry weight basis	Cooley, et al., 1971
	Aroclor 1248	10 ppb	40	↓		Cooley and Keltner, 1971
	Aroclor 1254	1 ppm	60	↓		Cooley, et al., 1971
	Aroclor 1260	1 ppm	79	↓		Cooley and Keltner, 1971
MOLLUSCS						
Hooked mussel <u>Brachidontes recurvus</u>	DDT	1 ppb	24,000	1 week	Whole body residues (Meats)	Butler, 1966
Hard-shell clam <u>Mercenaria mercenaria</u>	DDT	0.1 ppb	1,260	5 days		Butler, 1971
		1 ppb	6,000	1 week		Butler, 1966
	Aldrin	0.5 ppb	380	5 days		Butler, 1971



ORGANISM	CHEMICAL	EXPOSURE CONCENTRATION	CONCENTRATION FACTOR	TIME	SPECIAL DETAILS	REFERENCE
MOLLUSCS (continued)						
<u>M. mercenaria</u>	Dieldrin	0.5 ppb	760	5 days	Whole body residues (Meats) ↓	Butler, 1971
	Endrin	0.5 ppb	480	5 days		Butler, 1971
	Heptachlor	0.5 ppb	220	5 days		Butler, 1971
	Lindane	5.0 ppb	12	5 days		Butler, 1971
	Methoxychlor	1.0 ppb	470	5 days		Butler, 1971
Soft-shell clam						
<u>Mya arenaria</u>	Aldrin	0.5 ppb	4,600	5 days		Butler, 1971
	DDT	0.1 ppb	8,800	5 days		Butler, 1971
	Dieldrin	0.5 ppb	1,740	5 days		Butler, 1971
	Endrin	0.5 ppb	1,240	5 days		Butler, 1971
	Heptachlor	0.5 ppb	2,600	5 days		Butler, 1971
	Lindane	5.0 ppb	40	5 days		Butler, 1971
	Methoxychlor	1.0 ppb	1,500	5 days		Butler, 1971
Pacific oyster						
<u>Crassostrea gigas</u>	DDT	1.0 ppb	20,000	7 days		Butler, 1966
European oyster						
<u>Ostrea edulis</u>	DDT	1.0 ppb	15,000	7 days		Butler, 1966
Crested oyster						
<u>O. equestris</u>	DDT	1.0 ppb	23,000	7 days		Butler, 1966

ORGANISM	CHEMICAL	EXPOSURE CONCENTRATION		CONCENTRATION FACTOR	TIME	SPECIAL DETAILS	REFERENCE
MOLLUSCS (continued) Eastern oyster <u>Crassostrea virginica</u>	DDT	10	ppb	15,000	7 to 15 days	Whole body residues (Meats) ↓	Butler, 1967
		1	ppb	30,000			Butler, 1967
		0.1	ppb	70,000			Butler, 1967
		0.01	ppb	70,000			Butler, 1967
		0.0001	ppb	0			Butler, 1967
	Aroclor <sup>®</sup> 1254	0.01	ppb	50,000	8 weeks		Parrish, 1972
		1	ppb	76,000	24 weeks		Lowe, et al., 1970
		0.01	ppb	160,000	8 weeks		Parrish, 1972
		1	ppb	101,000	30 weeks		Parrish, et al., 1972
CRUSTACEAN Grass shrimp <u>Palaemonetes pugio</u>	Dieldrin	0.01	ppb	8,000	8 weeks		Parrish, 1972
	Aroclor <sup>®</sup> 1254	0.62	ppb	2,069	1 week	Whole body residues (Meats) ↓	Nimmo and Heitmuller, 1972
				11,920	2 weeks		Nimmo and Heitmuller, 1972
				10,903	3 weeks		Nimmo and Heitmuller, 1972
				17,425	4 weeks		Nimmo and Heitmuller, 1972
				26,580	5 weeks		Nimmo and Heitmuller, 1972

ORGANISM	CHEMICAL	EXPOSURE CONCENTRATION	CONCENTRATION FACTOR	TIME	SPECIAL DETAILS	REFERENCE
CRUSTACEAN (continued)						
<u>P. pugio</u>	Aroclor 1254 <sup>®</sup>	0.09 ppb	3,611	1 week	Whole body residues	Nimmo and Heitmuller, 1972
			4,800	2 weeks		Nimmo and Heitmuller, 1972
			5,000	3 weeks		Nimmo and Heitmuller, 1972
			17,400	4 weeks		Nimmo and Heitmuller, 1972
			8,355	5 weeks		Nimmo and Heitmuller, 1972
		0.037 ppb	1,594	1 week		Nimmo and Heitmuller, 1972
			3,405	2 weeks		Nimmo and Heitmuller, 1972
			3,918	3 weeks		Nimmo and Heitmuller, 1972
			4,567	4 weeks		Nimmo and Heitmuller, 1972
			5,729	5 weeks		Nimmo and Heitmuller, 1972
Pink shrimp <u>Penaeus duorarum</u>	Mirex	0.1 ppb	2,600	3 weeks	Whole body residues	Lowe, et al., 1971
			24,000	3 weeks	Hepatopancrease	Lowe, et al., 1971
	DDT	0.14 ppb	1,500	3 weeks	Whole body residues	Nimmo, et al., 1970
	Aroclor 1254 <sup>®</sup>	2.5 ppb	1,800	2 days	Whole body residues	Nimmo, et al., 1971
			2,760	4 days		Nimmo, et al., 1971
			6,800	6 days		Nimmo, et al., 1971
			7,600	9 days		Nimmo, et al., 1971

ORGANISM	CHEMICAL	EXPOSURE CONCENTRATION	CONCENTRATION FACTOR	TIME	SPECIAL DETAILS	REFERENCE
CRUSTACEAN (continued)						
<u>P. duorarum</u>	Aroclor <sup>®</sup> 1254	2.5 ppb	9,600	12 days	Whole body residues	Nimmo, et al., 1971
			15,600	15 days	↓	Nimmo, et al., 1971
			12,400	22 days	↓	Nimmo, et al., 1971
Mud crab (larvae)						
<u>Rhithropanopeus harrisi</u>	Mirex	0.1 ppb	1,000	7 weeks	Static culture bowl method with a change to fresh medium + chemical each day	Bookhout, et al., 1972
	Malathion	10 ppb	0 (larvae) 0 (adults)	4 weeks		Tyler, 1971
Blue crab (juveniles)						
<u>Callinectes sapidus</u>	Mirex	0.1 ppb	1,100 - 5,200	3 weeks	Whole body residues	Lowe
FISH						
Pinfish						
<u>Lagodon rhomboides</u>	DDT	0.1, 1.0 ppb	10,600 - 38,000	2 weeks	Whole body residues	Hansen and Wilson, 1970
	Aroclor <sup>®</sup> 1254	5 ppb	2,800 - 21,800	2 - 15 weeks	↓	Hansen, et al., 1971
Spot						
<u>Leiostomus xanthurus</u>	Aroclor <sup>®</sup> 1254	1 ppb	17,000 - 27,000	4 - 8 weeks	Whole body residues	Hansen, et al., 1971
		5 ppb	9,200 - 30,400	3 - 6 weeks	↓	Hansen, et al., 1971
Atlantic croaker						
<u>Micropogon undulatus</u>	DDT	0.1, 1.0 ppb	10,000 - 14,000	3 weeks	Whole body residues	Hansen and Wilson, 1970

ORGANISM	CHEMICAL	EXPOSURE CONCENTRATION	CONCENTRATION FACTOR	TIME	SPECIAL DETAILS	REFERENCE	
VASCULAR PLANTS							
Turtle grass							
<u>Thalassia testudinum</u>	Tordon 101 (39.6% 2,4-D; 14.3% Picolinic acid)	1 ppm	leaves 0 (2,4-D) 0 (Picolinic acid)	10 days	Plants exposed to chemical through rhizomes; concentration factor on wet weight basis	Walsh and Hollister, 1971	
			rhizomes 0.05 (2,4-D) 0 (Picolinic acid)		↓		
		5 ppm	leaves 0 (2,4-D) 0 (Picolinic acid)				
			rhizomes 0.12 (2,4-D) 0.02 (Picolinic acid)				
	Aroclor <sup>®</sup> 1254	5,820 ppb	0 leaves 0 rhizomes	10 days			Walsh and Hollister, 1971
	Mirex	0.1 ppb	0 leaves 0.36 rhizomes	10 days			Walsh and Hollister, 1971

ORGANISM	CHEMICAL	EXPOSURE CONCENTRATION	CONCENTRATION FACTOR	TIME	SPECIAL DETAILS	REFERENCE
VASCULAR PLANTS (continued)						
Red mangrove						
<u>Rhizophora mangle</u>	Tordon 101	14.4 ppb	roots	20 days	Seedlings treated when two pairs of leaves were present; concentration factor on wet weight basis	Walsh, et al., 1972
	(39.6% 2,4-D;		1.28 (2,4-D)			
	14.3% Picolinic acid)		0.64 (Picolinic acid)			
			hypocotyl			
			0.64 (2,4-D)			
			2.1 (Picolinic acid)			
			stems			
			1.28 (2,4-D)			
			0.64 (Picolinic acid)			
			1st leaves		↓	
			1.28 (2,4-D)			
			0.63 (Picolinic acid)			
			2nd leaves		↓	
			9.0 (2,4-D)			
			4.2 (Picolinic acid)		↓	

ORGANISM	CHEMICAL	EXPOSURE CONCENTRATION	CONCENTRATION FACTOR	TIME	SPECIAL DETAILS	REFERENCE
VASCULAR PLANTS (continued)						
Red mangrove						
<u>Rhizophora mangle</u>	Tordon 101	14.4 ppb		40 days	Seedlings treated when two pairs of leaves were present; concentration factor on wet weight basis	Walsh, et al., 1972
	(39.6% 2,4-D;		1.28	roots		
	14.3% Picolinic acid)		0.64	(2,4-D)		
				(Picolinic acid)		
				hypocotyl		
			16.0	(2,4-D)		
			6.0	(Picolinic acid)		
				stems		
			16.0	(2,4-D)		
			6.0	(Picolinic acid)		
				1st leaves		
			20.0	(2,4-D)		
			6.0	(Picolinic acid)		
				2nd leaves		
			24.3	(2,4-D)		
			6.0	(Picolinic acid)		

ORGANISM	CHEMICAL	EXPOSURE CONCENTRATION	CONCENTRATION FACTOR	TIME	SPECIAL DETAILS	REFERENCE
VASCULAR PLANTS (continued)						
Red mangrove						
<u>Rhizophora mangle</u>	Tordon 101 (39.6% 2,4-D; 14.3% Picolinic acid)	144 ppb	roots 10.8 (2,4-D) 2.9 (Picolinic acid)	10 days	Seedlings treated when two pairs of leaves were present; concentration factor on wet weight basis	Walsh, et al., 1972
			hypocotyl 14.7 (2,4-D) 4.3 (Picolinic acid)			
			stems 9.0 (2,4-D) 3.8 (Picolinic acid)			
			1st leaves 5.5 (2,4-D) 2.1 (Picolinic acid)			
			2nd leaves 7.7 (2,4-D) 3.6 (Picolinic acid)			



## ACKNOWLEDGEMENT

Appendix Tables 2 and 3 are based on data assembled by the National Academy of Sciences - National Academy of Engineering Committee on Water Quality Criteria.

