Edwin E. Geldreich U.S. Environmental Protection Agency Risk Reduction Engineering Laboratory Cincinnati, Ohio 45268

INTRODUCTION

The heterotrophic bacterial population in drinking water is composed of many transient organisms that never colonize the distribution system, while other associate organisms are more opportunistic, being capable of surviving on minimal nutrients, attachment to pipe sediments and becoming a participant in the developing biofilm. While these bacteria are generally of no public health significance, some opportunistic colonizers of the pipe network may, in addition, become colonizers of the human body through contact with water supply.

Opportunistic pathogens are generally understood to include those organisms which may exist as part of the normal body microflora but under certain conditions cause disease in compromised hosts. Such organisms become particularly invasive to susceptible individuals (elderly, newborns, AIDS victims, cancer patients receiving chemotherapy, burn cases, dialysis patients, trauma patients, and individuals receiving organ transplants). The route of exposure may be ingestion, inhalation or body contact with water supply during bathing, (whirlpool use, dental equipment, etc.) and indoor air climate control devices (humidification, air cooling). The purpose of this presentation is to place the subject in a more realistic aspect relative to management of water supply quality.

OCCURRENCE

Opportunistic pathogen infections are a serious public health threat anywhere there are large numbers of people in close confinement, such as nurseries, pre-schools, summer camps and in particular, hospitals and senior care facilities. At least 5 percent of patients admitted to hospitals acquire nosocomial infections and about 1 percent of the patients die as a direct result (1). Many of these organisms occur in the diverse heterotrophic flora found in water supplies (2-7). In general, these are the organisms that when found in large enough numbers and in the wrong place at the right time, have the potential to cause an infection. Some examples of nosocomial outbreaks associated with contaminated potable water are shown in Table 1.

Heterotrophic bacterial densities in most municipal water supplies are generally below 100 organisms per mL except at static water locations in buildings where densities are often one or two logs higher because of warm ambient temperatures. By contrast, the infective dose levels for a 50 percent attack rate for an opportunistic pathogen in the heterotrophic population, may range up to 10^{10} cells per dose. While the number of cells required to achieve an infective dose by ingestion may seem unlikely to occur often, the volume of water used to take a shower or bath can easily supply this density during a given exposure period.

^{*}Presented at the AWWA Water Quality Technology Conference, Nov. 12, 1991, Orlando, Florida

By contrast, the density of such opportunistic organisms needed to establish infection in newborn babies, post-operative or immunosuppressed patients, the elderly and infirm is generally lower than for healthy children and adults.

An additional factor to consider is colonization by these organisms in water attachment devices used in hospitals and clinics. Acinetobacter infections have been associated with the use of ventilator spirometers (24), room humidifiers (25, 26) and moisturized Wright respirometers (27). Serratia marcescens infections have been transmitted via medical solutions (28), and peritoneal-dialysis effluents (30). While such water related systems and equipment may be amplifiers of opportunistic pathogens, the source of these organisms may be the water supply, handling of the device, or airborne contaminants, to name a few. The contribution that water supply plays in the problem has been the subject of two studies on water supply associated bacteria and patient illnesses (11, 31). Both studies suggested that water supply organisms are part of the problem but not necessarily the major source of nosocomial infections in the hospital environment.

Water supply systems in large housing projects, highrise office buildings, hotels, and large public buildings exacerbate the problem of deteriorating water quality as a consequence of static water or infrequent water demands. Static water in building plumbing networks is often at warm water temperatures that stimulate bacterial growth in the accumulated sediments. Even new building plumbing networks may present a problem of deteriorating water quality as a result of construction practices that introduce dust, dirt and excessive solder flux in the lines during pipe assembly. Solder flux can be a nutrient source as well as an attachment site for numerous heterotrophic bacteria. This type of a water quality problem was experienced by a Boston hospital after the acceptance of a new building addition to their facility complex (32). Water entering the hospital lines was of high quality (no coliforms per 100 mL and heterotrophic plate counts averaging 3 organisms per mL). As the staff began to phase in the use of the new building addition, there were numerous complaints of malaise by hospital personnel and the water supply became a prime suspect. Laboratory analyses of the water supply in this new facility revealed no detectable coliforms but heterotrophic bacterial densities ranged from 3,000 to 4,000 organisms per mL. Turning on all faucets throughout the new building for a minimum of 15 hours discharge of building water supply was successful in flushing the contaminants from the plumbing system. The bacterial densities decreased to 15 organisms per mL and by the following day averaged 7 heterotrophic bacteria per mL, thereby achieving a water quality similar to that of the municipal water supply. The ill-defined health complaints of the hospital staff declined following this action response, however, incrimination of the microbial quality of the water supply remained circumstantial.

Building water supply lines and their attachment devices have a significant impact on the microbial quality of water. Long stagnation of public water supply in the warm environment of a building water system encourages various heterotrophic bacteria, including <u>Legionella</u>, to colonize pipe joint packing materials, valve stem seals, vacuum breakers (used in back flow prevention) and faucet aerators (34,35).

Hot water tanks in homes and building water systems attachment devices should not be overlooked as a cause of water quality deterioration in home or care centers. If thermostats on hot water tanks are set below 55°C as an energy conservation measure or to prevent scalding of

patients, growth of <u>Legionella</u> may occur in the hot water tank. In such situations legionellae densities may reach infective dose concentrations either in the hot water tank or in an attached shower head (36-41). Most residential hot water tanks are heated from the bottom, near the cold water entrance pipe so the water supply can be quickly heated to above 55°C, however, accumulating sediments at the bottom of the tank provide a heat buffered environment for <u>Legionella</u> colonization. Water in large institutions is often heated by internal steam coils located at mid depth in the tank, thus the cooler water in the bottom may not be heated sufficiently to kill <u>Legionella</u>. Recirculation of the hot water may spread the organisms to all parts of the system.

Cold water storage tanks in highrise buildings must be covered to prevent introduction of contamination from nesting birds and atmospheric dust and to reduce or prevent algae growth. Various heterotrophic bacteria may enter via this route and colonize in the accumulating bottom sediments. Algae may also be introduced by the same routes and proceed to grow in the available light from the open storage tank. Growth in an open water supply tank and subsequent release of algal toxin was the cause of one waterborne outbreak of diarrhea confined to a Chicago apartment building (42).

Increasingly, water utility customers who are dissatisfied with taste, odor or fear the potential health risks alleged to be associated with their municipal water supply are attempting to further refine the water quality at the tap. While treatment devices may be very effective initially in providing aesthetic treatment of the water, their usefulness over time may diminish because of unpredictable service capacity to adsorb a varying mixture of trace contaminants, guality characteristics of water supply and the volume of water processed over time. Poor design of attachment devices may provide recesses that do not drain. Such tiny pools of water become active sites for biofilm development that accelerates in the warm ambient environment and periodically diffuses into the interrupted flows of product water. Another aspect of bacterial proliferation is colonization in devices with carbon filters. While some organisms may pass through the device with little or no retention, others are amplified in these units and bacteria are released at densities higher than those found in the in-coming public water supply. Challenge of carbon filter devices with coliforms, opportunistic organisms and primary pathogens (bacteria anticipated in cross-connections, line breaks or backsiphonage) revealed that such treatment units do not provide an effective barrier. While Escherichia coli, Salmonella and other organisms pass through the filter, other organisms such as <u>Klebsiella</u> pneumoniae, Aeromonas hydrophila and Legionella pneumophila can colonize these devices. As a consequence, devices using carbon filters should not be used on an untreated water supply of questionable quality (43-45).

REPRESENTATIVE ORGANISMS

There is a variety of heterotrophic organisms that can occur in any "safe water supply." They are indeed opportunists in the broad sense, adjusting to a harsh environment and taking advantage of selected sites in the water supply system to colonize. Contacts with breaks in the human body barriers against disease result in a similar pattern of colonization of selected sites that lead to illness if natural defenses prove ineffective. While many heterotrophic organisms in water supply may be capable of colonizing the pipe network, only a few have the potential to be significant opportunistic pathogens. Such is the nature of four candidates among a variety of bacteria, fungi and yeast that are often reported to be waterborne opportunistic pathogens.

ACID-FAST (NONTUBERCULOUS) BACTERIA

Water supply may be significant in the transport of nontuberculous bacteria that pass through treatment barriers in very low densities. Speciation of water supply isolates reveals this group of acid-fast bacteria includes M. fortuitum, M. phei, M. gordonae, M. xenopei, M. kansasii, <u>M. avium</u> and <u>M. chelonae</u> (46-51). The pathological significance of these organisms is that human colonization may occur in the lungs and lymph nodes, or cause skin lesions, septicemia, and cause post-surgery infections. Furthermore, nontuberculous mycobacterial disease is the third most common opportunistic fatal infection in patients with AIDS (46). Waterborne mycobacterial infections present the greatest risk to patients in the hospital setting, particularly those susceptible older individuals that bathe in aerosolized water during the Extensive searches for the cause of two nosocomial summer months. outbreaks of Mycobacterium infection (M. fortuitum and M. gordonae) in different hospitals revealed that these organisms were associated with ice and ice water taken from contaminated ice machines (52, 53).

Nontuberculous <u>Mycobacteria</u> can be isolated from human fecal material (54, 55). These organisms were isolated from 40 percent of stool samples examined from healthy subjects; the mean density was 19 acid-fast organisms per gram of feces (55). Wastes from pig farms also contained <u>Mycobacteria</u> (56) and waste-water effluents were reported to contain an approximate 10^4 organisms per 100 mL (57).