ABBREVIATIONS USED

LC<sub>50</sub> median lethal concentration: the concentration of toxicant in the  
or environment which kills 50 percent of the organisms exposed to it.  
LC-50 Usually duration of exposure is specified, e.g., 96-hour LC<sub>50</sub>.

ppb parts per billion

ppm parts per million

mg/kg milligrams per kilogram = parts per million

mg/l milligrams per liter = parts per million

TLm median tolerance limit: the concentration of a test material in  
experimental water at which just 50 percent of the test animals are  
able to survive for a specified period of exposure, e.g., 96 hours.

ug/g Micrograms per gram - parts per million

ug/kg micrograms per kilogram - parts per billion

ug/l micrograms per liter - parts per billion

LIST OF PESTICIDES MENTIONED IN REPORT

ABATE

Chemical name: 0,0,0',0'-tetramethyl 0,0'-thiodi-  
p-phenylene phosphorothioate  
Other name: Biothion  
Action: Insecticide

ALDRIN

Chemical name: 1,2,3,4,10,10-hexachloro-1,4,4a,5,8,  
8a-hexahydro-1,4-endo-exo-5,8-dimethanonaphthalene

AMETRYNE

Chemical name: 2-(ethylamino)-4-(isopropylamino)-  
6-(methylthio)-s-triazine  
Other name: Gesapax  
Action: Herbicide

AMIBEN

Chemical name: 3-amino-2,5-dichlorobenzoic acid  
Other name: Chloramben  
Action: Herbicide

ALTRAZINE

Chemical name: 2-chloro-4-ethylamino-6-isopropylamino-  
s-triazine  
Other names: Aatrex, Fenamine, Fenatrol, Gesaprim, Primatol A  
Action: Herbicide

AZINPHOS-METHYL

Chemical name: 0,0-dimethyl S-[4-oxo-1,2,3-benzotriazin-  
3(4H)-ylmethyl] phosphorodithioate  
Other names: Carfene, DBD, Gusathion, Gusathion M,  
Gustathion, Guthion, Methyl Guthion  
Action: Insecticide

BAYGON

See PROPOXUR

BAYTEX

See FENTHION

BENZOIC ACID

Action: Fungicide

BHC

See LINDANE

CARBARYL

Chemical name: 1-naphthyl methylcarbamate  
Other name: Sevin  
Action: Insecticide

CHLORAMBEN

See AMIBEN

CHLORDANE

Chemical name: 1,2,4,5,6,7,8,8-octachloro-2,3,3a,4,7,  
7a-hexahydro-4,7-methanoindene  
Other names: Chlordan, Chlor Kil, Corodane, Kypchlor,  
Octachlor, Octa-Klor, Ortho-Klor, Synklor, Topiclor 20,  
Velsicol 1068  
Action: Insecticide

CO-RAL

See COUMAPHOS

COUMAPHOS

Chemical name: 0,0-diethyl 0-[3-chloro-4-methyl-2-oxo-(2H)-benzopyran-7-yl] phosphorothioate

Other names: Agridip, Asuntol, Co-Ral, Muscatox, Resistox

Action: Insecticide

2,4-D

Chemical name: 2,4-dichlorophenoxyacetic acid or its sodium salt or amine

Other names: Chloroxone, Crop Rider, Ded-Weed, Weed-Ag-Bar, Weedar 64, Weed-B-Gon, Weedone

Action: Herbicide

DALAPON

Chemical name: 2,2-dichloropropionic acid

Other names: Ded-Weed, Dowpon, Gramevin, Radapon, Unipon

Action: Herbicide

DDD

See TDE

DDT

Chemical name: dichloro diphenyl trichloroethane

Other names: Anofex, Chlorophenothane, Dedelo, Genitox, Gesapon, Gesarex, Gesarol, Gyron, Ixodex, Kopsol, Neocid, Pentachlorin, Rukseam, Zerdane

Action: Insecticide

DDVP

See DICHLORVOS

DELNAV

See DIOXATHION

DIAZINON

Chemical name: 0,0-diethyl 0-(2-isopropyl-6-methyl-4-pyrimidinyl) phosphorothioate

Other names: Basudin, Dazzel, Diazajet, Diazide, Gardentox, Spectracide

Action: Insecticide

DICAPTHON

Chemical name: 0-(2-chloro-4-nitrophenyl) 0,0-dimethyl phosphorothioate

Other name: Di-Captan

Action: Insecticide

DICHLOBENIL

Chemical name: 2,6-dichlorobenzonitrile

Other names: Casoron, Du-Sprex, 2,6-DBN

Action: Herbicide

DICHLORVOS

Chemical name: 2,2-dichlorovinyl 0,0-dimethyl phosphate

Other names: DDVF, DDVP, Dedevap, Dichlorphos, Herkol, Mafu, Marvex, Nogos, No-Pest, Nuvan, Oko, Phosvit, Vapona

Action: Insecticide

DIELDRIN

Chemical name: 1,2,3,4,10,10-hexachloro-exo-6,7-epoxy-1,4,4a,5,6,7,8,8a-octahydro-1,4-endo-exo-5,8-dimethanonaphthalene, and related compounds  
Other names: HEOD, Octalox, Panoram D-31  
Action: Insecticide

DIOXATHION

Chemical name: 2,3-p-dioxanedithiol S,S-bis-(0,0-diethyl phosphorodithioate)  
Other names: Delnav, Navadel, Ruphos  
Action: Insecticide

DIPHTHEREX

See TRICHLORFON

DIQUAT

Chemical name: 1,1'-ethylene-2,2'-dipyridylum dibromide  
Other names: Aquacide, Dextrone, FB/2, Reglone  
Action: Herbicide

DISULFOTON

Chemical name: 0,0-diethyl S-2-(ethylthio)ethyl phosphorodithioate  
Other names: Diethylethylthioethyl dithiophosphate, Di-syston, Dithiodemeton, Dithiosystox, Frumin A1, Frumin G, Solvirex, Thiodemeton  
Action: Insecticide

DI-SYSTON

See DISULFOTON

DIURON

Chemical name: 3-(3,4-dichlorophenyl)-1,1-dimethylurea  
Other names: DCMU, DMU, Karmex, Marmer  
Action: Herbicide

DURSBAN

Chemical name: 0,0-diethyl 0-3,5,6-trichloro-2-pyridyl phosphorothioate  
Action: Insecticide

ENDOSULFAN

Chemical name: 6,7,8,9,10,10-hexachloro-1,5,5a,-6,9,9a-hexahydro-6,9-methano-2,4,3-benzodioxathiepin 3-oxide  
Other names: Chlorthiepin, Cyclodan, Insectophene, Kop-Thiodan, Malic, Malix, Thifor, Thimul, Thiodan  
Action: Insecticide

ENDOTHALL

Chemical name: 7-oxabicyclo (2,2,1) heptane-2,3-dicarboxylic acid  
Other names: Accelerate, Aquathol, Des-i-cate, Endothal, Hydrothol, Niagrathal, Tri-Endothal  
Action: Herbicide

ENDRIN

Chemical name: 1,2,3,4,10,10-hexachloro-6,7-epoxy-1,4,4a,5,6,7,8,8a-octahydro-1,4-endo-endo-5,8-dimethanonaphthalene  
Other names: Hexadrin, Mendrin  
Action: Insecticide

**FENTHION**

Chemical name: 0,0-dimethyl 0[4-(methylthio)-m-tolyl]  
phosphorothioate

Other names: Baytex, DMPT, Entex, Lebaycid, Mercaptophos,  
Quelatox, Queletox, Tiguvon

Action: Insecticide

**FENURON**

Chemical name: 3-phenyl-1,1-dimethylurea

Other names: Dybar, Fenidim, Fenulon, PDU

Action: Herbicide

**GUTHION**

See AZINPHOS-METHYL

**HEPTACHLOR**

Chemical name: 1,4,5,6,7,8,8-heptachloro-3a,4,7,-  
7a-tetrahydro-4,7-methanoindane

Other names: Drinox H-34, Heptamul

Action: Insecticide

**LEAD ARSENATE**

Other names: Gypsine, Soprabel

Action: Insecticide

**LINDANE**

Chemical name: gamma isomer of 1,2,3,4,5,6-hexachloro-  
cyclohexane; also known as gamma benzene hexachloride

Other names: Gamaphex, Gamma BHC, Gammaline, Gammex,  
Gammexane, Isotox, Lindafor, Lindagam, Lintox, Novigam,  
Silvanol, Tri-6-Dust

Action: Insecticide

**MALATHION**

Chemical name: 0,0-dimethyl S-(1,2-dicarbethoxyethyl)  
dithiophosphate

Other names: Carbofos, Carbophos, Cythion, Emmatos,  
Karbofos, Kop-Thion, Kypfos, Malamar, Malaspray,  
Malathon, Mercaptothion, Zithiol

Action: Insecticide

**MCPA**

Chemical name: 4-chloro-2-methylphenoxyacetic acid

Other names: Agroxone, Chiptox, Hormotuh, Kilsem,  
MCP, Mephanac, Metaxon, Methoxone, Rhomene, Rhonox

Action: Herbicide

**MERCURY**

Action: Fungicide

**METHOXYCHLOR**

Chemical name: 1,1,1-trichloro-2,2-bis(p-methox-  
yphenyl)ethane

Other names: Dianisyltrichloroethane, Dimethoxy-DT,  
DMDT, Marlate, Methoxy DDT

Action: Insecticide

**METHYL PARATHION**

Chemical name: 0,0-dimethyl 0-p-nitrophenyl phosphorothioate

MEVINPHOS

Chemical name: 2-methoxycarbonyl-1-methyl-vinyl  
dimethyl-phosphate  
Other names: Phosdrin, Phosfene  
Action: Insecticide

MIREX

Chemical name: dodecachlorooctahydro-1,3,3-metheno-  
2H-cyclobuta(cd)pentalene  
Other name: Dechlorane  
Action: Insecticide

MONURON

Chemical name: 3-(p-chlorophenyl)-1,1-dimethylurea  
Other names: Chlorfenidim, Telvar  
Action: Herbicide

NEBURON

Chemical name: 1-n-butyl-3-(3,4-dichlorophenyl)-  
1-methylurea  
Other names: Kloben, Neburea  
Action: Herbicide

PARAQUAT

Chemical name: 1,1'-dimethyl-4,4'-bipyridinium ion  
Other names: Gramoxone, Weedol  
Action: Herbicide

PARATHION

Chemical name: 0,0-diethyl 0-p-nitrophenyl phos-  
phorothioate  
Other names: AAT, Alkron, Alleron, Aphonite, Corothion,  
DNTP, Ethyl Parathion, Etilon, Folidol, Niran,  
Nitrostigmine, Orthophos, Panthion, Paramar, Paraphos,  
Parathene, Parawet, Phoskil, Rhodiatox, SNP, Soprathion,  
Stathion, Thiophos  
Action: Insecticide

PHORATE

Chemical name: 0,0-diethyl S-(ethylthio)-methyl  
phosphorodithioate  
Other names: Thimet, Timet  
Action: Insecticide

PHOSDRIN

See MEVINPHOS

PICLORAM

Chemical name: 4-amino-3,5,6-trichloropicolinic acid  
Other names: Borolin, Tordon  
Action: Herbicide

POLYCHLORINATED BIPHENYLS

Chemical name: mixture of chlorinated terphenyls  
Other names: Chlorinated biphenyls, PCB's, Aroclors  
Action: Insecticide

PROMETONE

Chemical name: 2-methoxy-4,6-bis(isopropylamino)-  
s-triazine  
Other names: Gesafram, Pramitol, Prometon  
Action: Herbicide

PROPOXUR

Chemical name: o-isopropoxyphenyl methylcarbamate  
Other names: Arprocarb, Baygon, Blattanex, Suncide, Unden  
Action: Insecticide

SEVIN

See CARBARYL

SILVEX

Chemical name: 2-(2,4,5-trichlorophenoxy)propionic acid  
Other names: Esteron, Fenoprop, Garlon, Kuron, Kurosai,  
2,4,5-TP  
Action: Herbicide

SIMAZINE

Chemical name: 2-chloro-4,6-bis (ethylamino)-s-triazine  
Other names: Gesatop, Princep  
Action: Herbicide