Raw source waters at water supply intakes have been shown to contain acid-fast bacteria (50, 58, 59). Acid-fast bacteria were found in the raw water to the Oakwood, Illinois water treatment plant at a geometric mean density of 271 organisms per liter, while at Decatur, Illinois, raw water densities were approximately one order of magnitude less. Upon passage through treatment, the most significant reductions of acid-fast bacteria occurs during sand filtration. In an 18 month study of these two water systems, reductions in the concentrations of acid-fast bacteria by rapid sand filtration ranged from 59 to 74 percent (60). In the finished water these organisms could be isolated in 36 percent of all one liter samples.

Reported findings of acid-fast bacteria in finished water demonstrate that these organisms are resistant to the usual chlorine disinfectant C·T values applied to inactivate coliforms and viruses (61). Experiments using recent Mycobacterium isolates from chlorinated water supply (M. fortuitum, M. gordonae and M. avium) plus clinical isolates of M. chelonae, M. kansasii and M. intracellulare revealed that chlorine levels of less than 1.0 mg/L may not be adequate for effective inactivation of these opportunistic pathogens (62). Even the presence of a free chelorine residual (<0.2 mg/L) at a low water pH (5.9 to 7.1) did little to reduce acid-fast bacteria in the distribution system. Mycobacterium were also reported to be more resistant than <u>E. coli</u> to inactivation by inorganic chloramines (63) and by ozone (64).

While densities of mycobacteria entering the distribution system may be only a few organisms per liter, this density may change significantly during warm water periods in the static sections of the distribution network. Regrowth may also be intensified in older portions of the pipe network where corrosion is a problem and water pH is elevated to combat corrosivity. The trade-off is less effective disinfectant action of free chlorine at higher pH. Some regrowth was also noted in dead end areas where chlorine residuals disappeared and total organic carbon concentration and turbidity were higher. Devices attached to building plumbing systems may also be amplifiers of mycobacteria. Mycobacteria are among the first organisms to colonize reverse osmosis membranes used in producing reagent grade water, reuse water systems and medical devices. For example, nontuberculous mycobacteria were detected in water from 95 of 115 hemodialysis centers that reused disposable hemodialyzers (artificial kidneys) for the same patient (65). Increased incidence of patient infections caused by acidfast mycobacteria prompted an investigation that concluded water was the source of these opportunistic pathogens.

The water tap may also be a source of mycobacteria with the organisms colonizing the sediment accumulations in the device itself. Apparently, the presence of these organisms can invariably be found in scrapings or swabbings from the cold and hot water taps (66). In a study involving three hospitals, <u>M. xenopei</u> was recovered from 61 of 111 pairs of hot and cold water taps, 20 of 74 tap pairs in another hospital, but from only 3 of 61 pairs of taps in the third hospital. Positive findings were more often reported from the hot water tap, an observation which is not surprising since the optimum growth temperature for acid-fast bacteria is 42 to $44^{\circ}C$.

FECAL KLEBSIELLA

The sanitary significance of the klebsiellae group of coliform bacteria can be perplexing. Most of these organisms are of environmental origin without sanitary significance while other strains of the same genus have their source in the intestinal tract of warm-blooded animals (67-69). The genus includes K. pneumoniae, K. oxytoca, K. ozaenae, K. planticola, K. terrigena and K. rhinoscheromatis. Most of these species have been detected in coliform contaminated public water supplies (70-79). K. pneumoniae and K. oxytoca have often been reported to be the predominant organisms in distribution biofilm occurrences. These occurrences in water supply pose the questions: are these klebsiellae of fecal origin; can they be a potential opportunistic pathogen to susceptible individuals in the community?

In response to the first question, approximately 30 to 40 percent of all warm-blooded animals, humans included, have <u>Klebsiella</u> in their intestinal tracts, with individual densities ranging up to 10⁸ <u>Klebsiella</u> per gram of feces (70-82). An estimated 60-85 percent of all <u>Klebsiella</u> isolated from feces and clinical specimens were positive in the fecal coliform test were identified as K. pneumoniae (83-86).

K. pneumoniae, particularly antibiotic resistant serotypes, can cause human infections of the respiratory system, genito-urinary tract, nose and throat, and occasionally meningitis and septicemia (87, 88). Klebsiella-caused infection is sometimes of apparent primary etiology, but more often is found in mixed infection or as a secondary invader (89). In the hospital environment, the nosocomial infection rate for pathogenic K. pneumoniae was 16.7 infections per 100 patients from 94 hospitals (90). Klebsiella pneumoniae was the cause of 1.1 percent of all nosocomial hospital deaths during the same period. Infections of the urinary system, lower respiratory tract and surgical wounds were the most frequent cause of Klebsiella associated illnesses or deaths. The lack of evidence of increased illness in a community during a coliform biofilm event may relate to difficulties in gathering reports of water related illness cases among susceptible people at home or in the work environment vs. patients in the hospital setting.

Most of the <u>Klebsiella</u> waterborne occurrences are not of fecal origin. In those infrequent situations where the laboratory analyses reveal fecal <u>Klebsiella</u> in the distribution system, there should be a high priority effort to destroy the colonization sites because of the concern for more frequent releases of this opportunistic pathogen at higher densities into the water supply. Infective dose (ID_{50}) values for environmental and clinical isolates of <u>Klebsiella</u> have been reported to be between 3.5 x 10^1 to 7.9 x 10^5 cells per mL (91). Therefore, ingestion of 100 mL of drinking water (approximately one glass of water) containing 3.5 x 10^1 <u>Klebsiella</u> per mL could present a risk to susceptible individuals. Inhalation of moisture from vaporizers using drinking water contaminated with Klebsiella should also be considered a risk to some individuals.

LEGIONELLA

Legionella pneumophila is an important waterborne opportunistic pathogen that causes Legionnaire's disease in susceptible individuals exposed to contaminated aerosols from shower baths and air conditioner heat exchanges. The respiratory disease results in a complex colonization of the body that is responsible for pneumonia with significant mortality rates among senior citizens. Pontiac fever, another illness caused by legionellae, is a non-pneumonic, non-fatal and self-limiting disease. Apparently, there is no human carrier state or reservoir for legionellae bacteria in warm-blooded animals.

While this group of small, gram-negative bacteria have an absolute nutritional requirement for L-cysteine (92) it is somewhat surprising to find legionellae widespread in the aquatic environment. These organisms have been detected in freshwater streams and lakes in both North America and Europe, plus the tropical waters of Puerto Rico (93-95). In one study of 793 water samples collected from 67 different lakes and rivers throughout the United States, virtually all sources were positive for <u>L. pneumophila</u>, using the direct fluorescent antibody technique for detection (96). There is some indication that legionellae are very infrequently found in groundwater, unless there is some surface water runoff seepage or poor soil barrier protection (97,98).

Water treatment processes may play some role in the development of an ecological niche for <u>Legionella</u> through the release of assimilable organic nutrients, particulates and various heterotrophic organisms into the distribution system as a result of uneven, interrupted or failed treatment processes (99-104). Among the microorganisms in the raw source water that sequester <u>Legionella</u> and provide safe passage through the disinfection process are algae, amoebae and ciliates (105, 106). Airborne legionellae in dust or particulate laden rain showers may find their way to the open air treatment basins. Common pathways for their entry into the distribution system include reservoir air vents, main construction, pipe line repairs, cross-connections and back-siphonage (107).

Establishment of <u>Legionella</u> in the distribution system is most likely to occur in biofilm locations where symbiotic relationships with other heterotrophic bacteria (<u>Flavobacterium breve</u>, <u>Pseudomonas</u>, <u>Alcaligenes</u>, or <u>Acinetobacter</u>) provide the critical nutrient requirements necessary for long term persistence of this opportunistic pathogen (108-110). Many of these sites will be found at the periphery of the system (long pipe runs into dead ends) and in little used service connections throughout the pipe network where the water can stagnate. Densities of <u>Legionella</u> may be only a few cells per liter in water supply (105-106) and the mere presence of these few Legionella in drinking water does not pose a direct health threat until there are opportunities for amplification (hot water tanks, shower heads, water evaporator cooling devices, etc.).

Efforts to eliminate low levels of these organisms (a few per liter) in water supply treatment processes and in the distribution system network are not cost-effective. Water utility operations, however, can minimize regrowth potential through good housekeeping practices that include removal of scums and biofilm accumulations at air-water interfaces in treatment basins, connecting flumes and attachments to agitator paddles in flocculation basins. System-wide flushing with particular emphasis on dead end sections during warm water periods, will significantly suppress further development of biofilm and introduce detectable disinfectant residuals to those areas where <u>Legionella</u> and other heterotrophic bacteria may persist. The net effect desired is to suppress microbial symbiotic relationshies that are essential to Legionella metabolism.

PSEUDOMONAS AERUGINOSA

<u>Pseudomonas</u> species are ubiquitous bacteria that are able to flourish in a wide variety of habitats (surface waters, aquifers, sea water, soils and vegetation). Some pseudomonads are among the prominent denitrifiers while others grow prodigiously in and on tertiary treatment devices such as reverse osmosis and electrodialysis membranes and in sand or carbon filtration beds. Pseudomonads reported in some drinking water supplies include: <u>P. aeruginosa</u>, <u>P. cepacia</u>, <u>P. fluorescens</u>, <u>P. mallei</u>, <u>P. maltophilia</u>, <u>P. putida</u> and <u>P. testosteroni</u> (111,112). Perhaps the most significant species of concern in drinking water is <u>P. aeruginosa</u>. To this list can be added: <u>P. stutzeri</u>, <u>P. diminuta</u> and <u>P. acidovoran</u> which have been found in bottled waters at densities ranging from 10³ to 10⁵ organisms per mL (113-115). These organisms metabolically adapt to survive on minimal nutrient concentrations typical of protected aquifers and treated drinking water.

The ability of <u>P. aeruginosa</u> to rapidly colonize a variety of environments, including the susceptible human, makes it a major opportunistic pathogen; particularly <u>P. aeruginosa</u> serogroups 11 and possibly serogroup 9 which are the most frequently isolated pathogenic strains. Bacteremia attributable to <u>Pseudomonas</u> has become a major concern in the management of trauma as well as in the management of susceptible patients recovering from burns, intensive surgery and others exposed to cancer therapy (116-120). Other serious infections for susceptible individuals involve eye, ear, nose, and occasionally the gastrointestinal tract (121, 122).

The infrequent occurrence (3-19 percent) of <u>Pseudomonas</u> in the human intestinal tract (123) suggests that colonization of the gastrointestinal system rarely occurs in healthy adults, indicating that there are potent host-defense mechanisms against this group of gram negative bacteria (124, 125). Since municipal sewage contains a mixture of domestic wastes, industrial discharges and intermittent stormwater runoff, it is not unexpected to find <u>P. aeruginosa</u> in 90% of sewage samples (126). Densities of <u>P. aeruginosa</u> in surface waters receiving waste and stormwater discharges may range from 1 x 10° to 1 x 10⁴ cells per 100 mL, and are influenced by available nutrients and seasonal water temperatures (127).

P. aeruginosa found in a contaminated water supply has been linked to one waterborne outbreak that occurred in a newborn nursery (128). In this case study, the ground water supply was a starinated by seepage of sewage

and infiltration of contaminated surface water. Since <u>P. aeruginosa</u> is the most prevalent <u>Pseudomonas</u> in human disease (129) its occurrence has been limited to less than one organism in 250 mL of bottled drinking water by the European community. Other <u>Pseudomonas</u> species found occasionally in water (<u>P. fluorescens</u>, <u>P. putida</u>, <u>P. multivorans</u>, <u>P. maltophilia</u> and <u>P. stutzeri</u>) have not yet been linked to waterborne outbreaks suggesting they are indigenous aquatic bacteria in every water environment.

JOINT RESPONSIBILITY FOR WATER QUALITY

Fulfilling the obligation for the production, delivery, and maintenance of high quality water supply to the consumer is the joint responsibility of the water utility and the user community of hospitals, highrise building complex management and the individual family. Water quality is created at the water plant and is a reflection of treatment operations and distribution system maintenance and management. While treatment technology will provide a water free of health risks associated with primary pathogens, treatment processes were never intended to produce a sterile water supply. Opportunistic organisms will pass through or circumvent treatment barriers as now defined. Some of these pathways include passage of dust contaminants into open air process basins, organism protection in clumping, viable cell transport by aquatic invertebrates, movement with carbon fines, or unsettled particulates and infiltration through fractured pipes, line breaks and line repair practices.

WATER UTILITIES

Opportunistic organisms are very adaptable in establishment of a biofilm colonization that promotes the amplification of cell densities to levels that may be several logs higher than the initial levels. In treatment basins, colonization occurs at the solid surface-water interfaces of process basins and connecting flumes and agitator paddles in the coagulation basin. This problem is best controlled by scheduling application of high pressure washing of compartment walls and mechanical scraping of paddle surfaces. Colonization in the distribution system may occur in the slow flow and dead end sections of the pipe network, and on the walls and in the accumulated sediments of water storage tanks. Static water locations in the pipe system and stratified water in storage tanks promotes colonization by a variety of bacteria during warm water periods.

The key to suppressing colonization is to keep the water moving throughout the system and to remove accumulating pipe sediments. Flushing that is done at least every Spring in a systematic fashion from water plant to end of the pipe network often contributes to control of biofilm incursions. Draining and cleaning of all water storage tanks and standpipes may be more difficult to manage but nevertheless should be done as frequently as possible to suppress biofilm growth at these sites. For water utilities that use chloramination as the post-disinfectant, it may be desirable to change to free chlorine for a two week period each year to effectively reduce chloramine resistant heterotrophic bacteria population in biofilms.

HOSPITALS

Hospitals also have a responsibility in the management of water supply quality. The entire pipe network needs to be flushed every six months because this water is always in a warm environment regardless of seasonal weather changes. Flushing in this case needs to be done at each faucet throughout the facility for a 15 minute period or until a measurable disinfectant residual is obtained. Attachment devices must also be disassembled and cleaned of incrustations and sediments before reassembly. Flexible hose connections should be replaced at this time as well as any gaskets and washers that have visible slime growth or incrustations. Prior to the activation of a new hospital wing or reactivation of closed patient wards or operating theaters, all faucets should be opened for 24 hours to flush out the stagnate water and sediments in an effort to achieve water supply that is representative of public water supply quality.

BUILDING MANAGERS

Highrise buildings also have complex water supply networks that are subject to quality changes. Water supply tanks in apartment buildings, office buildings and hotels should be flushed and cleaned each year. Unoccupied apartments, office space and hotel rooms vacant for 3 months or more may have a significant deterioration in the water quality in the static service lines. Flushing overnight from each water tap in the vacant rooms will restore water quality which is characteristic of the building supply.

INDIVIDUAL CONSUMERS

Consumers must also be aware of their responsibility to protect the public water supply in their home. Perhaps the most common problem with static water quality is in the first draw of water supply in the morning. Microbial growth will occur in the water during overnight periods of no flow due to warm ambient temperatures associated with proximity to furnace pipes, hot water lines and under the sink locations. As a general practice, it is a good habit to flush the tap water line for 30 seconds each morning before ingesting that first glass of water. If the family has been away on an extended vacation, again flushing water lines from the bathroom and kitchen taps for several minutes will do much to remove higher bacterial densities in static water lines. For families that attach point-of-use devices to the water supply line for additional treatment, there should be a scheduled effort to change carbon filters every 4 to 6 weeks, depending on usage, or as recommended by the manufacturer. Morning flush of these devices is very important because of the microbial build-up in the unit overnight that often exceeds what occurs in static water at other home faucets.

SUMMARY

Organisms that become established in water supply may also be opportunistic pathogens. Representative opportunistic pathogens that are waterborne include acid-fast bacteria, fecal klebsiellae, <u>Legionella</u> and <u>Pseudomonas</u> <u>aeruginosa</u>. These organisms may be found in the heterotrophic bacterial population of treated drinking water and if appropriate conditions exist, may colonize and become part of the biofilm.

Maintaining a high quality water supply requires careful treatment and a clean water distribution system. Users of the community water supply also have a responsibility to preserve this water quality from deterioration as it leaves the service meter and traverses the building supply lines. The goal is to minimize exposure to various heterotrophic bacteria that may pose a risk to those consumers of varying health status in the community of people.

Etiologic Agent	Illness	Reference
Pseudomonas	Wound	Cross, et al. (9)
	Wound	Bassett, et al. (10)
	Dermatitis	Highsmith, et al. (11)
	Meningitis	Ho, et al. (12)
	Respiratory	duMoulin, et al. (13)
	Respiratory	Saepan, et al. (14)
	Cellulitis	McGuekin, et al. (15)
Acinetobacter	Peritonitis	Abrutyn, et al. (16)
Mycobacterium	Septicemia	Carson, et al. (17)
	Bacteremia	Bolan, et al. (18)
	Peritonitis	duMoulin and Stottmeir (19)
Flavobacterium	Septicemia	Herman and Himmelsbach (20)
	Respiratory	duMoulin, (21)
Legionella	Respiratory	Cordes, et al. (22)
Klebsiella	Urinary, Respiratory	Kelly, et al. (23)
*Information adapte	ad from Wighemith at al	(9)

Table 1. Documented Nosocomial Outbreaks Associated With Contaminated Potable Water*

_

.

*Information adapted from Highsmith, et al. (8)

-

REFERENCES

- Hughes, J.M. and W.R. Jarvis. Epidemiology of Nosocomial Infections. In: <u>Manual of Clinical Microbiology</u>, 4th edition, eds E. H. Lennette, A. Balows, W.J. Hausler, Jr., and H.J. Shadomy. Amer. Soc. for Microbiol., Washington, D.C. (1985).
- Olson, B.H. and L. Hanami. Seasonal Variation of Bacterial Populations in Water Distribution Systems. pp 137-151. Proc. Amer. Water Works Assoc., Water Quality Technol. Confr., Miami Beach, FL. (1980).
- Lamka, K.G., M.W. LeChevallier and R.J. Seidler. Bacterial Contamination of Drinking Water Supplies in a Modern Rural Neighborhood. Appl. Environ. Microbiol. 39:734-738 (1980).
- Nash, H.D. and E.E. Geldreich. Effect on Storage on Coliform Detection in Potable Water Samples, Water Quality Technol. Confr. Proc. 123-136 (1980).
- Reilly, J.K. and J.S. Kippin. Relationship of Bacterial Counts with Turbidity and Free Chlorine in Two Distribution Systems. Jour. Amer. Water Works Assoc. 75:309-312 (1983).
- 6. Bateman, J.L., R.P. Tu, M.H. Strampher and B.A. Cunha. <u>Aeromonas</u> <u>hydrophila</u> cellulis and Wound Infection Caused by Waterborne Organisms. Heart Lung 17:99-102 (1988).
- Notermans, S., A. Havelaar, W. Jansen, S. Kozaki and P. Guinee. Production of "Asao Toxin" by <u>Aeromonas</u> Strains Isolated from Feces and Drinking Water. Jour. Clin. Microbiol. <u>23</u>:1140-1142. (1986).
- Highsmith, A.K., T.G. Emori, S.M. Aguero, et al. Heterotrophic Bacteria Isolated From Hospital Water Systems. In: <u>International</u> <u>Symposium on Water-Related Health Issues</u>. C.L. Tate, Jr. ed., 181-187. Amer. Water Resources Assoc., Bethesda, MD. (1986).
- Cross, D., A. Benchimol and E. Dimond. Faucet Aerator -- A Source of <u>Pseudomonas</u> Infection. N. Eng. Jour. Med. <u>274</u>:1430-1431 (1966).
- 10. Bassett, D.C.J., K.J. Stokes and W.R.G. Thomas. Wound Infections with <u>Pseudomonas multivorans</u>: Waterborne Contaminant of Disinfectant Solutions. Lancet 1:1188-1191 (1970).
- Highsmith, A.K., P.N. Le, R.F. Khabbag and V.P. Mann. Characteristics of <u>Pseudomonas aeruginosa</u> Isolated From Whirlpools and Bathers. Infection Control, <u>6</u>:407-412 (1985).
- Ho, J.L., A.K. Highsmith, E.S. Wong, et al. Common-Source <u>Pseudomonas</u> <u>aeruginosa</u> Infection in Neurosurgery. Proc. Ann. <u>Meeting. Am. Soc. Microbiol.</u>, Dallas, Texas, Ll0, p. 80 (1981).
- duMoulin, G., G. Doyle, J. Mackay and J. Hedley-Whyte. Bacterial Fouling of a Hospital Closed-Loop Cooling System by Pseudomonas sp. Jour. Clin. Microbiol. <u>13</u>:1060-1065.
- 14. Saepan, M.S., H.O. Bodman, R.B. Kundsin. Microorganisms in Heated Nebulizers. Health Lab. Sci. <u>12</u>:316-320 (1975).