2,4,5-T

Chemical name: 2,4,5-trichlorophenoxyacetic acid  
Other names: Ded-Weed Brush Killer, Esteron 245  
Concentrate, Fence Rider, Inverton 245, Line Rider, Reddon  
Action: Herbicide

TDE

Chemical name: 2,2-bis(p-chlorophenyl)-1,1-dichloroethane  
Other names: DDD, Rhothane  
Action: Insecticide

TEPP

Chemical name: tetraethyl pyrophosphate and other  
ethyl phosphates  
Other names: Bladan, HETP, Kilmit 40, TEP, Tetron,  
Vapotone  
Action: Insecticide

THIODAN

See ENDOSULFAN

TORDON

See PICLORAM

TOXAPHENE

Chemical name: mixture of various chlorinated camphenes  
Other names: Alltox, Chlorinated camphene, Octachloro-  
camphene, Phenacide, Phenatox, Polychlorocamphene,  
Strobane-T, Toxakil  
Action: Insecticide

TRICHLORFON

Chemical name: dimethyl (2,2,2-trichloro-1-hydroxyethyl)  
phosphonate  
Other names: Anthon, Chlorofos, Dipterex, Dylox,  
Neguvon, Trichlorphon, Tugon  
Action: Insecticide

TRIFLURALIN

Chemical name: a,a,a-trifluoro-2,6-dinitro-N,N-dipropyl-  
p-toluidine  
Other names: Elancolan, Treflan  
Action: Herbicide



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Appendix Table 1. PESTICIDE INTAKE FROM FOOD AND WATER

	(1)	(2)	(3)	(4)	(5)
	Maximum concentration in water samples from 5-year survey <u>b/</u> ugm/l (ppb)	Computed daily intake (Column 1 x 2 liters per person per day) Mg	6-year average daily intake from food by 70 kg person <u>c/</u> Mg	WHO-FAO acceptable daily intake <u>d/</u> 70 kg person Mg	Fraction: intake from water (Column 2) intake from food (Column 3)
Aldrin	0.085	0.0002			
Dieldrin	0.407	0.0008			
Total	0.492	0.0010	0.006	0.007	1/6
Chlordane	0.169	0.0003			
DDT	0.316	0.0006			
DDE	0.050	0.0001			
DDD	0.840	0.0018			
Total	1.206	0.0025	0.049	0.35	1/20
Endrin	0.133	0.0003	0.0004		3/4
Heptachlor	0.048	0.0001			
Heptachlor Epoxide	0.067	0.0001			
Total	0.115	0.0002	0.002	0.035	1/10
Lindane (BHC)	0.112	0.0002	0.002	0.875	1/10
Total organo- phosphates <u>a/</u>	0.380	0.0008	0.013	0.35	1/16
Total chlorinated pesticides		0.0045	0.077		1/17

a/ Columns 1 and 2 are organophosphates plus carbamates. Columns 3 and 4 are organophosphates only. Column 5 therefore overestimates contribution from water of organophosphates. Column 4 is the value of the most toxic organophosphate (Parathion).

b/ Data from Lichtenberg, et al. (1970).

c/ Data from Duggan and Corneliussen (1972).

d/ Acceptable daily intake set by WHO-FAO expert committees, as presented by Duggan and Corneliussen (1972).

APPENDIX TABLE 2. TOXICITY DATA ON PESTICIDES FOR FRESHWATER ORGANISMS

## ORGANOCHLORINE INSECTICIDES

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	
ALDRIN	<u>CRUSTACEANS</u>				
	<u>Gammarus lacustris</u>	9800	96		Sanders, 1969
	<u>Gammarus fasciatus</u>	4300	96		Sanders, in press
	<u>Palaemonetes kadiakensis</u>	50	96		Sanders, in press
	<u>Asellus brevicaudus</u>	8	96		Sanders, in press
	<u>Daphnia pulex</u>	28	48		Sanders and Cope, 1966
	<u>Simocephalus serrulatus</u>	23	48		Sanders and Cope, 1966
	<u>INSECTS</u>				
	<u>Pteronarcys californica</u>	1.3	96		Sanders and Cope, 1968
	<u>Pteronarcys californica</u>	180	96	2.5 ug/liter (30-day LC-50)	Jensen and Gaufin, 1966
	<u>Acroneuria pacifica</u>	200	96	2.2 ug/liter (30-day LC-50)	Jensen and Gaufin, 1966
	<u>FISHES</u>				
	<u>Pimephales promelas</u>	28	96		Henderson, et al., 1959
	<u>Lepomis macrochirus</u>	13	96		Henderson, et al., 1959
	<u>Salmo gairdneri</u>	17.7	96		Katz, 1961
	<u>Oncorhynchus kisutch</u>	45.9	96		Katz, 1961
	<u>Oncorhynchus tshawytscha</u>	7.5	96		Katz, 1961
DDT	<u>CRUSTACEANS</u>				
	<u>Gammarus lacustris</u>	1.0	96		Sanders, 1969
	<u>Gammarus fasciatus</u>	0.8	96		Sanders, in press
	<u>Palaemonetes kadiakensis</u>	2.3	96		Sanders, in press
	<u>Orconectes nais</u>	0.24	96		Sanders, in press
	<u>Asellus brevicaudus</u>	4.0	96		Sanders, in press
	<u>Simocephalus serrulatus</u>	2.5	48		Sanders and Cope, 1966
	<u>Daphnia pulex</u>	0.36	48		Sanders and Cope, 1966
	<u>INSECTS</u>				
	<u>Pteronarcys californica</u>	7.0	96		Sanders and Cope, 1968
	<u>Pteronarcella badia</u>	1.9	96		Sanders and Cope, 1968
	<u>Claassenia sabulosa</u>	3.5	96		Sanders and Cope, 1968

APPENDIX TABLE 2 (continued)

## ORGANOCHLORINE INSECTICIDES

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	
DDT	<u>FISHES</u>				
	<u>Pimephales promelas</u>	19	96		Macek and McAllister, 1970
	<u>Lepomis macrochirus</u>	8	96		Macek and McAllister, 1970
	<u>Lepomis microlophus</u>	5	96		Macek and McAllister, 1970
	<u>Micropterus salmoides</u>	2	96		Macek and McAllister, 1970
	<u>Salmo gairdneri</u>	7	96		Macek and McAllister, 1970
	<u>Salmo gairdneri</u>			0.26 ug/l (15-day LC-50)	FPRL Annual Report
	<u>Salmo trutta</u>	2	96		Macek and McAllister, 1970
	<u>Oncorhynchus kisutch</u>	4	96		Macek and McAllister, 1970
	<u>Perca flavescens</u>	9	96		Macek and McAllister, 1970
	<u>Ictalurus punctatus</u>	16	96		Macek and McAllister, 1970
	<u>Ictalurus melas</u>	5	96		Macek and McAllister, 1970
TDE (DDD) RHOTHANE ®	<u>CRUSTACEANS</u>				
	<u>Gammarus lacustris</u>	0.64	96		Sanders, 1969
	<u>Gammarus fasciatus</u>	0.86	96		Sanders, in press
	<u>Palaemonetes kadiakensis</u>	0.68	96		Sanders, in press
	<u>Asellus breviacaudus</u>	10.0	96		Sanders, in press
	<u>Simocephalus serrulatus</u>	4.5	48		Sanders and Cope, 1966
	<u>Daphnia pulex</u>	3.2	48		Sanders and Cope, 1966
	<u>INSECT</u>				
DIELDRIN	<u>Pteronarcys californica</u>	380	96		Sanders and Cope, 1968
	<u>CRUSTACEANS</u>				
	<u>Gammarus lacustris</u>	460	96		Sanders, 1969
	<u>Gammarus fasciatus</u>	600	96		Sanders, in press
	<u>Palaemonetes kadiakensis</u>	20	96		Sanders, in press
	<u>Orconectes nais</u>	740	96		Sanders, in press
	<u>Asellus brevicaudus</u>	5	96		Sanders, in press
	<u>Simocephalus serrulatus</u>	190	48		Sanders and Cope, 1966
	<u>Daphnia pulex</u>	250	48		Sanders and Cope, 1966

APPENDIX TABLE 2 (continued)

## ORGANOCHLORINE INSECTICIDES

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	
DIELDRIN	<u>INSECTS</u>				
	<u>Pteronarcys californica</u>	0.5	96		Sanders and Cope, 1968
	<u>Pteronarcys californica</u>	39	96	2.0 (30-day LC-50)	Jensen and Gaufin, 1966
	<u>Acroneuria pacifica</u>	24	96	0.2 (30-day LC-50)	Jensen and Gaufin, 1966
	<u>Pteronarcella badia</u>	0.5	96		Sanders and Cope, 1968
	<u>Claassenia sabulosa</u>	0.58	96		Sanders and Cope, 1968
	<u>FISHES</u>				
	<u>Pimephales promelas</u>	16	96		Henderson, et al., 1959
	<u>Lepomis macrochirus</u>	8	96		Henderson, et al., 1959
	<u>Salmo gairdneri</u>	10	96		Katz, 1961
	<u>Oncorhynchus kisutch</u>	11	96		Katz, 1961
	<u>Oncorhynchus tshawytscha</u>	6	96		Katz, 1961
	<u>Poecillia latipinna</u>			3.0 (19-week LC-50)	Lane and Livingston, 1970
	<u>Poecillia latipinna</u>			0.75 (reduced growth & reproduction - 34-week)	Lane and Livingston, 1970
	<u>Lepomis gibbosus</u>	6.7	96	1.7 (affected swimming ability and oxygen consumption - 100-day)	Cairns and Scheir, 1964 FPRL
	<u>Ictalurus punctatus</u>	4.5	96		
CHLORDANE	<u>CRUSTACEANS</u>				
	<u>Gammarus lacustris</u>	26	96		Sanders, 1969
	<u>Gammarus fasciatus</u>	40	96		Sanders, in press
	<u>Palaemonetes kadiakensis</u>	4.0	96	2.5 (120-hour LC-50)	Sanders, in press
	<u>Simocephalus serrulatus</u>	20	48		Sanders and Cope, 1966
	<u>Daphnia pulex</u>	29	48		Sanders and Cope, 1966
	<u>INSECT</u>				
	<u>Pteronarcys californica</u>	15	96		Sanders and Cope, 1968

APPENDIX TABLE 2 (continued)

## ORGANOCHLORINE INSECTICIDES

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	
CHLORDANE	<u>FISHES</u>				
	<u>Pimephales promelas</u>	52	96		Henderson, et al., 1959
	<u>Lepomis macrochirus</u>	22	96		Henderson, et al., 1959
	<u>Salmo gairdneri</u>	44	96		Katz, 1961
	<u>Oncorhynchus kisutch</u>	56	96		Katz, 1961
	<u>Oncorhynchus tshawytscha</u>	57	96		Katz, 1961
ENDOSULFAN THIODAN	<u>CRUSTACEANS</u>				
	<u>Gammarus fasciatus</u>	6.0	96		Sanders, 1969
	<u>Daphnia magna</u>	52.9	96		Schoettger, 1970
	<u>INSECTS</u>				
	<u>Pteronarcys californica</u>	2.3	96		Sanders and Cope, 1968
	<u>Ischnura sp.</u>	71.8	96		Schoettger, 1970
	<u>FISHES</u>				
	<u>Salmo gairdneri</u>	0.3	96		Schoettger, 1970
	<u>Catostomus commersoni</u>	3.0	96		Schoettger, 1970
	<u>CRUSTACEANS</u>				
ENDRIN	<u>Gammarus lacustris</u>	3.0	96		Sanders, 1969
	<u>Gammarus fasciatus</u>	0.9	120		Sanders, in press
	<u>Palaemonetes kadiakensis</u>	0.4	120		Sanders, in press
	<u>Orconectes nais</u>	3.2	96		Sanders, in press
	<u>Asellus brevicaudus</u>	1.5	96		Sanders, in press
	<u>Simocephalus serrulatus</u>	26	48		Sanders and Cope, 1966
	<u>Daphnia pulex</u>	20	48		Sanders and Cope, 1966
	<u>INSECTS</u>				
	<u>Pteronarcys californica</u>	0.25	96		Sanders and Cope, 1968
	<u>Pteronarcys californica</u>	2.4	96	1.2 (30-day LC-50)	Jensen and Gaufin, 1966
	<u>Acroneuria pacifica</u>	0.32	96	0.03 (30-day LC-50)	Jensen and Gaufin, 1966