- McGuekin, M.B., R.J. Thorpe and E. Abrutyn. Hydrotherapy: An Outbreak of <u>Pseudomonas</u> <u>aeruginosa</u> Wound Infections Related to Hubbard Tank Treatments. Arch. Phys. Med. Rehabil. <u>62</u>:283-285 (1981).
- 16. Abrutyn, G.A., B.J. Collins, J.R. Babb et al. <u>Pseudomonas</u> aeruginosa in Hospital Sinks. Lancet 1:578-580 (1974).
- 17. Carson, L., G. Bolan, N.J. Petersen, et al. Antimicrobial and Formaldehyde Resistance Patterns of Non-Tuberculous Mycobacteria Associated With Reprocessed Hemodialyzers. Proc. Ann. Meet. Intersci. Conf. Antimicrobial Agents and Chemotherapy, Amer. Soc. Microbiol. Washington, D.C. (1983).
- Bolan, G., A.L. Reingold, L.A. Carson, et al. Infections With Mycobacterium chelonei in Patients Receiving Dialysis and Using Processed Hemodialyzers. Jour. Infect. Dis. <u>154</u>:1013-1019 (1985).
- 19. duMoulin, G.C., and K.D. Stottmeir. Waterborne Mycobacteria: An Increased Threat of Health. ASM News, <u>52</u>:525-529 (1986).
- Herman, L. and C. Himmelsbach. Detection and Control of Hospital Sources of Flavobacteria. Hospitals 39:72-76 (1965).
- duMoulin, G.C. Airway Colonization by <u>Flavobacterium</u> in an Intensive Care Unit. Jour. Clin. Microbiol. 10:155-160 (1979).
- 22. Cordes, L., et al. Isolation of Legionella pneumophila from Hospital Showerheads. Annals. Internal. Med. 94:195-197 (1981).
- Kelly, M.T., D.J. Brenner, and JaJ. Farmer, III. Enterobacteriaceae. In: Manual of Clinical Microbiology, 4th ed. Eds. E.H. Lennette, A. Ballows, W.J. Hausler, Jr., and H.J. Shadomy. Amer. Soc. Microbiol., Washington, D.C. (1985).
- 24. Irwin, R.S., R.R. Demers, M.R. Pratter, et al. An Outbreak of <u>Acinetobacter Infection Associated with the Use of a Ventilator</u> Spirometer. Respir. Care 25:232-237 (1980).
- Gervich, D.H. and L.S. Grout. An Outbreak of Nosocomial <u>Acinetobacter Infection from Humidifiers</u>. Amer. Jour. Infect. Contr. 13:210-215 (1985).
- Smith, P.W. and R.M. Massanari. Room Humidifiers as the Source of <u>Acinetobacter</u> Infections. Jour. Amer. Med. Assoc. 237:795-797 (1997).
- Chunha, B.A., J.J. Klimek, J. Graceuski and J.C. McLoughlin. A Common Source Outbreak of <u>Acinetobacter</u> Pulmonary Infections Traced to Wright Respirometers. Post Grad. Med. Jour. <u>56</u>:169-172 (1980).
- Nakashima, A.K., M.A. McCarthy, W.J. Martone and L. Anderson. Epidemic Septic Arthritis Caused by <u>Serratia marcescens</u> and Associated with a Benzalkonium Chloride Antiseptic. Jour. Clin. Microbiol. 25:1014-1018 (1987).

834

- McCormack, R.C. and C.M. Kunin. Control of a Single Source Nursery Epidemic due to <u>Serratia marcescens</u>. Pediatrics <u>37</u>:750-755 (1966).
- 30. Connacher, L.A., D.C. Old, G. Phillips, et al. Recurrent Peritonitis Caused by <u>Serratia marcescens</u> in a Diabetic Patient Receiving Continuous Ambulatory Peritoneal Dialysis. Jour. Hosp. Infect. <u>11</u>:155-160 (1988).
- 31. Lee, Y-L., L. Thrupp, C. Richards, et al. Water Supply As a Potential Source of Opportunistic Pathogens Causing Nosocomial Infections. Appl. Environ. Microbiol. (In Press) (1990).
- 32. Eichhorn, J.H., M.L. Bancroft, L.H. Laasberg, G.C. duMoulin and A.J. Saubermann. Contamination of Medical Gas and Water Pipelines in a New Hospital Building. Anesthesiology, <u>46</u>:286-289 (1977).
- 34. Colbourne, J.S., M.G. Smith, S.P. Fisher-Hock, and D. Harper. Source of <u>Legionella pneumophila</u> Infection in a Hospital Hot Water System: Materials Used in Water Fittings Capable of Supporting <u>L. pneumophila</u> Growth. pp. 305-307. In: <u>Legionella</u> Proceedings of the 2nd International Symposium, eds. C. Thornsberry, A. Balows, J.C. Feeley and W. Jakubowski. Amer. Soc. Microbiol. Washington, D.C. (1984).
- 35. Ciesielski, C.A., M.J. Blason, F.M. LaForce and W.L.L. Wang. Role of Stagnation and Obstruction of Water Flow in Isolation of Legionella pneumophila from Hospital Plumbing. pp. 307-309. In: Legionella Proceedings of the 2nd International Symposium, eds. C. Thornsberry, A. Balows, J.C. Feeley and W. Jakubowski. Amer. Soc. Microbiol., Washington, D.C. (1984).
- 36. Barnstein, N., C. Vieilly, M. Nowicki, J.C. Paucod and J. Fleurette. Epidemiological Evidence of Legionellosis Transmission Through Domestic Hot Water Supply Systems and Possibilities of Control. Israel Jour. Med. Sci. <u>22</u>:655-661 (1986).
- Wadowsky, R.M., R.B. Yee, and L. Megmar. Hot Water Systems as Source of Legionella pneumophila in Hospital and Non-Hospital Plumbing Fixtures. Appl. Envir. Microbiol. <u>43</u>:1104-1110 (1982).
- Plouffe, J.F., L.R. Webster, and B. Hackman. Relationship Between Colonization of Hospital Buildings with <u>Legionella</u> <u>pneumophila</u> and Hot Water Temperatures. Appl. Envir. Microbiol., 46:769-770 (1983).
- 39. Arnow, P.M., D. Weil and M.F. Para. Prevalence and Significance of <u>Legionella pneumophila</u> and Hot Tap Water Systems. Jour. Infect. Dis. <u>152</u>:145-151 (1985).
- Groothius, D.G., H.R. Veenendool, and H.L. Dijkstra. Influence of Temperature on the Number of Legionella pneumophila in Hot Water Systems. J. Appl. Bacteriol., <u>59</u>:529-536 (1985).
- Botzenhart, K., W. Heizmann, S. Sedaghat, P. Huy and T. Hahn. Bacterial Colonization and Occurrence of <u>Legionella pneumophila</u> in warm and cold water, in faucet aerators, and in drains of hospitals. Zbl. Bakt. Hyg. B, 183:79-85 (1986).

- Epidemiologic Notes and Reports. Outbreaks of Diarrheal Illness Associated with Cyanobacteria (Blue-Green Algae)-Like Bodies -Chicago and Nepal, 1989 and 1990. MMWR, <u>40</u>:325-327 (1990).
- Geldreich, E.E., R.H. Taylor, J.C. Blannon, and D.J. Reasoner. Bacterial Colonization of Point-of-Use Water Treatment Devices. Jour. Amer. Water Works Assoc., <u>77</u>:72-80 (1985).
- Reasoner, D.J., J.C. Blannon and E.E. Geldreich. Microbiological Characteristics of Third-Faucet Point-of-Use Devices. Jour. Amer. Water Works Assoc., 79:60-66 (1987).
- Geldreich, E.E. and D.J. Reasoner. Home Treatment Devices and Water Quality. In: <u>Drinking Water Microbiology</u>, G.A. McFeters ed., Springer-Verlag, New York (1990).
- Good, R.C. Opportunistic Pathogens in the Genus Mycobacterium. Annual Rev. Microbiol. 39:347-369 (1985).
- duMoulin, G.C., I.H. Sherman, D.C. Hoaglin, and K.D. Stottmeier. <u>Mycobacterium avium Complex</u>, an Emerging Pathogen in Massachusetts. Jour. Clin. Microbiol. 22:9-12 (1985).
- Ganzadharam, P.R.J., J.A. Lockhart, R.J. Awe, and D.E. Jenkins. Mycobacterial Contamination Through Tap Water. Amer. Rev. Respiratory Dis. <u>113</u>:894 (1976).
- McSwiggan, D.A. and C.H. Collins. The Isolation of <u>M. kansasii</u> and M. xenopei from Water Systems. Tubercle <u>55</u>:291-297 (1974).
- 50. Goslee, S. and E. Wolinsky. Water as a Source of Potentially Pathogenic Mycobacteria. Amer. Review of Respiratory Dis. 113:287-292 (1976).
- 51. Bullin, C.H., E.I. Tanner and C.H. Collins. Isolation of <u>Mycobacterium xenopei</u> from Water Taps. Jour. Hyg. Camb. <u>68</u>:97-100 (1970).
- Panwalker, A.P. and E. Fuhse. Nosocomial <u>Mycobacterium gordonae</u> Pseudo-Infections from Contaminated Ice Machines. Infect. Contr. 7:67-70 (1986).
- 53. Laussueq, S. A.L. Baltch, R.P. Smith, R.W. Smithwick, B.J. Davis, et al. Nosocomial <u>Mycobacterium</u> fortuitum Colonization from a Contaminated Ice Machine. Amer. Rev. Respir. Dis. 138:891-894 (1988).
- 54. Rosebury, T. <u>Microorganisms Indigenous to Man</u>, McGraw-Hill Book Co., New York (1962).
- 55. Engelbrecht, R.S., D.F. Foster, E.O. Grenning and S.H. Lee. New Microbial Indicators of Wastewater Chlorination Efficiency. EPA-670/2-72-082. U.S. Environmental Protection Agency, Cincinnati, OH. (1974).
- 56. Jones, P.W., J. Bew, M.R. Burrows, P.R. J. Matthews and P. Collins. The Occurrence of <u>Salmonella</u>, <u>Mycobacteria</u> and Pathogenic Strains of <u>E. coli</u> in Pig Slurry. Jour. Hyg. <u>77</u>:43-50 (1976).

- 57. Engelbrecht, R.S. and C.N. Haas. Acid-fast Bacteria and Yeasts as Disinfection Indicators: Enumeration Methodology pp. 2B-1 to 2B-20. Proc. Amer. Water Works Assoc., Water Quality Technol. Confr., Kansas City, MO. (1977).
- 58. Engelbrecht, R.S., B.F. Severin, M.T. Masarek, S. Faroog, S.H. Lee, C.M. Haas and A. Lalchandani. New Microbial Indicators of Disinfection Efficiency. EPA-600/2-77-052, U.S. Environmental Protection Agency, Cincinnati, OH. (1977).
- 59. Shular, J.A. The Occurrence of Indicator Organisms in the Decatur, Illinois South Water Treatment Plant and in One Branch of the Distribution System. M.S. Special Problem Report. Dept. Civil Engr., Univ. IL., Urbana-Champaign. (1978).
- 60. Haas, C., M.A. Meyer and M.S. Paller. The Ecology of Acid-fast Organisms in Water Supply, Treatment, and Distribution Systems. Jour. Amer. Water Works Assoc., <u>75</u>:139-144 (1983).
- 61. Surucu, F. and C.N. Haas. Inactivation of New Indicator Organisms of Disinfection Efficiency. Part I. Free Available Chlorine Species Kinetics. Presented at the 96th Annual Meeting, Amer. Water Works Assoc., New Orleans (1976).
- Pelletier, P.A., G.C. duMoulin and K.D. Stlottmeir. Mycobacteria in Public Water Supplies: Comparative Resistance to Chlorine. Microbiological Sci. <u>5</u>:147-148 (1988).
- 63. Severin, B.F. Inactivation of New Indicator Organisms of Disinfection Efficiency. Part II. Combined Chlorine as Chloramines. Presented at the 96th Annual Meeting, Amer. Water Works Assoc., New Orleans (1976).
- 64. Farooq, S. Kinetics of Inactivation of Yeasts and Acid-Fast Organisms with Ozone. PhD Thesis, Univ. Illinois, Dept. Civil Engr., Urbana-Champaign (1976).
- 65. Carson, L.H., L.A. Bland, L.B. Cusick, M.S. Favero, G.A. Bolan, A.L. Reingold and R.C. Good. Prevalence of Nontuberculous Mycobacteria in Water Supplies of Hemodialysis Centers. Appl. Environ. Microbiol. 54:3122-3125 (1988).
- 66. Bullin, C.H., E.I. Tanner and C.H. Collins. Isolation of Mycobacterium from Water Taps. Jour. Hyg. Camb., <u>68</u>:97-100 (1970).
- 67. Bagley, S.T. Habitat Association of <u>Klebsiella</u> Species. Infection Control <u>6</u>:52-58 (1985).
- Geldreich, E.E. and E. W. Rice. Occurrence, Significance, and Detection of <u>Klebsiella</u> in Water Systems. Jour. Amer. Water Works Assoc. <u>79</u>:74-80 (1987).
- 69. Nunez, W.J. and A.R. Colmer. Differentiation of <u>Aerobacter-Klebsiella</u> Isolated From Sugarcane. Appl. Microbiol., <u>16</u>:1375-1878 (1968).
- 70. Geldreich, E.E., H.D. Nash and D.F. Spino. Characterizing Bacterial Populations in Treated Water Supplies: A Progress Report. Proc. Amer. Water Works Assoc. Water Qual. Technol. Confr. Kansas City, MO. (1977).

- LeChevallier, M.W., R.J. Seidler and T.M. Evans. Enumeration and Characterization of Standard Plate Count Bacteria in Chlorinated and Raw Water Supplies. Appl. Envir. Microbiol. 40:522-930 (1980).
- Herson, D.S. and H. Victoreen. Identification of Coliform Antagonists. Proc. Amer. Water Works Assoc. Water Qual. Technol. Confr., Miami Beach, FL. (1980).
- Reilly, J.K. and J. Kippin. Relationship of Bacterial Counts with Turbidity and Free Chlorine in Two Distribution Systems. Jour. Amer. Water Works Assoc. 75:309-312 (1983).
- 74. Clark, J.A., C.A. Burger, and L.E. Sabatinos. Characterization of Indicator Bacteria in Municipal Raw Water, Drinking Water, and New Main Water Samples. Can. Jour. Microbiol., <u>28</u>:1002-1013 (1982).
- 75. Ptak, O.J., W. Ginsburg and B.F. Wiley. Identification and Incidence of <u>Klebsiella</u> in Chlorinated Water Supplies. Jour. Amer. Water Works Assoc., 65:604-608 (1973).
- 76. Olson, B.H. and L. Hanami. Seasonal Variations of Bacterial Populations in Water Distribution Systems. Proc. Amer. Water Works Assoc., Water Quality Technol. Confr., Miami Beach, FL. (1980).
- 77. Bagley, S.T., R.J. Seidler, and D.J. Brenner. <u>Klebsiella</u> <u>planticola</u> sp. nov.: A New Species of Enterobacteriaceae Found Primarily in Non-clinical Environments. Current Microbiol., <u>6</u>:105-109 (1981).
- Camper, A.K. et al. Bacteria Associated with Granular Activated Carbon Particles in Drinking Water. Appl. Envir. Microbiol. <u>52</u>:434-438 (1986).
- Edberg, S.C., V. Piscitelli and M. Carter. Phenotype Characteristics of Coliform and Noncoliform Bacteria From a Public Water Supply Compared with Regional and National Clinical Species. Appl. Environ. Microbiol. <u>52</u>:474-478. (1986).
- 80. Thom, B.T. <u>Klebsiella</u> in Faeces. Lancet, <u>2</u>:1033. (1970).
- 81. Davis, T.J. and J.M. Matsen. Prevalence and Characteristics of <u>Klebsiella</u> Species. Relation to Association with a Hospital Environment. Jour. Infect. Dis., <u>130</u>:402-405. (1974).
- Cooke, E.M. et al. Further Studies on the Sources of <u>Klebsiella</u> aerogenes in Hospital Patients. Jour. Hygiene <u>83</u>:391-395 (1979).
- Bagley, S.T. and R.J. Seidler. Significance of Fecal Coliform Positive <u>Klebsiella</u>. Appl. Envir. Microbiol. <u>33</u>:1141-1148 (1977).
- Edmondson, A.E., E.M. Cooke, A.P.D. Wilcock and R. Shinebaum. A Comparison of the Properties of <u>Klebsiella</u> Strains Isolated from Different Sources. Jour. Med. Microbiol., <u>13</u>:541-550 (1980).
- 85. Naemura, L.G. and R.J. Seidler. Significance of Low Temperature Growth Associated with the Fecal Coliform Response, Indole

Production and Pectin (sic) Liquefaction in <u>Klebsiella</u>. Appl. Environ. Microbiol. <u>35</u>:392-396 (1978).