APPENDIX TABLE 2 (continued)

## ORGANOCHLORINE INSECTICIDES

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	
ENDRIN	<u>INSECTS</u>				
	<u>Pteronarcella badia</u>	0.54	96		Sanders and Cope, 1968
	<u>Claassenia sabulosa</u>	0.76	96		Sanders and Cope, 1968
	<u>FISHES</u>				
	<u>Pimephales promelas</u>	0.5	96		Henderson, et al., 1959
	<u>Lepomis macrochirus</u>	0.6	96		Henderson, et al., 1959
	<u>Salmo gairdneri</u>	0.6	96		Katz, 1961
	<u>Oncorhynchus kisutch</u>	0.5	96		Katz, 1961
	<u>Oncorhynchus tshawytscha</u>	1.2	96		Katz, 1961
	<u>CRUSTACEANS</u>				
HEPTACHLOR	<u>Gammarus lacustris</u>	29	96		Sanders, 1969
	<u>Gammarus fasciatus</u>	40	96		Sanders, in press
	<u>Palaemonetes kadiakensis</u>	1.8	96		Sanders, in press
	<u>Orconectes nais</u>	7.8	96		Sanders, in press
	<u>Simocephalus serrulatus</u>	47	48		Sanders and Cope, 1966
	<u>Daphnia pulex</u>	42	48		Sanders and Cope, 1966
	<u>INSECTS</u>				
	<u>Pteronarcys californica</u>	1.1	96		Sanders and Cope, 1968
	<u>Pteronarcella badia</u>	0.9	96		Sanders and Cope, 1968
	<u>Claassenia sabulosa</u>	2.8	96		Sanders and Cope, 1968
	<u>FISHES</u>				
	<u>Pimephales promelas</u>	56	96		Henderson, et al., 1959
	<u>Lepomis macrochirus</u>	19	96		Henderson, et al., 1959
	<u>Lepomis microlophus</u>	17	96		Bridges, 1961
	<u>Salmo gairdneri</u>	19	96		Katz, 1961
	<u>Oncorhynchus kisutch</u>	59	96		Katz, 1961
	<u>Oncorhynchus tshawytscha</u>	17	96		Katz, 1961

APPENDIX TABLE 2 (continued)

## ORGANOCHLORINE INSECTICIDES

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	
LINDANE	<u>CRUSTACEANS</u>				
	<u>Gammarus lacustris</u>	48	96		Sanders, 1969
	<u>Gammarus fasciatus</u>	10	96		Sanders, in press
	<u>Asellus brevicaudus</u>	10	96		Sanders, in press
	<u>Simocephalus serrulatus</u>	520	48		Sanders and Cope, 1966
	<u>Daphnia pulex</u>	460	48		Sanders and Cope, 1966
	<u>INSECT</u>				
	<u>Pteronarcys californica</u>	4.5	96		Sanders and Cope, 1968
	<u>FISHES</u>				
	<u>Pimephales promelas</u>	87	96		Macek and McAllister, 1970
	<u>Lepomis macrochirus</u>	68	96		Macek and McAllister, 1970
	<u>Lepomis microlophus</u>	83	96		Macek and McAllister, 1970
	<u>Micropterus salmoides</u>	32	96		Macek and McAllister, 1970
	<u>Salmo gairdneri</u>	27	96		Macek and McAllister, 1970
	<u>Salmo trutta</u>	2	96		Macek and McAllister, 1970
	<u>Oncorhynchus kisutch</u>	41	96		Macek and McAllister, 1970
	<u>Perca flavescens</u>	68	96		Macek and McAllister, 1970
	<u>Ictalurus punctatus</u>	44	96		Macek and McAllister, 1970
	<u>Ictalurus melas</u>	64	96		Macek and McAllister, 1970
METHOXYCHLOR	<u>CRUSTACEANS</u>				
	<u>Gammarus lacustris</u>	0.8	96		Sanders, 1969
	<u>Gammarus fasciatus</u>	1.9	96		Sanders, in press
	<u>Palaemonetes kadiakensis</u>	1.0	96		Sanders, in press
	<u>Orconectes nais</u>	0.5	96		Sanders, in press
	<u>Asellus brevicaudus</u>	3.2	96		Sanders, in press
	<u>Simocephalus serrulatus</u>	5	48		Sanders and Cope, 1966
	<u>Daphnia pulex</u>	0.78	48		Sanders and Cope, 1966

APPENDIX TABLE 2 (continued)

## ORGANOCHLORINE INSECTICIDES

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	
METHOXYCHLOR	<u>INSECTS</u>				
	<u>Pteronarcys californica</u>	1.4	96		Sanders and Cope, 1968
	<u>Taeniopteryx nivalis</u>	0.98	96		Merna (Univ. of Mich.)
	<u>Stenonema</u> spp.	0.63	96		Merna (Univ. of Mich.)
	<u>FISHES</u>				
	<u>Pimephales promelas</u>	7.5	96	0.125 (reduced egg hatch- ability)	Merna (Univ. of Mich.)
	<u>Lepomis macrochirus</u>	62.0	96		Henderson, et al., 1959
	<u>Salmo gairdneri</u>	62.0	96		Katz, 1961
	<u>Oncorhynchus kisutch</u>	66.2	96		Katz, 1961
	<u>Oncorhynchus tshawytscha</u>	27.9	96		Katz, 1961
	<u>Perca flavescens</u>	20.0	96	0.6 (reduced growth - 8-month)	Merna (Univ. of Mich.)
TOXAPHENE	<u>CRUSTACEANS</u>				
	<u>Gammarus lacustris</u>	26	96		Sanders, 1969
	<u>Gammarus fasciatus</u>	6	96		Sanders, in press
	<u>Palaemonetes kadiakensis</u>	28	96		Sanders, in press
	<u>Simocephalus serrulatus</u>	10	48		Sanders and Cope, 1966
	<u>Daphnia pulex</u>	15	48		Sanders and Cope, 1966
	<u>INSECTS</u>				
	<u>Pteronarcys californica</u>	2.3	96		Sanders and Cope, 1968
	<u>Pteronarcella badia</u>	3.0	96		Sanders and Cope, 1968
	<u>Claassenia sabulosa</u>	1.3	96		Sanders and Cope, 1968
	<u>FISHES</u>				
	<u>Pimephales promelas</u>	14	96		Macek and McAllister, 1970
	<u>Lepomis macrochirus</u>	18	96		Macek and McAllister, 1970
	<u>Lepomis microlophus</u>	13	96		Macek and McAllister, 1970
	<u>Micropterus salmoides</u>	2	96		Macek and McAllister, 1970



APPENDIX TABLE 2 (continued)

ORGANOCHLORINE INSECTICIDES

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u>		<u>SUB-ACUTE EFFECTS</u>	<u>REFERENCE</u>
		<u>LC-50</u>			
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	
TOXAPHENE	<u>FISHES</u>				
	<u>Salmo gairdneri</u>	11	96		Macek and McAllister, 1970
	<u>Salmo trutta</u>	3	96		Macek and McAllister, 1970
	<u>Oncorhynchus kisutch</u>	8	96		Macek and McAllister, 1970
	<u>Perca flavescens</u>	12	96		Macek and McAllister, 1970
	<u>Ictalurus punctatus</u>	13	96		Macek and McAllister, 1970
	<u>Ictalurus melas</u>	5	96		Macek and McAllister, 1970

APPENDIX TABLE 2 (continued)

## ORGANOPHOSPHORUS INSECTICIDES

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter</u>	
ABATE ®	<u>CRUSTACEAN</u>					
	<u>Gammarus lacustris</u>	82	96			Sanders, 1969
	<u>INSECT</u>					
	<u>Pteronarcys californica</u>	10	96			Sanders and Cope, 1968
	<u>FISH</u>					
	<u>Salmo gairdneri</u>	158	96			FPRL
56 AZINPHOSMETHYL GUTHION ®	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	0.15	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	0.10	96			Sanders, in press
	<u>Gammarus pseudolimneaus</u>				0.10 (30-day)	Bell, unpublished
	<u>Palaemonetes kadiakensis</u>	1.2	120	0.16 (20-day LC-50)		Sanders, in press
	<u>Asellus brevicaudus</u>	21.0	96			Sanders, in press
	<u>INSECTS</u>					
	<u>Pteronarcys dorsata</u>	12.1	96	4.9 (30-day LC-50)		Bell, unpublished
	<u>Pteronarcys californica</u>	1.5	96			Sanders and Cope, 1968
	<u>Acroneuria lycorias</u>			1.5 (30-day LC-50)	1.36 (30-day)	Bell, unpublished
	<u>Ophiogomphus rupinsulensis</u>	12.0	96	2.2 (30-day LC-50)	1.73 (30-day)	Bell, unpublished
	<u>Hydropsyche bettoni</u>			7.4 (30-day LC-50)	4.94 (30-day)	Bell, unpublished
	<u>Ephemerella subvaria</u>			4.5 (30-day LC-50)	2.50 (30-day)	Bell, unpublished
	<u>FISHES</u>					
	<u>Pimephales promelas</u>	93	96			Henderson, 1959
	<u>Lepomis macrochirus</u>	5.2	96			Henderson, 1959
	<u>Lepomis microlophus</u>	52	96			Macek and McAllister, 1970
	<u>Micropterus salmoides</u>	5	96			Macek and McAllister, 1970
	<u>Salmo gairdneri</u>	14	96			Macek and McAllister, 1970
	<u>Salmo trutta</u>	17	96			Macek and McAllister, 1970
	<u>Oncorhynchus kisutch</u>	17	96			Macek and McAllister, 1970

APPENDIX TABLE 2 (continued)

## ORGANOPHOSPHORUS INSECTICIDES

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter</u>	
AZINPHOSMETHYL GUTHION ®	<u>FISHES</u>					
	<u>Perca flavescens</u>	13	96			Macek and McAllister, 1970
	<u>Ictalurus punctatus</u>	3290	96			Macek and McAllister, 1970
	<u>Ictalurus melas</u>	3500	96			Macek and McAllister, 1970
AZINPHOSETHYL ETHYL GUTHION ®	<u>CRUSTACEANS</u>					
	<u>Simocephalus serrulatus</u>	4	48			Sanders and Cope, 1966
	<u>Daphnia pulex</u>	3.2	48			Sanders and Cope, 1966
	<u>FISH</u>					
	<u>Salmo gairdneri</u>	19	96			FPRL
CARBOPHENOTHION TRITHION ®	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	5.2	96			Sanders, 1969
	<u>Palaemonetes kadiakensis</u>	1.2	96			Sanders, in press
	<u>Asellus brevicaudus</u>	1100	96			Sanders, in press
CHLOROTHION	<u>CRUSTACEAN</u>					
	<u>Daphnia magna</u>	4.5	48			"Water Quality Criteria", 1968
	<u>FISHES</u>					
	<u>Pimephales promelas</u>	2700	96			Pickering, et al., 1962
	<u>Lepomis macrochirus</u>	700	96			Pickering, et al., 1962
CIODRIN ®	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	15	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	11	96			Sanders, in press
	<u>FISHES</u>					
	<u>Lepomis macrochirus</u>	250	96			FPRL
	<u>Micropterus salmoides</u>	1100	96			FPRL