- 86. Naemura, L.G., S.T. Bagley, R.J. Siedler, J.B. Kaper and R.R. Colwell. Numerical Taxonomy of <u>Klebsiella pneumoniae</u> Strains Isolated from Clinical and Nonclinical Sources. Current Microbiol. 2:175-780 (1979).
- Montgomerie, J.Z. Epidemiology of <u>Klebsiella</u> and Hospitalassociated Infections. Rev. Infect. Dis. 1:736-753 (1979).
- 88. Smith, S.M., J.T. Digori and R.H.K. Eng. Epidemiology of <u>Klebsiella</u> Antibiotic Resistance and Serotypes. Jour. Clin. Microbiol. <u>16</u>:868-873 (1982).
- Martin, W.J., P.K.W. Yu and T.A. Washington. Epidemiological. Significance of <u>Klebsiella pneumoniae</u>: A 3 Month Study. Mayo Clin. Proc. 46:785-783 (1971).
- 90. Jarvis, W.R. et al. The Epidemiology of Nosocomial Infections Caused by <u>Klebsiella pneumoniae</u>. Infection Control, <u>6</u>:68-74 (1985).
- 91. Bagley, S.T. and R.J. Seidler. Comparative Pathogenicity of Environmental and Clinical <u>Klebsiella</u>. Health Lab. Sci. <u>15</u>:104-111 (1978).
- 92. Lennette, E.H., A. Balows, W.J. Hausler, Jr. and H.J. Shadomy. Manual of Clinical Microbiology. 4th edition. Amer. Soc. Microbiol., Washington, D.C. (1985).
- Fliermans, C.B., et al. Ecological Distribution of <u>Legionella</u> <u>pneumophila</u> from Non-Epidemic Related Aquatic Habitats. Appl. Environ. Microbiol., <u>37</u>:1239-1242 (1979).
- 94. Morris, G.K., et al. Isolation of the Legionnaires Disease Bacterium from Environmental Samples. Ann. Intern. Med., <u>90:664-666</u> (1979).
- 95. Oritz-Roque, C. and T.C. Hazen. Isolation of <u>Legionella</u> <u>pneumophila</u> from Tropical Waters. Ann. Mtg. Amer. Soc. Microbiol. (Abstract) (1981).
- 96. Fliermans, C.B., W.B. Cherry, L.H. Orrison, S.J. Smith, D.L. Tison and D.H. Pope. Ecological Distribution of <u>Legionella</u> gneumophila. Appl. Envir. Microbiol., <u>41</u>:9-16 (1981).
- 97711ermans, C.B., G.E. Bettinger and A.W. Fynsk. Treatment of Cooling Systems Containing High Levels of <u>Legionella</u> pneumophila. Water Res. 16:903-909 (1982).
- 98. Spino, D.F., E.W. Rice and E.E. Geldreich. Occurrence of <u>Legionella</u> spp. and Other Aquatic Bacteria in Chemically Contaminated Ground Water Treated by Aeration. pp. 318-320. In: C. Thornsberry, A. Balows, J.C. Feeley and W. Jakubowski, eds. <u>Legionella: Proceedings of the 2nd International Symposium</u>, June 19-23, 1983; Atlanta, GA. American Society for Microbiology, Washington, D.C.

- 99. Tison, D.L. and R.J. Seidler. <u>Legionella</u> Incidence and Density in Potable Drinking Water Supplies. Appl. Environ. Microbiol. <u>45</u>:337-339 (1983).
- 100. Stout, J.E., V.L. Yu and M.G. Best. Ecology of <u>Legionella</u> <u>pneumophila</u> Within Water Distribution Systems. Appl. Environ. Microbiol., <u>49</u>:221-228 (1985).
- 101. Colbourne, J.S. and R.M. Trew. Presence of <u>Legionella</u> in London's Water Supplies. Israel Jour. Med. Sci., <u>22</u>:633-639 (1986).
- 102. Wadowsky, R.M. and R.B. Yee. Effect of Non-Legionellaceae Bacteria on the Multiplication of Legionella pneumophila in Potable Water. Appl. Environ. Microbiol., 49:1206-1210 (1985).
- 103. Colbourne, J.S., P.J. Dennis, R.M. Trew, C. Berry and G. Veseg. Legionella and Public Water Supplies. Wat. Sci. Tech., <u>30</u>:5-10 (1988).
- 104. Witherell, L.E., R.W. Duncan, K.M. Stone, L.J. Stratton, L. Oriciari, S. Kappel and D.A. Jillson. Investigation of <u>Legionella pneumophila</u> in Drinking Water. Jour. Amer. Water Works Assoc., <u>80</u>:87-93 (1988).
- 105. Fields, B.S., E.B. Shotts, Jr., J.C. Feeley, C.W. Gorman and W.T. Martin. Proliferation of <u>Legionella pneumophila</u> as an Intracellular Parasite of the Ciliated Protozoan <u>Tetrahymena</u> pyriformis. Appl. Environ. Microbiol. 47:467-471 (1984).
- 106. Wadowsky, R.M., L.J. Butler, M.K. Cook, et al. Growth-Supporting Activity for <u>Legionella pneumophila</u> in Tap Water Cultures and Implication of Hartmannellid Amoebae as Growth Factors. Appl. Environ. Microbiol. 54:2677-2682 (1988).
- 107. Barrow, G.I. Legionnaire's Disease and Its Impact on Water Supply Management. Jour. Institu. Water Environ. Manage. <u>1</u>:117-122 (1987).
- 108. Wadowsky, R.M. and R.B. Yee. Satellite Growth of Legionella pneumophila With an Environmental Isolate of Flavobacterium breve. Appl. Environ. Microbiol. 46:1447-1449 (1983).
- 109. Stout, J.E., V.L. Yu and M.G. Best. Ecology of <u>Legionella</u> <u>pneumophila</u> Within Water Distribution Systems. Appl. Environ. Microbiol. <u>49</u>:221-228 (1985).
- 110. Wadowsky, R.M. and R.B. Yee. Effect of Non-Legionellaceae Bacteria on the Multiplication of Legionella pneumophila in Potable Water. Appl. Environ. Microbiol. 49:1206-1210 (1985).
- 111. Geldreich, E.E. Microbiological Quality Control in Distribution Systems. In: Water Quality and Treatment. Technical Editor, F.W. Pontius, Amer. Water Works Assoc., McGraw-Hill, Inc., New York, NY. (1990).
- 112. Gambassini, L., C. Sacco, E. Lanciotti, D. Burrini and O. Griffini. Microbial Quality of the Water in the Distribution System of Florence. Aqua, <u>39</u>:258-264 (1990).

Ì

840

- 113. Gavin, F. and Leclerc, H. Etude des Bacilles Gram-Pigmentes en Joune Isoles de l'Eau. Intern. Ocean. Med. 37:17-68 (1974).
- 114. Ducluzeau, R., Bochan, J.M., and Defresne, S. La Microflora autochtone de l'Eau Mineral Nature Caracteres Phyrialogiques Signification Hygienique. Med. Nutr. <u>12</u>:115-120 (1976).
- 115. Hernandez Duquino, H. and Rosenberg, F.A. Antibiotic-resistant <u>Pseudomonas</u> in Bottled Drinking Water. Can. Jour. Microbiol. <u>33</u>:286-289 (1987).
- 116. Lindberg, R.B. Culture and Identification of Commonly Encountered Gram-negative Bacilli: <u>Pseudomonas</u>, <u>Klebsiella-Enterobacter</u>, <u>Serratia</u>, <u>Proteus</u> and <u>Providencia</u>. In: <u>Opportunistic Pathogens</u>. J.E. Prier and H. Friedman eds. University Park Press, Baltimore, MD. (1974).
- 117. Holder, I.A. Epidemiology of <u>Pseudomonas aeruginosa</u> in a burns hospital. In: <u>Pseudomonas aeruginosa</u>: Ecological Aspects and Patient Colonization. Ed. V.M. Young. Raven Press, New York, NY. (1977).
- 118. Tinne, J.E., A.M. Gordon, W.H. Bain and W.A. Mackey. Crossinfection by <u>Pseudomonas aeruginosa</u> as a Hazard of Intensive Surgery. Brit. Med. Jour., 4:313-315 (1967).
- 119. Schimpff, S.C., W.H. Greene, V.M. Young and P.H. Wiernik. <u>Pseudomonas Septicemia</u>: Incidence, Epidemiology, Prevention and Therapy in Patients with Advanced Cancer. Eur. Jour. Cancer, 9:449-455 (1973).
- 120. Schimpff, S.C., R.M. Miller, S. Polkavetz and R.B. Hornik. Infection in the Severely Traumatized Patient. Ann. Surg., <u>179</u>:352-357 (1974).
- 121. Jay A. Jacobson. Pool-Associated <u>Pseudomonas</u> <u>aeruginosa</u> Dermatitis and Other Bathing-associated Infections. Infection Control, <u>6</u>:398-401 (1985).
- 122. Hunter, C.A. and P.R. Ensign. An Epidemic of Diarrhea in a Newborn Nursery Caused by <u>Pseudomonas</u> <u>aeruginosa</u>. Am. J. Pub. Health, <u>37</u>:1166-1169 (1947).

Patient Colonization. Raven Press, New York, NY. (1977).

- 124. Stoodley, B.J. and B.T. Tom. Observations on the Intestinal Carriage of <u>Pseudomonas</u> <u>aeruginosa</u>. Jour. Med. Microbiol., <u>3</u>:367-375 (1970).
- .25. Buck, A.C. and E.M. Cooke. The Fate of Ingested <u>Pseudomonas</u> <u>aeruginosa</u> in Normal Persons. Jour. Med. Microbiol. <u>2</u>:521-525 (1969).
- 126. Ringen, L.M. and C.H. Drake. A Study of the Incidence of <u>Pseudomonas aeruginosa</u> from Various Natural Sources. Jour. Bact. <u>64</u>:841-845 (1952).

- 127. Hoadley, A.W., E. McCoy, and G.A. Rohlich. Untersuchungen uber <u>Pseudomonas aeruginosa</u> in Oberflachengewassern. I. Quellen. <u>Arch. Hyg. Bakteriol.</u> <u>152</u>:328-338.
- 128. Weber, G., H.P. Werner and H. Matschnigg. <u>Pseudomonas</u> <u>aeruginosa</u> in Trinkwasser als Todesursache bei Neugeborenen. Zentralbl. Bakteriol. Parasitenk. Infektionskr. Hyg. I Abt. Orig., <u>216</u>:210-214 (1971).
- 129. Lennette, E.H., Balows, A., Hausler, Jr., W.J. and Shadomy, H.J. <u>Manual of Clinical Microbiology</u>. 4th edition, p 350-373. Amer. Soc. Microbiol., Washington, D.C. (1985).