## APPENDIX TABLE 2 (continued)

## ORGANOPHOSPHORUS INSECTICIDES

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter</u>	
CIODRIN ®	<u>FISHES</u>					
	<u>Salmo gairdneri</u>	55	96			FPRL
	<u>Ictalurus punctatus</u>	2500	96			FPRL
COUMAPHOS CO-RAL ®	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	0.07	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	0.15	96			Sanders, in press
	<u>Daphnia magna</u>	1.0	48			"Water Quality Criteria", 1968
INSECTS	<u>Hydropsyche sp.</u>	5	24			Carlson, 1966
	<u>Hexagenia sp.</u>	430	24			Carlson, 1966
FISHES	<u>Pimephales promelas</u>	18000	96			Katz, 1961
	<u>Lepomis macrochirus</u>	180	96			Henderson, 1959
	<u>Salmo gairdneri</u>	1500	96			Katz, 1961
	<u>Oncorhynchus kisutch</u>	15000	96			Katz, 1961
DEMETON SYSTOX ®	<u>CRUSTACEAN</u>					
	<u>Gammarus fasciatus</u>	27	96			Sanders, in press
FISHES	<u>Pimephales promelas</u>	3200	96			Pickering, et al., 1962
	<u>Lepomis macrochirus</u>	100	96			Pickering, et al., 1962
DIAZINON	<u>CRUSTACEANS</u>					
	<u>Gammarus pseudolimneaus</u>			0.27 (30-day LC-50)	0.20 (30-day)	Bell, (NWQL - unpublished)
	<u>Gammarus lacustris</u>	200	96			Sanders, 1969
	<u>Simocephalus serrulatus</u>	1.4	48			Sanders and Cope, 1966
	<u>Daphnia pulex</u>	0.90	48			Sanders and Cope, 1966
	<u>Daphnia magna</u>				0.26 (21-day)	Biesinger, (NWQL - unpub.)

APPENDIX TABLE 2 (continued)

## ORGANOPHOSPHORUS INSECTICIDES

PESTICIDE	ORGANISM	ACUTE TOXICITY LC-50		SUB-ACUTE EFFECTS	NO EFFECT		REFERENCE
		ug/liter	hours		ug/liter	ug/liter	
DIAZINON	INSECTS						
	<u>Pteronarcys californica</u>	25	96				Sanders and Cope, 1968
	<u>Pteronarcys dorsata</u>			4.6 (30-day LC-50)	3.29 (30-day)		Bell, (NWQL - unpublished)
	<u>Acroneuria lyctorias</u>	1.7	96	1.25 (30-day LC-50)	0.83 (30-day)		Bell, (NWQL - unpublished)
	<u>Ophiogomphus rupinsulensis</u>			2.2 (30-day LC-50)	1.29 (30-day)		Bell, (NWQL - unpublished)
	<u>Hydropsyche pettoni</u>			3.54 (30-day LC-50)	1.79 (30-day)		Bell, (NWQL - unpublished)
	<u>Ephemerella subvaria</u>			1.05 (30-day LC-50)	0.42 (30-day)		Bell, (NWQL - unpublished)
DICHLOROVOS	CRUSTACEANS						
	DDVP						
	<u>Gammarus lacustris</u>	0.50	96				Sanders, 1969
	VAPONA (R)						
	<u>Gammarus fasciatus</u>	0.40	96				Sanders, in press
	<u>Simocephalus serrulatus</u>	0.26	48				Sanders and Cope, 1966
	<u>Daphnia pulex</u>	0.07	48				Sanders and Cope, 1966
	INSECT						
	<u>Pteronarcys californica</u>	0.10	96				Sanders and Cope, 1968
	FISH						
DIOXATHION	<u>Lepomis macrochirus</u>	869	96				FPRL
	DELNAV (R)						
	CRUSTACEANS						
	<u>Gammarus lacustris</u>	270	96				Sanders, 1969
	<u>Gammarus fasciatus</u>	8.6	96				Sanders, in press
	FISHES						
	<u>Pimephales promelas</u>	9300	96				Pickering, et al., 1962
	<u>Lepomis macrochirus</u>	34	96				Pickering, et al., 1962
	<u>Lepomis cyanellus</u>	61	96				Pickering, et al., 1962
	<u>Micropterus salmoides</u>	36	96				Pickering, et al., 1962
DISULFOTON	CRUSTACEANS						
	DI-SYSTON (R)						
	<u>Gammarus lacustris</u>	52	96				Sanders, 1969
	<u>Gammarus fasciatus</u>	21	96				Sanders, in press
	<u>Palaemonetes kadiakensis</u>	38	96				Sanders, in press

APPENDIX TABLE 2 (continued)

## ORGANOPHOSPHORUS INSECTICIDES

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter</u>	
DISULFOTON DI-SYSTON ®	<u>INSECTS</u>					
	<u>Pteronarcys californica</u>	5	96			Sanders and Cope, 1968
	<u>Pteronarcys californica</u>	21.4	96	1.7 (30-day LC-50)		Jensen and Gaufin, 1964
	<u>Acroneuria pacifica</u>	8.4	96	1.2 (30-day LC-50)		Jensen and Gaufin, 1964
	<u>FISHES</u>					
	<u>Pimephales promelas</u>	63	96			Pickering, et al., 1962
	<u>Lepomis macrochirus</u>	3700	96			Pickering, et al., 1962
	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	0.11	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	0.32	96			Sanders, in press
DURSBAN ®	<u>INSECTS</u>					
	<u>Pteronarcys californica</u>	10	96			Sanders and Cope, 1968
	<u>Pteronarcella badia</u>	0.38	96			Sanders and Cope, 1968
	<u>Claassenia sabulosa</u>	0.57	96			Sanders and Cope, 1968
	<u>FISHES</u>					
	<u>Lepomis macrochirus</u>	2.6	96			FPRL
	<u>Salmo gairdneri</u>	11	96			FPRL
	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	1.8	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	9.4	96			Sanders, in press
ETHION NIALATE ®	<u>Palaemonetes kadiakensis</u>	5.7	96			Sanders, in press
	<u>INSECT</u>					
	<u>Pteronarcys californica</u>	2.8	96			Sanders and Cope, 1968
	<u>FISHES</u>					
	<u>Lepomis macrochirus</u>	220	96			FPRL
	<u>Micropterus salmoides</u>	150	96			FPRL
	<u>Salmo gairdneri</u>	560	96			FPRL
	<u>Salmo clarkii</u>	720	96			FPRL
	<u>Ictalurus punctatus</u>	7500	96			FPRL

## APPENDIX TABLE 2 (continued)

## ORGANOPHOSPHORUS INSECTICIDES

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter</u>	
EPN	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	15	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	7	96			Sanders, in press
	<u>Palaemonetes kadiakensis</u>	0.56	96			Sanders, in press
	<u>FISHES</u>					
	<u>Pimephales promelas</u>	110	96			Solon and Nair, 1970
	<u>Lepomis macrochirus</u>	100	96			Pickering, et al., 1962
	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	8.4	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	110	96			Sanders, in press
FENTHION BAYTEX®	<u>Palaemonetes kadiakensis</u>	5	120	1.5 (20-day LC-50)		Sanders, in press
	<u>Orconectes nais</u>	50	96			Sanders, in press
	<u>Asellus brevicaudus</u>	1800	96			Sanders, in press
	<u>Simocephalus serrulatus</u>	0.62	48			Sanders and Cope, 1966
	<u>Daphnia pulex</u>	0.80	48			Sanders and Cope, 1966
	<u>INSECT</u>					
	<u>Pteronarcys californica</u>	4.5	96			Sanders and Cope, 1968
	<u>FISHES</u>					
	<u>Pimephales promelas</u>	2440	96			Macek and McAllister, 1970
	<u>Lepomis macrochirus</u>	1380	96			Macek and McAllister, 1970
	<u>Lepomis microlophus</u>	1880	96			Macek and McAllister, 1970
	<u>Micropterus salmoides</u>	1540	96			Macek and McAllister, 1970
	<u>Salmo gairdneri</u>	930	96			Macek and McAllister, 1970
	<u>Salmo trutta</u>	1330	96			Macek and McAllister, 1970
	<u>Oncorhynchus kisutch</u>	1320	96			Macek and McAllister, 1970
	<u>Perca flavescens</u>	1650	96			Macek and McAllister, 1970
	<u>Ictalurus punctatus</u>	1680	96			Macek and McAllister, 1970
	<u>Ictalurus melas</u>	1620	96			Macek and McAllister, 1970

## APPENDIX TABLE 2 (continued)

## ORGANOPHOSPHORUS INSECTICIDES

PESTICIDE	ORGANISM	ACUTE TOXICITY		SUB-ACUTE EFFECTS	NO EFFECT		REFERENCE
		LC-50					
		ug/liter	hours	ug/liter		ug/liter	
MALATHION	CRUSTACEANS						
	<u>Gammarus pseudolimneaus</u>			0.023 (30-day LC-50)		0.008 (30-day)	Bell, (NWQL - unpublished)
	<u>Gammarus lacustris</u>	1.0	96				Sanders, 1969
	<u>Gammarus fasciatus</u>	0.76	96	0.5 (120-hour LC-50)			Sanders, in press
	<u>Palaemonetes kadiakensis</u>	12	96	9.0 (120-hour LC-50)			Sanders, in press
	<u>Orconectes nais</u>	180	96				Sanders, in press
	<u>Asellus brevicaudus</u>	3000	96				Sanders, in press
	<u>Simocephalus serrulatus</u>	3.5	48				Sanders and Cope, 1966
	<u>Daphnia pulex</u>	1.8	48				Sanders and Cope, 1966
	<u>Daphnia magna</u>					0.6 (21-day)	Biesinger, (NWQL - unpub.)
	INSECTS						
	<u>Pteronarcys californica</u>	10	96				Sanders and Cope, 1968
	<u>Pteronarcys dorsata</u>			11.1 (30-day LC-50)		9.4 (30-day)	Bell, (NWQL - unpublished)
	<u>Acroneuria lycorias</u>	1.0		0.3 (30-day LC-50)		0.17 (30-day)	Bell, (NWQL - unpublished)
	<u>Pteronarcella badia</u>	1.1	96				Sanders and Cope, 1968
	<u>Claassenia sabulosa</u>	2.8	96				Sanders and Cope, 1968
	<u>Boyeria vinosa</u>			2.3 (30-day LC-50)		1.65 (30-day)	Bell, (NWQL - unpublished)
	<u>Ophiogomphus rupinsulensis</u>			0.52 (30-day LC-50)		0.28 (30-day)	Bell, (NWQL - unpublished)
	<u>Hydropsyche bettoni</u>			0.34 (30-day LC-50)		0.24 (30-day)	Bell, (NWQL - unpublished)
	FISHES						
	<u>Pimephales promelas</u>	9000	96	580 (spinal deformity, 10-month)	200 (10-month exposure)		Mount and Stephan, 1967
	<u>Lepomis macrochirus</u>	110	96	7.4 (spinal deformity, several months)	3.6 (11-month)		Eaton, 1971
	<u>Lepomis cyanelus</u>	120	96				Pickering, et al., 1962
	<u>Lepomis microlophus</u>	170	96				Macek and McAllister, 1970
	<u>Micropterus salmoides</u>	285	96				Macek and McAllister, 1970
	<u>Salmo gairdneri</u>	170	96				Macek and McAllister, 1970
	<u>Salmo trutta</u>	200	96				Macek and McAllister, 1970
	<u>Oncorhynchus kisutch</u>	101	96				Macek and McAllister, 1970
	<u>Perca flavescens</u>	263	96				Macek and McAllister, 1970
	<u>Ictalurus punctatus</u>	8970	96				Macek and McAllister, 1970
	<u>Ictalurus melas</u>	12900	96				Macek and McAllister, 1970