L

ł

PAPORTNO. 2. p. PERFAGO/A-92/094 2. p. ATTREAMD SUBTICE Deportunistic Organisms and the Water Supply s. MEPORT DATE Opportunistic Organisms and the Water Supply s. MEPORT DATE s. MEPORT DATE Opportunistic Organisms and the Water Supply s. MEPORT DATE s. MEPORT DATE Opportunistic Organisms and the Water Supply s. MEPORT DATE s. MEPORT DATE Connection S. MEPORTMAKE ORGANIZATION AGONG s. MEPORT DATE Risk Reduction Engineering Laboratory-Cincinnati, OH 10. PROFINANC ORGANIZATION AND ADDRESS 11. CONTRACT.GRANT NO. Risk Reduction Engineering Laboratory-Cincinnati, OH 11. CONTRACT.GRANT NO. 11. CONTRACT.GRANT NO. U.S. Environmental Protection Agency 11. TYPE OF REPORT AND FERIOD COVERES 11. TYPE OF REPORT AND FERIOD COVERES Risk Reduction Engineering Laboratory-Cincinnati, OH 11. CONTRACT.GRANT NO. 11. CONTRACT.GRANT NO. U.S. Environmental Protection Agency 11. TYPE OF REPORT AND FERIOD COVERES 11. TYPE OF REPORT AND FERIOD COVERES Risk Reduction Engineering Laboratory-Cincinnati, OH 11. CONTRACT.GRANT NO. 11. CONTRACT.GRANT NO. U.S. Envitantary ANTES DO = Edvin E. Goldreich (513)559-72	TECHNICAL R (Please read Instructions on th		
LITTLE AND SUBTICE B. REPORT DATE Opportunistic Organisms and the Water Supply E. PERFORMUNG ORGANIZATION CODE Connection F. PERFORMUNG ORGANIZATION REPORT NO. T. AUTHORISI Edwin E. Geldreich S. FERFORMUNG ORGANIZATION NAME AND ADDRESS II. PROGRAM ELIMENT NO. Risk Reduction Engineering Laboratory-Cincinnati, OB II. CONTRACLORANT NO. U.S. Environmental Protection Agency II. VIPE OF REPORT AND FERICE COVERED Cincinnati, OB J. Store Control Continue Control (State Control (Control (State Control (Control (State Control (Control (State Control (State Control (Control (State Control (Control (Con		3.	
Connection Provide ORGANIZATION COSE T.AUTHOR(5) Edwin E. Geldreich Protection Agency Cincinnati, OR d5268 Nisk Reduction Engineering Laboratory-Cincinnati, OR Office of Research and Development U.S. Environmental Protection Agency Cincinnati, OR d5268 Nisk Reduction Engineering Laboratory-Cincinnati, OR Dffice of Research and Development U.S. Environmental Protection Agency Cincinnati, OR d5268 Nisk Reduction Engineering Laboratory-Cincinnati, OR Diffice of Research and Development U.S. Environmental Protection Agency Cincinnati, OR d5268 Nisk Reduction Engineering Laboratory-Cincinnati, OR Diffice of Research and Development U.S. Environmental Protection Agency Cincinnati, OR d5268 Nisk Reduction Engineering Laboratory-Cincinnati, OR Diffice of Research and Development U.S. Environmental Protection Agency Cincinnati, OR d5268 Nisk Reduction Engineering Laboratory-Cincinnati, OR Diffice of Research and Development U.S. Environmental Protection Agency Cincinnati, OR d5268 Nisk Reduction Engineering Laboratory-Cincinnati, OR Discription Agency Cincinnati, OR d5268 Nisk Reduction Engineering Laboratory-Cincinnati, OR Discription Agency Cincinnati, OR d5268 Discription Agency Discription Agency Discription Agency Discription Agency Discription Discription Dis		5. REPORT DATE	
Connection ANTHORIG T. AUTHORIG Edwin E. Geldreich C.FARJOANNG ORGANIZATION NAME AND ADDRESS ID. FRAGORMING ORGANIZATION NAME AND ADDRESS Risk Reduction Engineering Laboratory-Cincinnati, OH U.S. Environmental Protection Agency Cincinnati, OH 45268 U.S. Environmental Protection Agency Distribution Addew Contention Cincinnati, OH 45268 U.S. Environmental Protection Agency Distribution Addew Contention Cincinnati, OH 5266 Visuation Control and Development Distribution Addew Contention U.S. Environmental Protection Agency Distribution Addew Contention Cincinnati, OH 45268 Superimental Protection Agency EPA/600/14 Superimental Protection Agency EPA/600/14 Superimental Protection Agency EPA/600/14 Superimental Protection Agency Continuation of Protection Agency Continuition of Protection Agency EPA/600/14 Superimental Protection Agency EPA/600/14 Superimental Protection Agency EPA/600/14 Notestation Protection Agency EPA/600/14 Superimental Protection Agency EPA/600/14 Superim	Opportunistic Organisms and the Water Suppl	y S REPEORNING ORGANIZATION CODE	
Edwin E. Geldreich 10. PAGGRAM ELEMENT NO. PERFORMING ORGANIZATION NAME AND ADDRESS 10. PAGGRAM ELEMENT NO. Risk Reduction Engineering Laboratory-Cincinnati, OH 11. CONTRACTORANT NO. U.S. Environmental Protection Agency 11. CONTRACTORANT NO. Cincinnati, OH 45268 Risk Reduction Engineering Laboratory-Cincinnati, OH 11. STONGONG CADENT NAME AND ADDRESS Risk Reduction Engineering Laboratory-Cincinnati, OH 11. STONGONG CADENT NAME AND ADDRESS Risk Reduction Engineering Laboratory-Cincinnati, OH 11. STONGONG CADENT NAME AND ADDRESS Risk Reduction Engineering Laboratory-Cincinnati, OH 11. STONGONG CADENT NO. U.S. Environmental Protection Agency 11. STONGONG CADENT Cincinnati, OH 45268 12. STANGONG CADENT NO. 11. Subvisionary Notics PO = Edwin E. Geldreich (513)569-7232, Technology Conference Proceedings, Part II, Sessions EN Through STG, AWWA Water Quality Technology Conf, 11/10-14/91, Orlando, Florida, P:823-842 * Organisms that become established in water supply may also be opportunistic pathogens. Representative opportunistic pathogens that are waterborne include acid-fast bacteria, fecal klebsiella, Acid-fast bacteria, fecal klebsiella, Legionella and Pseudomonas aeruginosa. * Organisms may be found in the heterotrophic bacteria that may pose a risk to those consumers of varying health status in the community water supply allso have a resp	Connection	S. FERFORMING ONGANIZATION COSE	
PERFORMING ORGANIZATION NAME AND ADDRESS ID. PROGRAM ELEMENT NO. Fisk Reduction Engineering Laboratory-Cincinnati, OH Office of Research and Development II. CONTRACT.GRANT NO. U.S. Environmental Protection Agency Cincinnati, OH Office of Research and Development II. CONTRACT.GRANT NO. U.S. Environmental Protection Agency Cincinnati, OH 45268 II. TOPE OF REPORT AND PERIOD COVERCD Published Paper 15. SuperStructure Advances II. TOPE OF REPORT AND PERIOD COVERCD Published Paper 16. SuperStructure Advances II. TOPE OF REPORT AND PERIOD COVERCD Published Paper 17. Sourcestive Advances II. TOPE OF REPORT AND PERIOD COVERCD Published Paper 18. SuperStructure Advances II. TOPE OF REPORT AND PERIOD COVERCD Published Paper 18. SuperStructure Advances II. TOPE OF REPORT AND PERIOD COVERCD Published Paper 19. Statistic Advances II. Contract.GRANT NO. 11. Contract.GRANT NO. II. TOPE OF REPORT AND PERIOD COVERCD Published Paper 11. Sessions EM Fordeficing Cover Code 11. Contract.GRANT NO. II. TOPE OF REPORT AND PERIOD COVERCD Published Paper 11. Contract.GRANT NO. II. TOPE OF REPORT AND PERIOD COVERCD Published Paper 11. Contract.GRANT NO. II. Contract.GRANT NO. 11. Contract.GRANT NO. II. TOPE OF REPORT AND COVERCOVERCOVERCE	7. AUTHOR(S)	8. PERFORMING ORGANIZATION REPOR	IT NO.
Risk Reduction Engineering Laboratory-Cincinnati, OH 11. CONTRACLGRANTING. Office of Research and Development 11. CONTRACLGRANTING. U.S. Environmental Protection Agency 11. TYPE OF AFFORT AND FENDO COVERED Cincinnati, OH 45268 U.S. Environmental Protection Agency 11. TYPE OF AFFORT AND FENDO COVERED Published Paper 11. STONSGAUNCY NOTES U.S. Environmental Protection Agency 11. STONSGAUNCY CODE U.S. Environmental Protection Agency 11. Stons Ed Through STG. AWAW Water Quality Technology Conference Proceedings, Part II, Sessions Ed Through STG. AWAW Water Quality Technology Conf, 11/10-14/91, Orlando, Florida, P:823-842 Organisms that become established in water supply may also be opportunistic pathogens. Representative opportunistic pathogens that are waterborne include acid-fast bacteria, fecal Klebsiellae, Legionella and Pseudomonas aeruginosa. These organisms may be found in the heterotrophic bacterial population of treated drinking water and if appropriate conditions exist, may colonize and become part of the biofilm. Maintaining a high quality water supply requires careful treatment and a clean water distribution system. Users of the community water supply also have a responsibility to preserve this water quality from deterioration as it leaves the service meter and traverses the building supply lines. The goal is to minimize exposure to various heterotrophic bacteria 12. CONTENTSTATEMENT DISENTIFIERS.OPENTANALYSIS 13. DIMAISUMENE	Edwin E. Geldreich		