## APPENDIX TABLE 2 (continued)

## ORGANOPHOSPHORUS INSECTICIDES

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter</u>	
METHYL PARATHION BAYER E601	<u>FISHES</u>					
	<u>Pimephales promelas</u>	8900	96			Macek and McAllister, 1970
	<u>Lepomis macrochirus</u>	5720	96			Macek and McAllister, 1970
	<u>Lepomis microlophus</u>	5170	96			Macek and McAllister, 1970
	<u>Micropterus salmoides</u>	5220	96			Macek and McAllister, 1970
	<u>Salmo gairdneri</u>	2750	96			Macek and McAllister, 1970
	<u>Salmo trutta</u>	4740	96			Macek and McAllister, 1970
	<u>Oncorhynchus kisutch</u>	5300	96			Macek and McAllister, 1970
	<u>Perca flavescens</u>	3060	96			Macek and McAllister, 1970
	<u>Italurus punctatus</u>	5710	96			Macek and McAllister, 1970
	<u>Italurus melas</u>	6640	96			Macek and McAllister, 1970
MEVINPHOS. PHOSDRIN ®	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	130	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	2.8	96			Sanders, in press
	<u>Palaemonetes kadiakensis</u>	12	96			Sanders, in press
	<u>Asellus brevicaudus</u>	56	96			Sanders, in press
	<u>Simocephalus serrulatus</u>	0.43	48			Sanders and Cope, 1966
	<u>Daphnia pulex</u>	0.16	48			Sanders and Cope, 1966
	<u>INSECT</u>					
	<u>Pteronarcys californica</u>	5.0	96			Sanders and Cope, 1968
	<u>FISHES</u>					
	<u>Lepomis macrochirus</u>	70	96			FPRL
	<u>Micropterus salmoides</u>	110	96			FPRL
NALED DIBROM ®	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	110	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	14	96			Sanders, in press
	<u>Palaemonetes kadiakensis</u>	90	96			Sanders, in press
	<u>Orconectes nais</u>	1800	96			Sanders, in press
	<u>Asellus brevicaudus</u>	230	96			Sanders, in press
	<u>Simocephalus serrulatus</u>	1.1	48			Sanders and Cope, 1966
	<u>Daphnia pulex</u>	0.35	48			Sanders and Cope, 1966

## APPENDIX TABLE 2 (continued)

## ORGANOPHOSPHORUS INSECTICIDES

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter</u>	
NALED DIBROM ®	<u>INSECT</u>					
	<u>Pteronarcys californica</u>	8.0	96			Sanders and Cope, 1968
	<u>FISHES</u>					
	<u>Lepomis macrochirus</u>	180	96			FPRL
	<u>Salmo gairdneri</u>	132	96			FPRL
OXYDEMETON METHYL META-SYSTOX ®	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	190	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	1000	96			Sanders, in press
	<u>INSECT</u>					
	<u>Pteronarcys californica</u>	35	96			Sanders and Cope, 1968
	<u>FISHES</u>					
	<u>Lepomis macrochirus</u>	14000	96			FPRL
	<u>Salmo gairdneri</u>	4000	96			FPRL
	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	3.5	96			Sanders, 1969
PARATHION	<u>Gammarus fasciatus</u>	2.1	96	1.6 (120-hour LC-50)		Sanders, in press
	<u>Palaemonetes kadiakensis</u>	1.5	96			Sanders, in press
	<u>Simocephalus serrulatus</u>	0.37	48			Sanders and Cope, 1966
	<u>Daphnia pulex</u>	0.60	48			Sanders and Cope, 1966
	<u>Orconectes nais</u>	0.04	96			Sanders, in press
	<u>Asellus brevicaudus</u>	600	96			Sanders, in press
	<u>INSECTS</u>					
	<u>Pteronarcys californica</u>	3.6	96	2.2 (30-day LC-50)		Jensen and Gaufin, 1964
	<u>Pteronarcys dorsata</u>	3.0	96	0.90 (30-day LC-50)		Bell, (NWQL - unpublished)
	<u>Pteronarcella badia</u>	4.2	96			Sanders and Cope, 1968
	<u>Claassenia sabulosa</u>	1.5	96			Sanders and Cope, 1968
	<u>Acroneuria pacifica</u>	3.0	96	0.44 (30-day LC-50)		Jensen and Gaufin, 1964
	<u>Acroneuria lycorias</u>			0.013 (30-day LC-50)		Bell, (NWQL - unpublished)

APPENDIX TABLE 2 (continued)

## ORGANOPHOSPHORUS INSECTICIDES

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter</u>	
PARATHION	<u>INSECTS</u>					
	<u>Ephemerella subvaria</u>	0.16	96	0.056 (30-day LC-50)		Bell, (NWQL - unpublished)
	<u>Ophiogomphus rupinsulensis</u>	3.25	96	0.22 (30-day LC-50)		Bell, (NWQL - unpublished)
	<u>Hydropsyche bettoni</u>			0.45 (30-day LC-50)		Bell, (NWQL - unpublished)
	<u>FISHES</u>					
	<u>Pimephales promelas</u>	1410	96			Solon and Nair, 1970
	<u>Lepomis macrochirus</u>	65	96			Pickering, et al., 1962
	<u>Lepomis cyanellus</u>	425	96			Pickering, et al., 1962
	<u>Micropterus salmoides</u>	190	96			Pickering, et al., 1962
	<u>CRUSTACEANS</u>					
8 PHORATE THIMET ®	<u>Gammarus lacustris</u>	9	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	0.60	96			Sanders, in press
	<u>Orconectes nais</u>	50	96			Sanders, in press
	<u>CRUSTACEANS</u>					
PHOSPHAMIDON	<u>Gammarus lacustris</u>	2.8	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	16	96			Sanders, in press
	<u>Orconectes nais</u>	7500	96			Sanders, in press
	<u>Simocephalus serrulatus</u>	6.6	48			Sanders and Cope, 1966
	<u>Daphnia pulex</u>	8.8	48			Sanders and Cope, 1966
	<u>INSECT</u>					
	<u>Pteronarcys californica</u>	150	96			Sanders and Cope, 1968
	<u>FISHES</u>					
	<u>Pimephales promelas</u>	100000	96			FPRL
	<u>Lepomis macrochirus</u>	4500	96			FPRL
	<u>Ictalurus punctatus</u>	70000	96			FPRL
RONNEL	<u>FISH</u>					
	<u>Pimephales promelas</u>	305	96			Solon and Nair, 1970

## APPENDIX TABLE 2 (continued)

## ORGANOPHOSPHORUS INSECTICIDES

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter</u>	
TEPP	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	39	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	210	96			Sanders, in press
	<u>FISHES</u>					
	<u>Pimephales promelas</u>	840	96			Pickering, et al., 1962
	<u>Lepomis macrochirus</u>	520	96			Pickering, et al., 1962
TRICHLOROPHON DIPTEREX DYLOX	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	40	96			Sanders, 1969
	<u>Simocephalus serrulatus</u>	0.32	48			Sanders and Cope, 1966
	<u>Daphnia pulex</u>	0.18	48			Sanders and Cope, 1966
	<u>INSECTS</u>					
	<u>Pteronarcys californica</u>	69	96	9.8 (30-day LC-50)		Jensen and Gaufin, 1966
	<u>Pteronarcys californica</u>	35	96			Sanders and Cope, 1968
	<u>Acroneuria pacifica</u>	16.5	96	8.7 (30-day LC-50)		Jensen and Gaufin, 1966
	<u>Pteronarcella badia</u>	11	96			Sanders and Cope, 1968
	<u>Claassenia sabulosa</u>	22	96			Sanders and Cope, 1968
	<u>FISHES</u>					
	<u>Pimephales promelas</u>	109000	96			Pickering, et al., 1962
	<u>Lepomis macrochirus</u>	3800	96			Pickering, et al., 1962

APPENDIX TABLE 2 (continued)

## CARBAMATE

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter Time</u>	
CARBARYL SEVIN ®	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	16	96			Sanders and Cope, 1969
	<u>Gammarus fasciatus</u>	26	96			Sanders, in press
	<u>Palaemonetes kadiakensis</u>	5.6	96			Sanders, in press
	<u>Orconectes nais</u>	8.6	96			Sanders, in press
	<u>Asellus brevicaudus</u>	240	96			Sanders, in press
	<u>Simocephalus serrulatus</u>	7.6	48			Sanders and Cope, 1966
	<u>Daphnia pulex</u>	6.4	48			Sanders and Cope, 1966
	<u>Daphnia magna</u>				5.0 (63-day)	Biesigner, (NWQL - unpub.)
	<u>INSECTS</u>					
	<u>Pteronarcys californica</u>	4.8	96			Sanders and Cope, 1968
	<u>Pteronarcys dorsata</u>			23.0 (30-day LC-50)	11.5 (30-day)	Bell, (NWQL - unpublished)
	<u>Pteronarcella badia</u>	1.7	96			Sanders and Cope, 1968
	<u>Claassenia sabulosa</u>	5.6	96			Sanders and Cope, 1968
	<u>Acroneuria lycorias</u>			2.2 (30-day LC-50)	1.3 (30-day)	Bell, (NWQL - unpublished)
	<u>Hydropysche bettoni</u>			2.7 (30-day LC-50)	1.8 (30-day)	Bell, (NWQL - unpublished)
	<u>FISHES</u>					
	<u>Pimephales promelas</u>	9000	96	680 (deline survival and reproduction, 6-month)	210 (6-mo.)	Carlson, (NWQL - unpublished)
	<u>Lepomis macrochirus</u>	6760	96			Macek and McAllister, 1970
	<u>Lepomis microlophus</u>	11200	96			Macek and McAllister, 1970
	<u>Micropterus salmoides</u>	6400	96			Macek and McAllister, 1970
	<u>Salmo gairdneri</u>	4340	96			Macek and McAllister, 1970
	<u>Salmo trutta</u>	1950	96			Macek and McAllister, 1970
	<u>Oncorhynchus kisutch</u>	764	96			Macek and McAllister, 1970
	<u>Perca flavescens</u>	745	96			Macek and McAllister, 1970
	<u>Ictalurus punctatus</u>	15800	96			Macek and McAllister, 1970
	<u>Ictalurus melas</u>	20000	96			Macek and McAllister, 1970
BAYGON	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	34	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	50	96			Sanders, in press
	<u>INSECT</u>					
	<u>Pteronarcys californica</u>	13	96			Sanders and Cope, 1968

APPENDIX TABLE 2 (continued)

<u>PESTICIDE</u>	<u>ORGANISM</u>	CARBAMATE				<u>REFERENCE</u>
		<u>ACUTE TOXICITY</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	
		<u>LC-50</u>				
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter</u>	
AMINOCARB	<u>CRUSTACEAN</u>					
MATACIL	<u>Gammarus lacustris</u>	12	96			Sanders, 1969
ZECTRAN	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	46	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	40	96			Sanders, in press
	<u>Palaemonetes kadiakensis</u>	83	96	25 (20-day LC-50)		Sanders, in press
	<u>Simocephalus serrulatus</u>	13	48			Sanders and Cope, 1966
	<u>Daphnia pulex</u>	10	48			Sanders and Cope, 1966
	<u>INSECT</u>					
	<u>Pteronarcys californica</u>	10	96			Sanders and Cope, 1968
	<u>FISHES</u>					
	<u>Pimephales promelas</u>	17000	96			Macek and McAllister, 1970
	<u>Lepomis macrochirus</u>	11200	96			Macek and McAllister, 1970
	<u>Lepomis microlophus</u>	16700	96			Macek and McAllister, 1970
	<u>Micropterus salmoides</u>	14700	96			Macek and McAllister, 1970
	<u>Salmo gairdneri</u>	10200	96			Macek and McAllister, 1970
	<u>Salmo trutta</u>	8100	96			Macek and McAllister, 1970
	<u>Oncorhynchus kisutch</u>	1730	96			Macek and McAllister, 1970
	<u>Perca flavescens</u>	2480	96			Macek and McAllister, 1970
	<u>Ictalurus punctatus</u>	11400	96			Macek and McAllister, 1970
	<u>Ictalurus melas</u>	16700	96			Macek and McAllister, 1970

APPENDIX TABLE 2 (continued)

## HERBICIDES, FUNGICIDES, DEFOLIANTS

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter</u>	
ACROLEIN AQUALIN	<u>FISHES</u>					
	<u>Lepomis macrochirus</u>	80	24			Bond, et al., 1960
	<u>Salmo trutta</u>	46	24			Burdick, et al., 1964
	<u>Lepomis macrochirus</u>	79	24			Burdick, et al., 1964
AMINOTRIAZOLE AMITROL	<u>CRUSTACEANS</u>					
	<u>Gammarus fasciatus</u>				100,000 ug/l 48 hr.	Sanders, 1970
	<u>Daphnia magna</u>	30000	48			Sanders, 1970
	<u>Cypridopsis vidua</u>	32000	48			Sanders, 1970
	<u>Asellus brevicaudus</u>				100,000 ug/l 48 hr.	Sanders, 1970
	<u>Palaemonetes kadiakensis</u>				100,000 ug/l 48 hr.	Sanders, 1970
	<u>Orconectes nais</u>				100,000 ug/l 48 hr.	Sanders, 1970
	<u>FISHES</u>					
	<u>Lepomis macrochirus</u>	100	48			Sanders, 1970
	<u>Oncorhynchus kisutch</u>	325000	48			Bond, et al., 1960
BALAN	<u>CRUSTACEAN</u>					
	<u>Gammarus fasciatus</u>	1100	96			Sanders, 1970
BENSULFIDE	<u>CRUSTACEAN</u>					
	<u>Gammarus fasciatus</u>	1400	96			Sanders, 1970
CHLOROXURON	<u>FISH</u>					
	<u>Lepomis macrochirus</u>	25000	48			Hughes and Davis, 1964
CIPC	<u>FISH</u>					
	<u>Lepomis macrochirus</u>	8000	48			Hughes and Davis, 1964
DACTHAL	<u>FISH</u>					
	<u>Lepomis macrochirus</u>	700000	48			Hughes and Davis, 1964

APPENDIX TABLE 2 (continued)

## HERBICIDES, FUNGICIDES, DEFOLIANTS

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter</u>	
DALAPON (SODIUM SALT)	<u>CRUSTACEANS</u>					
	<u>Simocephalus serrulatus</u>	16000	48			Sanders and Cope, 1966
	<u>Daphnia pulex</u>	11000	48			Sanders and Cope, 1966
	<u>INSECT</u>					
	<u>Pteronarcys californica</u>				100,000 ug/l 96 hr.	Sanders and Cope, 1968
	<u>FISHES</u>					
	<u>Pimephales promelas</u>	290000	96			Surber and Pickering, 1962
DEF	<u>Lepomis macrochirus</u>	290000	96			Surber and Pickering, 1962
	<u>Oncorhynchus kisutch</u>	340000	48			Bond, et al., 1960
	<u>CRUSTACEAN</u>					
	<u>Gammarus lacustris</u>	100	96			Sanders, 1969
	<u>INSECT</u>					
DEXON	<u>Pteronarcys californica</u>	2100	96			Sanders and Cope, 1968
	<u>CRUSTACEAN</u>					
	<u>Gammarus lacustris</u>	3700	96			Sanders, 1969
	<u>INSECT</u>					
	<u>Pteronarcys californica</u>	24000	96			Sanders and Cope, 1968



APPENDIX TABLE 2 (continued)

## HERBICIDES, FUNGICIDES, DEFOLIANTS

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>			
DICAMBA	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	3900	96			Sanders, 1969
	<u>Gammarus fasciatus</u>			100,000 ug/l	48 hr.	Sanders, 1970
	<u>Daphnia magna</u>			100,000 ug/l	48 hr.	Sanders, 1970
	<u>Cypridopsis vidua</u>			100,000 ug/l	48 hr.	Sanders, 1970
	<u>Asellus brevicaudus</u>			100,000 ug/l	48 hr.	Sanders, 1970
	<u>Palaemonetes kadiakensis</u>			100,000 ug/l	48 hr.	Sanders, 1970
	<u>Orconectes nais</u>			100,000 ug/l	48 hr.	Sanders, 1970
	<u>FISH</u>					
	<u>Lepomis macrochirus</u>	20	48			Hughes and Davis, 1964
DICHLOBENIL CASARON (®)	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	11000	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	10000	96			Sanders, 1970
	<u>Hyallorella azteca</u>	8500	96			Wilson and Bond, 1969
	<u>Simocephalus serrulatus</u>	5800	48			Sanders and Cope, 1968
	<u>Daphnia pulex</u>	3700	48			Sanders and Cope, 1968
	<u>Daphnia magna</u>	10000	48			Sanders, 1970
	<u>Cypridopsis vidua</u>	7800	96			Sanders, 1970
	<u>Asellus brevicaudus</u>	34000	96			Sanders, 1970
	<u>Palaemonetes kadiakensis</u>	9000	96			Sanders, 1970
	<u>Orconectes nais</u>	22000				Sanders, 1970
	<u>INSECTS</u>					
	<u>Pteronarcys californica</u>	7000	96			Sanders and Cope, 1968
	<u>Tendipedid</u>	7800	96			Wilson and Bond, 1969
	<u>Calibrates sp.</u>	10300	96			Wilson and Bond, 1969
	<u>Limnephilus sp.</u>	13000	96			Wilson and Bond, 1969
	<u>Enallegma sp.</u>	20700	96			Wilson and Bond, 1969
	<u>FISH</u>					
	<u>Lepomis macrochirus</u>	20000	48			

APPENDIX TABLE 2 (continued)

## HERBICIDES, FUNGICIDES, DEFOLIANTS

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter</u>	
DICHLONE PHYGON XL	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	1100	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	100	96			Sanders, 1970
	<u>Daphnia magna</u>	125	48			Sanders, 1970
	<u>Cypridopsis vidua</u>	120	48			Sanders, 1970
	<u>Asellus brevicaudus</u>	200	48			Sanders, 1970
	<u>Palaemonetes kadiakensis</u>	450	48			Sanders, 1970
	<u>Orconectes nais</u>	3200	48			Sanders, 1970
	<u>FISHES</u>					
	<u>Lepomis macrochirus</u>	120	48			Bond, et al., 1960
	<u>Micropterus salmoides</u>	70	48			Hughes and Davis, 1962
	<u>CRUSTACEAN</u>					
	<u>Hyallella azteca</u>	48	96			Wilson and Bond, 1969
	<u>INSECTS</u>					
DIQUAT	<u>Callibrates</u> sp.	16400	96			Wilson and Bond, 1969
	<u>Limnephilus</u> sp.	33000	96			Wilson and Bond, 1969
	<u>Tendipedid</u>	100	96			Wilson and Bond, 1969
	<u>Enallagma</u> sp.	100	96			Wilson and Bond, 1969
	<u>FISHES</u>					
	<u>Pimephales promelas</u>	14000	96			Surber and Pickering, 1962
	<u>Lepomis macrochirus</u>	35000	96			Gilderhaus, 1967
	<u>Micropterus salmoides</u>	7800	96			Surber and Pickering, 1962
	<u>Esox lucius</u>	16000	48			Gilderhaus, 1967
	<u>Stizostedion vitreum vitreum</u>	2100	96			Gilderhaus, 1967
	<u>Salmo gairdneri</u>	11200	48			Gilderhaus, 1967
	<u>Oncorhynchus kisutch</u>	28500	48			Bond, et al., 1960

APPENDIX TABLE 2 (continued)

## HERBICIDES, FUNGICIDES, DEFOLIANTS

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter</u>	
DIURON	<u>CRUSTACEAN</u>					
	<u>Gammarus lacustris</u>	160	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	700	96			Sanders, 1970
	<u>Simocephalus serrulatus</u>	2000	48			Sanders and Cope, 1966
	<u>Daphnia pulex</u>	1400	48			Sanders and Cope, 1966
	<u>INSECT</u>					
	<u>Pteronarcys californica</u>	1200	96			Sanders and Cope, 1968
DIFOLITAN	<u>FISH</u>					
	<u>Oncorhynchus kisutch</u>	33000	48			Bond, et al., 1960
	<u>CRUSTACEAN</u>					
	<u>Gammarus lacustris</u>	800	96			Sanders, 1969
	<u>INSECT</u>					
DINITROBUTYL PHENOL	<u>Pteronarcys californica</u>	40	96			Sanders and Cope, 1968
	<u>CRUSTACEAN</u>					
	<u>Gammarus fasciatus</u>	1800	96			Sanders, 1970

APPENDIX TABLE 2 (continued)

## HERBICIDES, FUNGICIDES, DEFOLIANTS

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter</u>	
DIPHENAMID	<u>CRUSTACEANS</u>					
	<u>Gammarus fasciatus</u>				100,000 ug/1 48 hr.	Sanders, 1970
	<u>Daphnia magna</u>	56000	48			Sanders, 1970
	<u>Cypridopsis vidua</u>	50000	48			Sanders, 1970
	<u>Asellus brevicaudus</u>				100,000 ug/1 48 hr.	Sanders, 1970
	<u>Palaemonetes kadiakensis</u>	58000	48			Sanders, 1970
	<u>Orconectes nais</u>				100,000 ug/1 48 hr.	Sanders, 1970
DURSBAN	<u>CRUSTACEAN</u>					
	<u>Gammarus lacustris</u>	110	96			Sanders, 1969
	<u>INSECTS</u>					
	<u>Pteronarcys californica</u>	10	96			Sanders and Cope, 1968
	<u>Pteronarcella badia</u>	0.38	96			Sanders and Cope, 1968
	<u>Claassenia sabulosa</u>	0.57	96			Sanders and Cope, 1968
2-4, D (PGBE)	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	1600	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	2500	96			Sanders, 1970
	<u>Daphnia magna</u>	100	48			Sanders, 1970
	<u>Cypridopsis vidua</u>	320	48			Sanders, 1970
	<u>Asellus brevicaudus</u>	2200	48			Sanders, 1970
	<u>Palaemonetes kadiakensis</u>	2700	48		100,000 ug/1 48 hr.	Sanders, 1970
	<u>Orconectes nais</u>				100,000 ug/1 48 hr.	Sanders, 1970

APPENDIX TABLE 2 (continued)

## HERBICIDES, FUNGICIDES, DEFOLIANTS

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter</u>	
2-4, D (BEE)	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	440	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	5900	48			Sanders, 1970
	<u>Daphnia magna</u>	5600	48			Sanders, 1970
	<u>Cypridopsis vidua</u>	1800	48			Sanders, 1970
	<u>Asellus brevicaudus</u>	3200	48			Sanders, 1970
	<u>Palaemonetes kadiakensis</u>	1400	48		100,000 ug/l 96 hr.	Sanders, 1970
	<u>Orconectes nais</u>	60000	48			Sanders, 1970
	<u>INSECT</u>					
	<u>Pteronarcys californica</u>	1600	96			Sanders and Cope, 1968
2-4, D (IOE)	<u>FISH</u>					
	<u>Pimephales promelas</u>	5600	96	1500 ug/l lethal to eggs in 48 hour exposure	300 ug/l 10 mo.	Mount and Stephan, 1967
	<u>CRUSTACEAN</u>					
	<u>Gammarus lacustris</u>	2400	96			Sanders, 1969
	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	100000	96			Sanders, 1969
	<u>Gammarus fasciatus</u>				100,000 ug/l 48 hr.	Sanders, 1970
	<u>Daphnia magna</u>	4000	48			Sanders, 1970
	<u>Cryptidopsis vidua</u>	8000	48			Sanders, 1970
	<u>Asellus brevicaudus</u>				100,000 ug/l 48 hr.	Sanders, 1970
2-4, D (DIETHYLAMINE SALT)	<u>Palaemonetes kadiakensis</u>				100,000 ug/l 48 hr.	Sanders, 1970
	<u>Orconectes nais</u>				100,000 ug/l 48 hr.	Sanders, 1970