Office of Research and Development 11. CONTRACTORATION U.S. Environmental Protection Agency 11. CONTRACTORATION Cincinnati, OH 45268 12. TYPE OF REPORT AND PERIOD COVERED Risk Reduction Engineering Laboratory-Cincinnati, OH Published Paper 0ffice of Research and Development 12. TYPE OF REPORT AND PERIOD COVERED U.S. Environmental Protection Agency 13. SUPPLIEMENTARY NOTES Cincinnati, OH 45268 EPA/600/14 15. Supplies Variant Notes P0 = Edvin E. Geldreich (513)569-7232, Technology Conference Proceedings, Part II, Sessions Ed Through STG, AWWA Water Quality Technology Conf, 1/10-14/91, Orlando, Florida, P:823-842 * Organisms that become established in water supply may also be opportunistic pathogens. Representative opportunistic pathogens that are waterborne include acid-fast bacteria, fecal klebsiellae, Legionella and Pseudomonas aeruginosa. These organisms may be found in the heterotrophic bacterial population of treated drinking water and if appropriate conditions exist, may colonize and become part of the biofilm. Maintaining a high quality water supply requires careful treatment and a clean water distribution system. Users of the community water supply also have a responsibility to preserve this water quality from deterioration as it leaves the service meter and traverses the building supply lines. The goal is to minime exposure to various heterotrophic bacteria that may pose a risk to those consumers of varying health status in the community of people.s. 12. CONTRELIG			
U.S. Environmental Protection Agency Cincinnati, OH 45268 II. TYPE OF REPORT AND FERIOD COVERED Risk Reduction Engineering Laboratory-Cincinnati, OH Office of Research and Development U.S. Environmental Protection Agency Cincinnati, OH 45268 II. TYPE OF REPORT AND FERIOD COVERED Published Paper U.S. Environmental Protection Agency Cincinnati, OH 45268 EPA/600/14 U.S. SuperStandard Development U.S. Environmental Protection Agency Cincinnati, OH 45268 EPA/600/14 U.S. SuperStandard Development U.S. SuperStandard Development U.S. SuperStandard Development U.S. SuperStandard Development Cincinnati, OH 45268 EPA/600/14 U.S. SuperStandard Development U.S. SuperStandard Development Development Statistic Pathogens EPA/600/14 U.S. SuperStandard Development U.S. SuperStandard Development Development Statistic Pathogens EPA/600/14 U.S. SuperStandard Development U.S. SuperStandard Development Development Protection Development Developmen		INATI, OH	
Clincinnati, OH 45268 II. TYPE OF REPORT AND PERIOD COVERED Published Paper Risk Reduction Engineering Laboratory-Cincinnati, OH Office of Research and Development II. TYPE OF REPORT AND PERIOD COVERED Published Paper U.S. Environmental Protection Agency Cincinnati, OH 45268 II. TYPE OF REPORT AND PERIOD COVERED Published Paper IS.SUMPLEMENTARY NOTES Proceedings, Part II. Sessions EB Through STG, AWWA Water Quality Technology Conference Proceedings, Part II. Sessions EB Through STG, AWWA Water Quality Technology Conf, 11/10-14/91, Orlando, Florida, P:823-842 IS.SUMPLEMENTARY NOTES Proceedings, Part II. Sessions EB Through STG, AWWA Water Quality Technology Conf, 11/10-14/91, Orlando, Florida, P:823-842 IS.SUMPLEMENTARY NOTES Pathogens. Representative opportunistic pathogens that are waterborne include acid-fast bacteria, fecal klebsiellae, Legionella and Pseudomonas aeruginosa. These organisms may be found in the heterotrophic bacterial population of treated drinking water and if appropriate conditions exist, may colonize and become part of the biofilm. Maintaining a high quality water supply requires careful treatment and a clean water distribution system. Users of the community water supply also have a responsibility to preserve this water quality from deterioration as it leaves the service meter and traverses the building supply lines. The goal is to minimize exposure to various heterotrophic bacteria that may pose a risk to those consumers of varying health status in the community of people. II. KEV WCABS AND DECUMENT ANALYSIS II. DESCRIPTORS DESCRIPTORS DIDENTIFIESJOPEN ENDED TEAMS & COS			
Risk Reduction Engineering Laboratory-Cincinnati, OH Office of Research and Development Published Paper U.S. Environmental Protection Agency Cincinnati, OH 45268 Feadbook EPA/600/14 U.S. Environmental Protection Agency Cincinnati, OH 45268 EPA/600/14 U.S. Environmental Protection Agency Cincinnati, Protection Agency Cincinnation Protection Agency Cincinnation Age	Cincinnati, OH 45268		
Office of Research and Development [1:5PONSCRING AGENCY CODE U.S. Environmental Protection Agency EPA/600/14 SisSuprisukersany NOTES PO = Edwin E. Geldreich (513)569-7232, Technology Conference Proceedings, Part II, Sessions EB Through STG, AWWA Water Quality Technology Conf. 1/10-14/91, Orlando, Florida, P:823-842 To address that become established in water supply may also be opportunistic pathogens. Representative opportunistic pathogens that are waterborne include acid-fast bacteria, fecal klebsiellae, Legionella and Pseudomona geruginosa. These organisms may be found in the heterotrophic bacterial population of treated drinking water and if appropriate conditions exist, may colonize and become part of the biofilm. Maintaining a high quality water supply requires careful treatment and a clean water distribution system. Users of the community water supply also have a responsibility to preserve this water quality from deterioration as it leaves the service meter and traverses the building supply lines. The goal is to minimize exposure to various heterotrophic bacteria that may pose a risk to those consumers of varying health status in the community of people. 17. KEY WERES AND OCCUMENT AMALYSIS 17. KEY WERES AND OCCUMENT AMALYSIS </td <td></td> <td></td> <td>ERED</td>			ERED
U.S. Environmental Protection Agency Cincinnati, OH 45268 EPA/600/14 SUPPENENTARY NOTES PD = Edwin E. Geldreich (513)569-7232, Technology Conference Proceedings, Part II, Sessions EB Through STG, AWMA Water Quality Technology Conf, 11/10-14/91, Orlando, Florida, P:823-842 No regardings, Part II, Sessions EB Through STG, AWMA Water Quality Technology Conf, 11/10-14/91, Orlando, Florida, P:823-842 No regardings, Representative opportunistic pathogens that are waterborne include acid-fast bacteria, fecal klebsiellae, Legionella and Pseudomonas aeruginosa. These organisms may be found in the heterotrophic bacterial population of treated drinking water and if appropriate conditions exist, may colonize and become part of the biofilm. Maintaining a high quality water supply requires careful treatment and a clean water distribution system. Users of the community water supply also have a responsibility to preserve this water quality from deterioration as it leaves the service meter and traverses the building supply lines. The goal is to minimize exposure to various heterotrophic bacteria that may pose a risk to those consumers of varying health status in the community of people. 17. KIEV WCRES AND DOCUMENT ANALYSIS 18. DISECRIPTORS 19. DESCRIPTORS 10. Mater supply quality Fecal Klebsiella Acid-fast bacteria Legionella 19. SECUMITY CLASS (Ibm Majour, UNCLASSTIFIED 21. NO. OF PAGES 22. PRICE		INATI, UH FUDIISHED FAPET	<u></u>
Cincinati, ON 45268 15 SUPPENDENTARY NOTES 15 SUPPENDENTARY NOTES 16 Additional Action 17 Organisms that become established in water supply may also be opportunistic pathogens. Representative opportunistic pathogens that are waterborne include acid-fast bacteria, fecal klebsiellae, Legionella and Pseudomonas aeruginosa. These organisms may be found in the heterotrophic bacterial population of treated drinking water and if appropriate conditions exist, may colonize and become part of the biofilm. Maintaining a high quality water supply requires careful treatment and a clean water distribution system. Users of the community water supply also have a responsibility to preserve this water quality from deterioration as it leaves the service meter and traverses the building supply lines. The goal is to minimize exposure to various beterotrophic bacteria that may pose a risk to those consumers of varying health status in the community of people. 17. KEY WCRES AND DECLIMENT ANALYSIS 18. DESCRIPTIONS 19. DESCRIPTIONS 10. KEY WCRES AND DECLIMENT ANALYSIS 11. COSATIFIEM CLASS (Phur Report, 21.NO. OF \$ACES 12. DESCRIPTIONS 13. DESCRIPTIONS 14. SCHENTICK STATEMENT- 15. DESCRIPTIONS 16. DESCRIPTIONS 17. COSATIFIEM CLASS (Phur Report, 21.NO. OF \$ACES 18. <t< td=""><td></td><td>EBA /600/1/</td><td></td></t<>		EBA /600/1/	
Proceedings, Part 11, Sessions EB Through STG, AWA Water Quality Technology Conference Proceedings, Part 11, Sessions EB Through STG, AWA Water Quality Technology Conf, 11/10-14/91, Orlando, Florida, P:823-842 * Organisms that become established in water supply may also be opportunistic pathogens. Representative opportunistic pathogens that are waterborne include acid-fast bacteria, fecal klebsiellae, Legionella and Pseudomonas aeruginosa. These organisms may be found in the heterotrophic bacterial population of treated drinking water and if appropriate conditions exist, may colonize and become part of the biofilm. Maintaining a high quality water supply requires careful treatment and a clean water distribution system. Users of the community water supply also have a responsibility to preserve this water quality from deterioration as it leaves the service meter and traverses the building supply lines. The goal is to minimize exposure to various heterotrophic bacteria that may pose a risk to those consumers of varying health status in the community of people 			

EPA Form 2220-1 (Rov. 4-77) PREVIOUS EDITION IS OBSOLETE

•

.

•.

-