APPENDIX TABLE 2 (continued)

HERBICIDES, FUNGICIDES, DEFOLIANTS

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter</u>	
ENDOTHALL DI SODIUM SALT	<u>FISHES</u>					
	<u>Pimephales notatus</u>	10000	96			Walker, 1964
	<u>Lepomis macrochirus</u>	125000	96			Walker, 1964
	<u>Micropterus salmoides</u>	120000	96			Walker, 1964
	<u>Notropis umbratilis</u>	95000	96			Walker, 1964
ENDOTHALL DIPOTASSIUM SALT	<u>CRUSTACEAN</u>					
	<u>Gammarus lacustris</u>			100,000 ug/1	96 hr.	Sanders, 1969
	<u>FISHES</u>					
	<u>Pimephales promelas</u>	320000	96			Surber and Pickering, 1962
	<u>Lepomis macrochirus</u>	160000	96			Surber and Pickering, 1962
EPTAM	<u>Micropterus salmoides</u>	200000	96			Bond, et al., 1960
	<u>Oncorhynchus tshawytscha</u>	136000	96			Bond, et al., 1960
	<u>CRUSTACEAN</u>					
	<u>Gammarus fasciatus</u>	23000	96			Sanders, 1970
	<u>FENAC</u>					
(SODIUM SALT)	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	12000	96			Sanders, 1969
	<u>Gammarus fasciatus</u>			100,000 ug/1	48 hr.	Sanders, 1970
	<u>Daphnia pulex</u>	4500	48			Sanders and Cope, 1966
	<u>Simocephalus serrulatus</u>	6600	48			Sanders and Cope, 1966
	<u>Daphnia magna</u>			100,000 ug/1	48 hr.	Sanders, 1970
	<u>Cypridopsis vidua</u>			100,000 ug/1	48 hr.	Sanders, 1970
	<u>Asellus brevicaudus</u>			100,000 ug/1	48 hr.	Sanders, 1970
	<u>Palaemonetes kadiakensis</u>			100,000 ug/1	48 hr.	Sanders, 1970
	<u>Orconectes nais</u>					

APPENDIX TABLE 2 (continued)

## HERBICIDES, FUNGICIDES, DEFOLIANTS

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC 50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter</u>	
	<u>INSECT</u>					
	<u>Pteronarcys californica</u>	55000	96			Sanders and Cope, 1968
	<u>FISH</u>					
	<u>Lepomis</u>	15000	96			Hughes and Davis, 1962
HYAMINE 1622	<u>FISHES</u>					
	<u>Pimephales promelas</u>	1600	96			Surber and Pickering, 1962
	<u>Lepomis macrochirus</u>	1400	96			Surber and Pickering, 1962
	<u>Oncorhynchus kisutch</u>	53000	96			Bond, et al., 1960
HYAMINE 2389	<u>FISHES</u>					
	<u>Pimephales promelas</u>	2400	96			Surber and Pickering, 1962
	<u>Lepomis macrochirus</u>	1200	96			Surber and Pickering, 1962
HYDROTHAL 47	<u>CRUSTACEAN</u>					
	<u>Gammarus fasciatus</u>	510	96			Sanders, 1970
HYDROTHAL 191	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	500	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	480	96			Sanders, 1970

APPENDIX TABLE 2 (continued)

## HERBICIDES, FUNGICIDES, DEFOLIANTS

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter</u>	
HYDROTHAL PLUS	<u>FISH</u>					
	<u>Lepomis macrochirus</u>	3500	48			Hughes and Davis, 1964
IPC	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	10000	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	1900	96			Sanders, 1970
	<u>Simocephalus serrulatus</u>	10000	48			Sanders and Cope, 1966
	<u>Daphnia pulex</u>	10000	48			Sanders and Cope, 1966
KURON	<u>CRUSTACEANS</u>					
	<u>Simocephalus serrulatus</u>	2400	48			Sanders and Cope, 1966
	<u>Daphnia pulex</u>	2000	48			Sanders and Cope, 1966
MCDA	<u>FISH</u>					
	<u>Lepomis macrochirus</u>	1500	48			Hughes and Davis, 1964
MOLINATE	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	4500	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	390	48			Sanders, 1970
	<u>Daphnia magna</u>	600	48			Sanders, 1970
	<u>Asellus brevicaudus</u>	400	48			Sanders, 1970
	<u>Palaemonetes kadiakensis</u>	1000	48			Sanders, 1970
	<u>Orconectes nais</u>	5600	48			Sanders, 1970



APPENDIX TABLE 2 (continued)

## HERBICIDES, FUNGICIDES, DEFOLIANTS

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter</u>	
MONURON	<u>FISH</u> <u>Oncorhynchus kisutch</u>	110000	48			Bond, et al., 1960
PARAQUAT	<u>CRUSTACEANS</u> <u>Gammarus lacustris</u> <u>Simocephalus serrulatus</u> <u>Daphnia pulex</u>	110000 4000 3700	96 48 48			Sanders, 1969 Sanders and Cope, 1966 Sanders and Cope, 1966
	<u>INSECT</u> <u>Pteronarcys californica</u>				100,000 ug/l 96 hr.	Sanders and Cope, 1968
PEBULATE	<u>CRUSTACEAN</u> <u>Gammarus fasciatus</u>	10000	96			Sanders, 1970
	<u>INSECT</u> <u>Pteronarcys californica</u>	48000	96			Sanders and Cope, 1968
PROPANIL	<u>CRUSTACEAN</u> <u>Gammarus fasciatus</u>	16000	96			Sanders, 1969
SILVEX (BEE)	<u>CRUSTACEANS</u> <u>Gammarus fasciatus</u> <u>Daphnia magna</u> <u>Cypridopsis vidua</u> <u>Asellus brevicaudus</u> <u>Palaemonetes kadiakensis</u> <u>Orconectes nais</u>	250 2100 4900 40000 8000 60000	96 48 48 48 48 48			Sanders, 1970 Sanders, 1970 Sanders, 1970 Sanders, 1970 Sanders, 1970 Sanders, 1970

APPENDIX TABLE 2 (continued)

## HERBICIDES, FUNGICIDES, DEFOLIANTS

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter</u>	
	<u>FISH</u>					
	<u>Lepomis macrochirus</u>	1200	48			Hughes and Davis, 1963
SILVEX (PGBE)	<u>CRUSTACEANS</u>					
	<u>Gammarus fasciatus</u>	840	96			Sanders, 1970
	<u>Daphnia magna</u>	180	48			Sanders, 1970
	<u>Cypridopsis vidua</u>	200	48			Sanders, 1970
	<u>Asellus brevicaudus</u>	500	48			Sanders, 1970
	<u>Palaemonetes kadiakensis</u>	3200	48			Sanders, 1970
	<u>Orconectes nais</u>				100,000 ug/l 48 hr.	Sanders, 1970
	<u>FISH</u>					
	<u>Lepomis macrochirus</u>	16600	48			Hughes and Davis, 1963
SILVEX (IOE)	<u>FISH</u>					
	<u>Lepomis macrochirus</u>	1400	48			Hughes and Davis, 1963
SILVEX (POTASSIUM SALT)	<u>FISH</u>					
	<u>Lepomis macrochirus</u>	83000	48			Hughes and Davis, 1963
SIMAZINE	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	13000	96			Sanders, 1969
	<u>Gammarus fasciatus</u>				100,000 ug/l 48 hr.	Sanders, 1970
	<u>Daphnia magna</u>	1000	48			
	<u>Cypridopsis vidua</u>	3200	48			
	<u>Asellus brevicaudus</u>				100,000 ug/l 48 hr.	Sanders, 1970
	<u>Palaemonetes kadiakensis</u>				100,000 ug/l 48 hr.	Sanders, 1970
	<u>Orconectes nais</u>				100,000 ug/l 48 hr.	Sanders, 1970
	<u>FISH</u>					
	<u>Oncorhynchus kisutch</u>	6600	48			Bond, et al., 1960

APPENDIX TABLE 2 (continued)

## HERBICIDES, FUNGICIDES, DEFOLIANTS

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter</u>	
TRIFLURALIN	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	2200	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	1000	96			Sanders, 1970
	<u>Daphnia magna</u>	560	48			Sanders, 1970
	<u>Daphnia pulex</u>	240	48			Sanders and Cope, 1966
	<u>Simocephalus serrulatus</u>	450	48			Sanders and Cope, 1966
	<u>Cypridopsis vidua</u>	250	48			Sanders, 1970
	<u>Asellus brevicaudus</u>	200	48			Sanders, 1970
	<u>Palaemonetes kadiakensis</u>	1200	48			Sanders, 1970
	<u>Orconectes nais</u>	50000	48			Sanders, 1970
	<u>INSECT</u>					
	<u>Pteronarcys californica</u>	3000	96			
VERNOLATE	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	1800	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	13000	96			Sanders, 1970
	<u>Daphnia magna</u>	1100	48			Sanders, 1970
	<u>Cypridopsis vidua</u>	240	48			Sanders, 1970
	<u>Asellus brevicaudus</u>	5600	48			Sanders, 1970
	<u>Palaemonetes kadiakensis</u>	1900	48			Sanders, 1970
	<u>Orconectes nais</u>	24000	48			Sanders, 1970

APPENDIX TABLE 2 (continued)

## BOTANICALS

<u>PESTICIDE</u>	<u>ORGANISM</u>	<u>ACUTE TOXICITY</u> <u>LC-50</u>		<u>SUB-ACUTE EFFECTS</u>	<u>NO EFFECT</u>	<u>REFERENCE</u>
		<u>ug/liter</u>	<u>hours</u>	<u>ug/liter</u>	<u>ug/liter</u>	
ALLETHRIN	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	11	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	8	96			Sanders, in press
	<u>Simocephalus serrulatus</u>	56	48			Sanders and Cope, 1966
	<u>Daphnia pulex</u>	21	48			Sanders and Cope, 1966
	<u>INSECT</u>					
	<u>Pteronarcys californica</u>	2.1	96			Sanders and Cope, 1968
	<u>FISHES</u>					
	<u>Lepomis macrochirus</u>	56	96			FPRL
	<u>Salmo gairdneri</u>	19	96			FPRL
PYRETHRUM	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	12	96			Sanders, 1969
	<u>Gammarus fasciatus</u>	11	96			Sanders, 1969
	<u>Simocephalus serrulatus</u>	42	48			Sanders and Cope, 1966
	<u>Daphnia pulex</u>	25	48			Sanders and Cope, 1966
	<u>INSECT</u>					
	<u>Pteronarcys californica</u>	1.0	96			Sanders and Cope, 1968
ROTENONE	<u>CRUSTACEANS</u>					
	<u>Gammarus lacustris</u>	2600	96			Sanders, 1969
	<u>Simocephalus serrulatus</u>	190	48			Sanders and Cope, 1966
	<u>Daphnia pulex</u>	100	48			Sanders and Cope, 1966
	<u>INSECT</u>					
	<u>Pteronarcys californica</u>	380	96			Sanders and Cope, 1968

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